

# Molecular Docking of Mangrove Plantas Therapeutic Agentto Treat Non-Small Cell Lung Carcinoma

## ABSTRACT

**Background:** Cancer is one of the biggest health problems worldwide, with lung cancer as the first rank in the number of new cases and deaths. Non-small cell lung carcinoma (NSCLC) is a type of lung cancer that accounts for about 85% of all lung cancer cases. Previous research identified the role of epidermal growth factor receptor (EGFR) as the most suitable target to treat NSCLC. This study aims to identify the potential compounds derived from mangrove plants as agents to treat NSCLC using a molecular docking study.

**Methodology:** Six natural compounds, which include taraxasterol, stigmasterol, tretinoin, heritonin, ascochitine, and tricin, along with gefitinib as a drug comparative were used. Docking was carried out on EGFR as a receptor target by Autodock Tools. The visualizations of molecular interactions were carried out by BIOVIA Discovery Studio 2020.

**Results:** The results showed that all six compounds were compiled from several criteria as drugs based on Lipinski analysis and had an affinity to EGFR receptors. The docking results were found in the order of stigmasterol (-11.84 kcal/mol), taraxasterol (10.80 kcal/mol), tretinoin (-10.60 kcal/mol), tricin (-9.24 kcal/mol), ascochitine (-7.85 kcal/mol), heritonin (-7.81 kcal/mol), and gefitinib (-8.62 kcal/mol). Among these natural compounds, stigmasterol exhibited the highest binding affinity. ADME profile showed that these natural compounds are safe and drug-like compounds.

**Conclusion:** Stigmasterol exhibited the highest binding energy of -11.84 kcal/mol. All three compounds bind in the binding pocket of EGFR. All compounds have drug-likeness properties based on Lipinski rules

**Keywords:** Epidermal growth factor; molecular docking; non-small cell lung carcinoma; taraxasterol; stigmasterol.

## 1. INTRODUCTION

Cancer is one of the biggest health problems that lead to the leading causes of death worldwide[1]. This disease also shows an increasing trend in recent years and is predicted to increase every year[2]. Lung cancer is the leading cause of cancer death among both men and women in the United States so far. In 2022, American Cancer Society estimated 236,740 new cases of lung cancer will be diagnosed and 130,180 will die, approximately 350 deaths per day caused by lung cancer, the leading cause of death[3].

Lung cancer has been linked to several factors including smoking, genetic predisposition, and environmental factors[4]. Lung cancer can be classified into two types which are small

cell lung cancer (SCLC) and non-small cell lung carcinoma (NSCLC), with NSCLC responsible for approximately 85% of lung cancer cases[5]. Lung cancer may exist because of genetic and epigenetic changes of the cellular genome, which is critical to the disease progression. The comprehensive molecular dissection of NSCLC found the mutation in epidermal growth factor receptor (EGFR) genes for about 10-40% cases with 14-19% of western patients and 40-48% of Asian patients. EGFR is a kind of tyrosine kinase receptor located at the cell surface. EGFR can generate differentiation and proliferation of cells upon activation through the binding of one of its ligands. Based on the fact that EGFR mutation leads to NSCLC, research has shown that targeting EGFR is currently considered the most suitable way to treat it[6,7].

The current treatments which are usually used to treat NSCLC are surgery, chemotherapy, and targeted therapy[8]. NSCLC patients whose tumors activate kinase domain mutations in EGFR often respond to EGFR tyrosine kinase inhibitors (TKI) such as erlotinib, gefitinib, and afatinib[9]. However, some studies have found TKI drug resistance in some NSCLC patients with EGFR mutation[10]. NSCLC treatment with surgery is invasive and limited to stage I-II and IIIA[11]. Moreover, chemotherapy treatment which is often used in several types of cancer, had serious side effects[12]. Therefore, the development of novel and treatment for treating NSCLC patients is needed. In the last few decades, research on herbs as an alternative treatment with minimal side effects has been developed. Natural compounds are widely used in various therapeutic interventions due to their benefits as anticancer and minimum side effects[13,14].

Mangrove plant has abundant bioactive compounds and can serve as a reservoir for novel bioactive compounds such as amides, alkaloids, tannins, flavonoids, saponin, glycosides, terpenoid, phenolic, and phytosterol [16]. The pharmacological activities of terpenoid, phenolic, and phytosterol indicate that they have a potential as preventative supplements and pharmaceutical agents as anticancer, antifungal, antibacterial, antiviral, antioxidant, anti-inflammatory, and other activities. Some specific compounds pose prominent effects as anti-cancer. Terpenoid exhibits cytotoxic against many cancer cell lines[17,18]. Taraxasterol is one of terpenoid known as anticancer for a few types of cancer. Chen et al. revealed that taraxasterol cut the growth of gastric cancer by inhibiting of EGFR signaling[19]. The main phytosterol in mangrove, stigmasterol was also found to inhibit proliferation and promoted the apoptosis of lung cancer cells [20]. As a whole, Song et al. reported that mangrove compounds can be a multi-target inhibitors such as inhibit activities of HER2, HER3, HER4, RET, and EGFR in treating NSCLC[21]. The specific compounds that play a role in inhibiting EGFR remain unclear. Thus, the present study aims to identify the potential compounds derived from mangrove plants as therapeutic agents to treat NSCLC.

## 2. MATERIALS AND METHODS

This study was conducted utilizing molecular docking computational method. The materials used in this study were protein target, namely EGFR (PDB ID: 3G5Z) downloaded from <http://www.rcsb.org> and mangrove compounds were downloaded from <https://pubchem.ncbi.nlm.nih.gov>. The drug used as comparative was Gefitinib (Compound CID: 123631) which downloaded from <https://pubchem.ncbi.nlm.nih.gov>.

### 2.1 Selection of Mangrove Compounds

The mangrove compounds were retrieved from previous studies. Six mangrove compounds were used in this study including taraxasterol (Compound CID:115250), stigmasterol

79 (Compound CID: 5280794), tretinoin (Compound CID: 444795), heritonin (Compound CID:  
80 130118), ascochitine (Compound CID: 73486), and tricin (Compound CID: 5281702).

## 81 **2.2 Geometry Optimization**

82

83 After the compounds from Pubchem were downloaded, the compounds were saved in pdb  
84 format using Discovery studio. Then, geometry optimization was carried out in Argus Lab  
85 4.0.1. software using PM3 semi-empirical parameterization based on Hartree-Fock  
86 calculation method. Argus lab software computed the energy convergence (stopping point of  
87 the compound's molecule[22]. Furthermore, the compound's format was converted to pdb  
88 with OpenBabel software to make it readable with Autodock Tools program[23].

## 89 **2.3 Preparation of Target Protein and Compounds**

90

91 The target protein was used in this study was EGFR (PDB ID: 5UG8). The preparation of the  
92 target protein was performed by removing water molecules (H<sub>2</sub>O) contained in the target  
93 protein, adding polar hydrogen atoms, cleaning the target protein structure from natural  
94 ligands then saved its file in the pdbqt format[24]. The preparation of the  
95 compounds were carried out by changing sdf format to pdbqt format using Discovery Studio  
96 and AutoDock software.

## 97 **2.4 Validation**

98

99 Validation of the molecular docking method was done by redocking the native ligand (N-  
100 [(3R,4R)-4-fluoro-1-{6-[(1-methyl-1H-pyrazol-4-yl)amino]-9-(propan-2-yl)-9H-purin-  
101 2yl}pyrrolidin-3yl]propanamide) to the selected macromolecule (EGFR) using Autodock  
102 Tools software. The binding site and the parameters used in this study are considered valid  
103 the RMSD value is  $\leq 2\text{\AA}$ [25].

## 104 **2.5 Docking Protocol**

105

106 The molecular docking was carried out to predict the binding energies of the compounds  
107 toward target protein using the Autodock Tools, Autogrid4, and Autodock4  
108 software[26,27]. The docking simulation was done by arranging the docking parameters,  
109 which are the grid box size (x = 40, y = 40, z = 40), the grid box coordinate (x = -13.156, y =  
110 14.7, z = -25.718), 0.375Å spacing, 100 runs, medium number of evals, and Lamarckian  
111 Genetic Algorithm 4.2 The docking output is indlg format. The lowest binding affinity was  
112 selected from a set of 100 conformation poses. The interactions which exhibit the strong  
113 binding energy were analyzed using Discovery Studio software.

## 114 **2.7 Drug-likeness and Toxicity Analysis**

115

116 The Lipinski rule of five was used in this study to assess the drug-like properties of  
117 compounds[28]. The molecular weight, number of hydrogen donor and acceptor, solubility,  
118 permeability, level of GI absorption, and number of Lipinski violations were screened  
119 by employing the Swiss ADME web tools <http://www.swissadme.ch/index.php>[29]. AdmetSAR  
120 2.0 online tool (<http://lmmd.ecust.edu.cn/admetSAR2>) was used to predict the toxicological  
121 profile of selected compounds[30]. The finalized ligands' SMILES were submitted in the  
122 admetSAR website to check for toxicity[31].

123 **3. RESULTS AND DISCUSSION**

124

125 **3.1 Molecular Docking Study**

126

127 All compounds were docked to analyze their binding energy using Autodock Vina. Analysis  
128 of the molecular docking results were carried out by assessing the binding energy ( $\Delta G$ ).The  
129 validation was performed by redocking the native ligand to EGFR, using the determined  
130 parameters, showed the RMSD value of 1.91 Å.Since the value is less than 2 Å, the docking  
131 method can be used to dock the test compounds. The difference between the native ligand  
132 before and after the redocking procedure (Fig. 1). The docking results are represented in  
133 Table 1. Stigmasterol, taraxasterol, and tretinoin were found to have the highest binding  
134 energy of -11.84, -10.80, and -10.60 kcal/mol,respectively. The comparative drug, gefitinib  
135 was found to have binding energy of -8.62 kcal/mol.Hence, the ability of the compounds to  
136 bind EGFR was more exceptional. They showed that more robust and stable interactions  
137 that occur between the compounds and EGFR [32]In addition, the binding energy value is  
138 directly linear with the constant inhibition value ( $K_i$ ). So, the value of binding energy can be  
139 used to estimate the ability of a compound to inhibit protein target [33]. Based on the results,  
140 mangrove compounds have the ability to inhibit EGFR as the most suitable target to treat  
141 NSCLC. The top three compounds and gefitinib were selected to visualize their interaction.

142 **Table 1. Molecular docking results**

143

144

145

146

147

148

149 **Fig. 1.**  
150 **of**  
151 **docking,**  
152 **docking**  
153 **Green:**  
154 **ligand.**  
155 **Å**

1. \_\_\_\_\_



\_\_\_\_\_ergy  
\_\_\_\_\_l)

**Validation**  
**molecular**  
**Blue: re-**  
**results;**  
**native**  
**RMSD: 1.91**

156

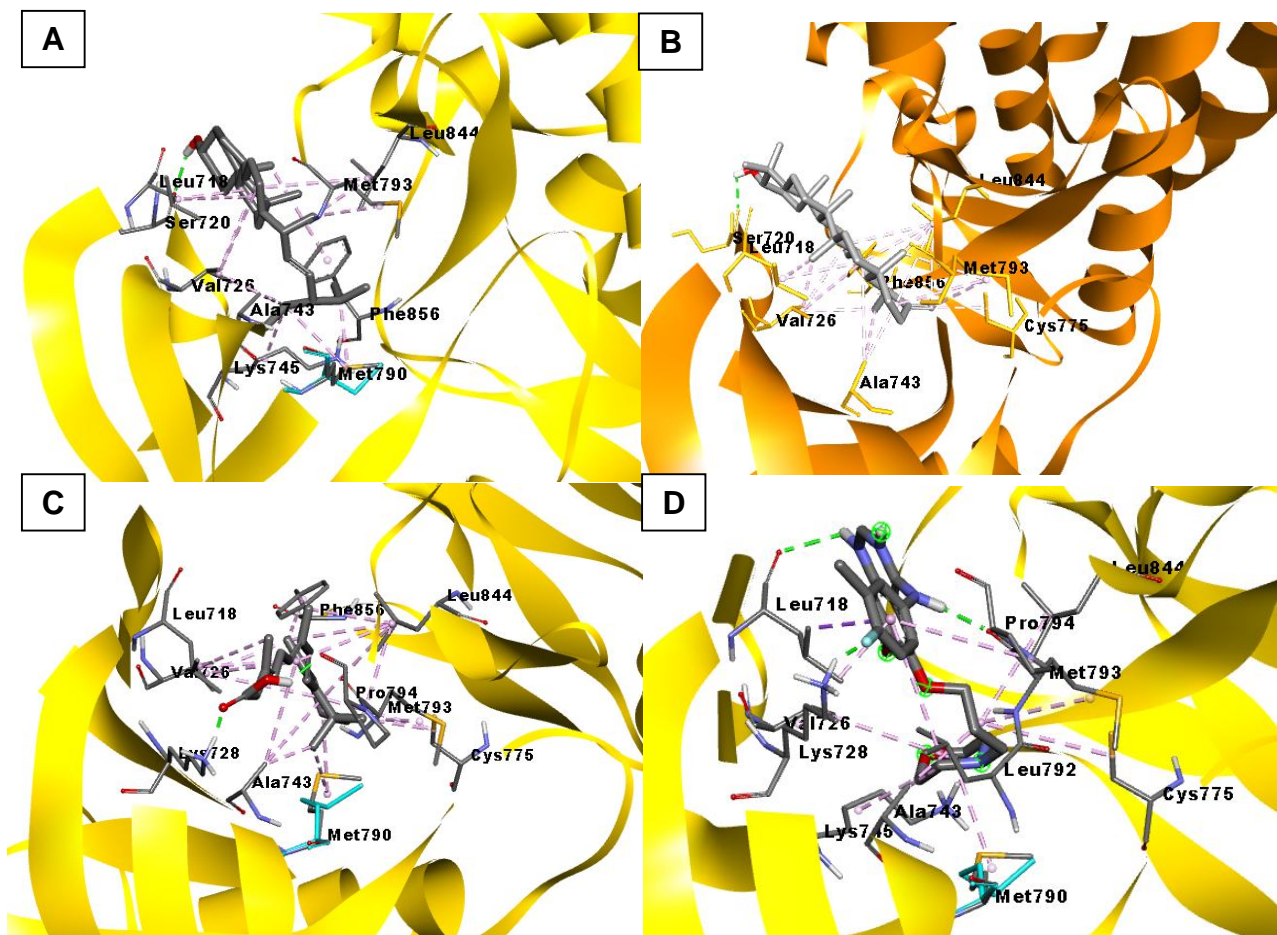
157 **3.2 Visualization of molecular docking study**

158 The visualization showed in 3D (Fig. 2) and 2D (Fig. 3) form resulted from many amino acid  
159 residues of EGFR that bind with the compounds and described in Table 2. Four amino acid  
160 residues, Leu718, Leu844, Ala743, and Val726 were found in all interactions between the  
161 compounds and EGFR.Previous molecular docking study by Ibrahim et al. revealed that the

162 active binding sites of EGFR were amino acid residues of Met793, Thr854, Leu718, Leu844,  
163 Met766, Val726, Ala743, Lys745, and Met790 [34]. Accordingly, those five amino acid  
164 residues were located in the active sites of EGFR. So, all compounds bind in the active site  
165 of EGFR. The active site or binding pocket is the binding area of enzyme that involve amino  
166 acid residues that play a role in the binding. The interaction of amino acid residues at the  
167 active site with the compounds causes compounds to have the ability to inhibit EGFR as a  
168 competitive inhibitor. There is a correlation between binding energy and the active sites  
169 (binding pocket) of protein target [35].

170 Stigmasterol and taraxasterol had hydrogen bond with residues of Ser720. Meanwhile,  
171 hydrogen bond also found in tretinoin and gefitinib with residues of Lys728. The remaining  
172 residues are hydrophobic interactions (Fig. 3). Hydrogen bond and hydrophobic interaction  
173 affect the binding energy value. The hydrogen bond is the interaction of hydrogen atoms with  
174 electronegative atoms such as fluorine (F), nitrogen (N), and oxygen (O), while hydrophobic  
175 interaction is an interaction that occurs between nonpolar molecules which include alkyl-  
176 alkyl, pi-alkyl, pi-pi stacked, and pi-pi T-shaped interactions [36,37].

177 A previous study stated that hydrogen bond and hydrophobic interaction could stabilize the  
178 compound when bind in the target protein and change the  $\Delta G$  value as well as enhance the  
179 efficacy of the compound when interacting with the target protein [38]. Similarly, a study  
180 revealed that hydrophobic interactions and hydrogen bonds both also make large  
181 contributions to compound stability [39]. Thus, hydrogen bond and hydrophobic interaction  
182 have a key role in strengthening molecular bond or enhancing binding energy, although it is  
183 still debatable between both of them regarding which type has more potential role in  
184 increasing the binding energy.

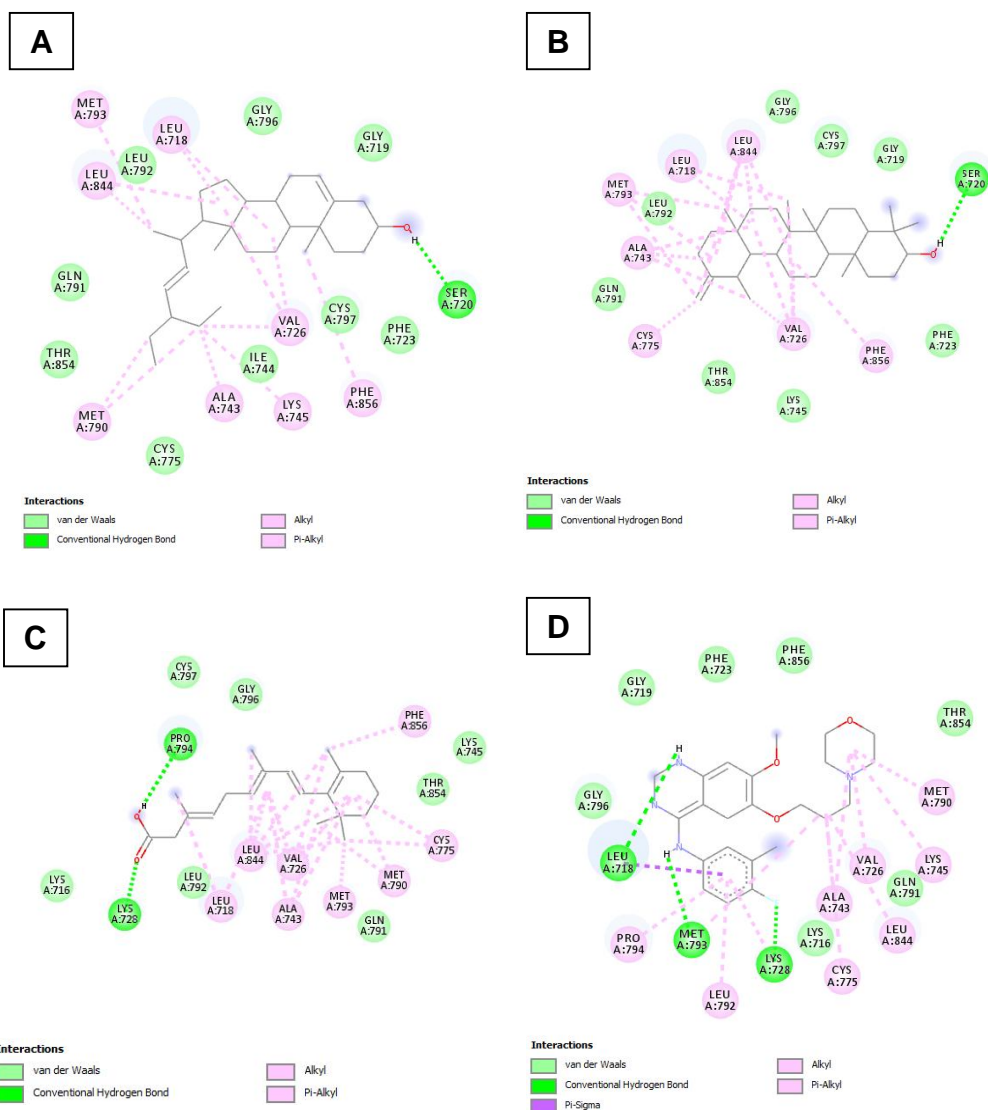


**Fig.2. 3D visualization of molecular docking results between EGFR and the compound**  
**A) stigmasterol;B) taraxasterol; C) tretinoin; D) gefitinib.**

**Table 2. The summary of visualization results**

Compounds	Amino Acid Residues	Molecular Interaction	
		Hydrophobic Interaction	Hydrogen bond
Taraxasterol	Ser720, Leu718(2), Val726(4), Ala743(3), Met793(3), Leu844(5), Cys775, Phe856	19	1
Stigmasterol	Ser720, Leu718(2), Val726(3), Ala743, Leu844(2), Met793, Met790(2), Lys745, Phe856, Lys728, Pro794, Leu718(2), Val726(3), Ala743(3), Cys775(2), Met790(2), Leu844(5), Met793, Phe856	13	1
Tretinoin	Lys728(2), Leu718(2), Met793(2), Val726, Ala743(2), Lys745, Cys775, Met790, Leu844, Leu792, Pro794	19	2
Gefitinib		12	3





**Fig. 3. 2D visualization of molecular docking results between EGFR and the compound A) stigmasterol;B) taraxasterol; C) tretinoin; D) gefitinib.**

### 3.3 Drug-likeness and Toxicity Analysis

All compounds were found to have no more than one violation (Table 3). Lipinski rules state that for any compound to be considered as a drug-like compound, a compound must obey these criteria: Molecular Weight (MW) <500 Dalton, number of H-bond acceptors <10, number of H-bond donors <5, Log P <5, and if more than one violation were found, then the compound cannot be considered as a drug-like compound [29]. All compounds in this study followed the Lipinski criteria.

On toxicity analysis, most of the compounds are a class of III on acute oral toxicity, which means that based on US EPA classification, the LD 50 values are between 500 mg/kg and

5000 mg/kg [40], while stigmasterol belongs to class I with LD 50 values of less than 50 mg/kg (Table 4). These findings give fundamental data in regards to the toxicological profile of the compounds and might be helpful in choosing the preferred dosage and the route of administration.

Compound	MW <500 (g/mol)	H-donor	H-acceptor	LogP	LogS	GI absorption	Violation
Taraxasterol	426.72	1	1	4.65	-8.24	Low	1
Stigmasterol	412.69	1	1	5.08	-7.46	Low	1
Tretinoin	300.44	1	2	4.28	-5.34	High	1
Heritonin	258.31	0	3	2.81	-3.41	High	0
Tricin	330.29	3	7	-0.07	-4.12	High	0
Ascochitine	276.28	2	5	1.84	-3.06	High	0

**Table 3. Lipinski results**

Compound	Carcinogenicity	Eye corrosion	Eye irritation	Ames mutagenesis	Hepato-toxicity	Acute oral toxicity
Taraxasterol	- (0.9571)	- (0.9834)	- (0.8878)	- (0.8400)	- (0.6250)	III (0.8879)
Stigmasterol	- (0.8571)	- (0.9886)	- (0.9673)	- (0.8300)	- (0.7750)	I (0.4287)
Tricin	- (1.0000)	- (0.9779)	+ (0.8092)	- (0.6800)	+ (0.7750)	III (0.5920)
Heritonin	- (0.9857)	- (0.9799)	- (0.6951)	+ (0.5200)	+ (0.6250)	III (0.4823)
Ascochitine	- (0.8714)	- (0.9852)	- (0.9051)	- (0.8700)	+ (0.6750)	III (0.4914)
Tretinoin	- (0.6714)	- (0.9886)	- (0.9569)	- (0.7800)	- (0.6000)	III

**Table 4. Toxicity prediction for taraxasterol, stigmasterol, triclin, heritonin, ascochitine, tretinoin, and gefitinib.**



	Gefitinib	- (0.9857)	- (0.9886)	- (0.9737)	- (0.5400)	+ (0.6750)	(0.8050) III (0.7006)
--	-----------	------------	------------	------------	------------	------------	-----------------------------

211 *\*“+” means toxic; “-“ means nontoxic. The numbers in brackets indicate the toxicity prediction*

212 Previous studies reported that taraxasterol and stigmasterol have anti-cancer activities. Bao,  
 213 et al revealed that taraxasterol inhibited liver cancer cells' growth by inducing cell cycle  
 214 arrest at G0/G1 phase and apoptosis in vitro and in vivo [41]. In gastric cancer, poor  
 215 prognosis is associated with overexpression of EGFR. Recent study showed that  
 216 taraxasterol might play a role as anti-gastric cancer by inactivation of EGFR/AKT1 signaling  
 217 pathway. It is shown that taraxasterol significantly downregulated EGFR, p-EGFR, AKT1,  
 218 and p-AKT1 level in the tumor tissues [42]. Another compound, stigmasterol prevents the  
 219 development of cholangiocarcinoma by downregulating TNF-alpha and VEGFR-2 and  
 220 suppresses skin cancer by increasing lipid peroxide levels and inducing DNA damage [52].  
 221 In the meantime, tricin had proven as anti-cancer. Naoko Seki, et al reported that tricin  
 222 inhibited proliferation of HSC (Hepatic Stellate Cells) in vitro[43,44,45].

223  
 224

## 225 4. CONCLUSION

226  
 227 Six compounds have the potential as a drug candidate. Stigmasterol exhibited the highest  
 228 binding energy. All three compounds bind in the binding pocket of EGFR. All compounds  
 229 have drug-likeness properties based on Lipinski rules. Moreover, further in vivo and in vitro  
 230 investigation are needed to bring these compounds at the clinical setting.

## 231 ACKNOWLEDGEMENTS

232  
 233 The authors would like to thank the Department of Pharmacology, Faculty of Medicine,  
 234 Universitas Sriwijaya for research support.

## 236 COMPETING INTERESTS

237  
 238 Authors have declared that no competing interests exist  
 239

## 240 AUTHORS' CONTRIBUTIONS

241  
 242 Author HAH, PMA, and T formulated, conceptualized, and designed the research. Author  
 243 MDR and DA did the program using software, organized the data, and analyzed the result.  
 244 Author HAH, DA, and RSD wrote the final drafts of the paper, reviewed, and edited the  
 245 language of the draft. All authors accepted the final draft and are responsible for the  
 246 manuscript's content and similarity index.

## 247 REFERENCES

- 248  
 249 1. World Health Organization. WHO Report On Cancer. World Health Organization. 2020.  
 250 399–438 p.  
 251 2. Kreatsoulas C, Anand SS, Subramanian S V. An emerging double burden of disease: The  
 252 prevalence of individuals with cardiovascular disease and cancer. J Intern Med.  
 253 2014;275(5):494–505.  
 254 3. American Cancer Society. Cancer Facts and Figures 2022. Atlanta; American Cancer  
 255 Society: 2022.

- 256 4. American Cancer Society. Lung Cancer Research Highlights. 2022. Available on:  
257 [https://www.cancer.org/research/acs-research-highlights/lung-cancer-research-](https://www.cancer.org/research/acs-research-highlights/lung-cancer-research-highlights.html)  
258 [highlights.html](https://www.cancer.org/research/acs-research-highlights/lung-cancer-research-highlights.html) (Accessed July 18th, 2022)
- 259 5. Zappa C, Mousa SA. Non-small cell lung cancer: Current treatment and future advances.  
260 *Transl Lung Cancer Res.* 2016;5(3):288–300.
- 261 6. Shaik NA, Al-Kreathy HM, Ajabnoor GM, Verma PK, Banaganapalli B. Molecular  
262 designing, virtual screening and docking study of novel curcumin analogue as mutation  
263 (S769L and K846R) selective inhibitor for EGFR. *Saudi J Biol Sci* [Internet]. 2019;26(3):439–  
264 48. Available from: <https://doi.org/10.1016/j.sjbs.2018.05.026>
- 265 7. Han B, Tjulandin S, Hagiwara K, Normanno N, Wulandari L, Laktionov K, et al. EGFR  
266 mutation prevalence in Asia-Pacific and Russian patients with advanced NSCLC of  
267 adenocarcinoma and non-adenocarcinoma histology: The IGNITE study. *Lung Cancer.*  
268 2017;113(June):37–44.
- 269 8. Zarogoulidis K, Zarogoulidis P, Darwiche K, Boutsikou E, Machairiotis N, Tsakiridis K, et  
270 al. Treatment of non-small cell lung cancer (NSCLC). *J Thorac Dis.* 2013;5(SUPPL.4):2–9.
- 271 9. Rosell R, Carcereny E, Gervais R, Vergnenegre A, Massuti B, Felip E, et al. Erlotinib  
272 versus standard chemotherapy as first-line treatment for European patients with advanced  
273 EGFR mutation-positive non-small-cell lung cancer (EURTAC): A multicentre, open-label,  
274 randomised phase 3 trial. *Lancet Oncol* [Internet]. 2012;13(3):239–46. Available from:  
275 [http://dx.doi.org/10.1016/S1470-2045\(11\)70393-X](http://dx.doi.org/10.1016/S1470-2045(11)70393-X)
- 276 10. Zhang Z, Lee JC, Lin L, Olivas V, Au V, Laframboise T, et al. Activation of the AXL  
277 kinase causes resistance to EGFR-targeted therapy in lung cancer. *Nat Genet.*  
278 2012;44(8):852–60.
- 279 11. Lackey A, Donington J. Surgical management of lung cancer. *Semin Intervent Radiol.*  
280 2013;30(2):133–40.
- 281 12. Aslam MS, Naveed S, Ahmed A, Abbas Z, Gull I, Athar MA. Side Effects of  
282 Chemotherapy in Cancer Patients and Evaluation of Patients Opinion about Starvation  
283 Based Differential Chemotherapy. *J Cancer Ther.* 2014;05(08):817–22.
- 284 13. Zhao GF, Huang ZA, Du XK, Yang ML, Huang DD, Zhang S. Molecular docking studies  
285 of Traditional Chinese Medicinal compounds against known protein targets to treat non-small  
286 cell lung carcinomas. *Mol Med Rep.* 2016;14(2):1132–8.
- 287 14. Changxing L, Galani S, Hassan F ul, Rashid Z, Naveed M, Fang D, et al.  
288 Biotechnological approaches to the production of plant-derived promising anticancer agents:  
289 An update and overview. *Biomed Pharmacother* [Internet]. 2020;132(November):110918.  
290 Available from: <https://doi.org/10.1016/j.biopha.2020.110918>
- 291 15. Lahjie AM, Nouval B, Lahjie AA, Ruslim Y, Kristiningrum R. Economic valuation from  
292 direct use of mangrove forest restoration in Balikpapan Bay, East Kalimantan, Indonesia.  
293 *F1000Research.* 2019;8:1–13.
- 294 16. Das G, Gouda S, Mohanta YK, Patra JK. Mangrove plants: A potential source for  
295 anticancer drugs. *Indian J Geo-Marine Sci.* 2015;44(5):666–72.
- 296 17. Simlai A, Roy A. Biological activities and chemical constituents of some mangrove  
297 species from Sundarban estuary: An overview. *Pharmacogn Rev.* 2013;7(14):170–8.
- 298 18. Patra JK, Mohanta YK. Antimicrobial compounds from mangrove plants: A  
299 pharmaceutical prospective. *Chin J Integr Med.* 2014;20(4):311–20.
- 300 19. Chen W, Li J, Li C, Fan HN, Zhang J, Zhu JS. Network pharmacology-based  
301 identification of the antitumor effects of taraxasterol in gastric cancer. *Int J*  
302 *Immunopathol Pharmacol.* 2020 Jan-Dec;34:2058738420933107. doi:  
303 10.1177/2058738420933107.
- 304 20. Dong Y, Chen C, Chen C, et al. Stigmasterol inhibits the progression of lung cancer by  
305 regulating retinoic acid-related orphan receptor C. *Histol Histopathol.* 2021;36(12):1285-  
306 1299. doi:10.14670/HH-18-388

21. Song X, Qi X, Wang Q, Zhu W, Li J. A novel multi-target inhibitor harboring selectivity of inhibiting EGFR T790M sparing wild-type EGFR. *Am J Cancer Res.* 2017;7(9):1884-1898. PMID: 28979811; PMCID: PMC5622223.
22. Bitencourt-Ferreira G, de Azevedo WF Jr. Molecular Docking Simulations with ArgusLab. *Methods Mol Biol.* 2019;2053:203-220. doi:10.1007/978-1-4939-9752-7\_13
23. Morris GM, Huey R, Lindstrom W, Sanner MF, Belew RK, Goodsell DS, et al. AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. *Journal of computational chemistry.* 2009;30(16):2785–91.
24. Madhavi Sastry G, Adzhigirey M, Day T, Annabhimoju R, Sherman W. Protein and ligand preparation: Parameters, protocols, and influence on virtual screening enrichments. *J Comput Aided Mol Des.* 2013;27(3):221–34.
25. Prasatiawati R, Suherman M, Permana B, Rahmawati R. Molecular Docking Study of Anthocyanidin Compounds Against Epidermal Growth Factor Receptor (EGFR) as Anti-Lung Cancer. *Indones J Pharm Sci Technol.* 2021;8(1):8.
26. Prieto-Martínez FD, Arciniaga M, Medina-Franco JL. Molecular docking: current advances and challenges. *TIP Rev Espec en Ciencias Químico-Biológicas.* 2018;21:65–87.
27. Trott, O., Olson AJ. AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization and multithreading. *J Comput Chem.* 2019;31(2):455–61.
28. Benet LZ, Hosey CM, Ursu O, Oprea TI. BDDCS, the Rule of 5 and Drugability. *Physiol Behav.* 2017;176(12):139–48.
29. Lipinski CA, Lombardo F, Dominy BW, Feeney PJ. Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Adv Drug Deliv Rev.* 2012;64(SUPPL.):4–17.
30. Hongbin Yang, Chaofeng Lou, Lixia Sun, Jie Li, Yingchun Cai, Zhuang Wang, Weihua Li, Guixia Liu, Yun Tang, admetSAR 2.0: web-service for prediction and optimization of chemical ADMET properties. *Bioinformatics.* 2019;35(6):1067–1069.
31. MOON A, KHAN D, GAJBHIYE P, JARIYA M. In silico prediction of toxicity of ligands utilizing admetSAR. *Int J Pharma Bio Sci.* 2017;8(3):674–7.
32. Du X, Li Y, Xia YL, Ai SM, Liang J, Sang P, et al. Insights into protein–ligand interactions: Mechanisms, models, and methods. *Int J Mol Sci.* 2016;17(2):1–34.
33. Tian G, Haffner CD. Linear Relationships between the Ligand Binding Energy and the Activation Energy of Time-dependent Inhibition of Steroid 5 $\alpha$ -Reductase by  $\Delta$ 1-4-Azasteroids. *J Biol Chem.* 2001;276(24):21359–64.
34. Ibrahim MT, Uzairu A, Shallangwa GA, Uba S. Computer-aided design of some quinazoline analogues as epidermal growth factor receptor inhibitors. *Egypt J Med Hum Genet.* 2021;22(1).
35. Kumar SP, Patel CN, Rawal RM, Pandya HA. Energetic contributions of amino acid residues and its cross-talk to delineate ligand-binding mechanism. *Proteins Struct Funct Bioinforma.* 2020;88(9):1207–25.
36. Głowacki ED, Irimia-Vladu M, Bauer S, Sariciftci NS. Hydrogen-bonds in molecular solids-from biological systems to organic electronics. *J Mater Chem B.* 2013;1(31):3742–53.
37. Almalki F, A G, M. H, O A. Profrens: A comparative molecular docking study into cyclooxygenase 1/2. *Drug Inven Today.* 2019;11:480–7.
38. Varma AK, Patil R, Das S, Stanley A, Yadav L, Sudhakar A. Optimized hydrophobic interactions and hydrogen bonding at the target-ligand interface leads the pathways of Drug-Designing. *PLoS One.* 2010;5(8).
39. Lot M, Hamblin MR, Rezaei N. COVID-19: Transmission, prevention, and potential therapeutic opportunities. *Int J Clin Chem Diagnostic Lab Med.* 2020;508(January):254–66.
40. United States Environmental Protection Agency. Health Effects Test Guidelines Acute Dermal Toxicity. 2002;(June).

- 359 41. Bao T, Ke Y, Wang Y, Wang W, Li Y, Wang Y, et al. Taraxasterol suppresses the growth  
360 of human liver cancer by upregulating Hint1 expression. *J Mol Med*. 2018;96(7):661–72.
- 361 42. Chen W, Li J, Li C, Fan HN, Zhang J, Zhu JS. Network pharmacology-based  
362 identification of the antitumor effects of taraxasterol in gastric cancer. *Int J*  
363 *ImmunopatholPharmacol*. 2020;34(600).
- 364 43. Yazawa K, Kurokawa M, Obuchi M, Li Y, Yamada R, Sadanari H, et al. Anti-influenza  
365 virus activity of tricin, 4',5,7-trihydroxy-3',5'-dimethoxyfavone. *Antivir Chem Chemother*.  
366 2012;22(1):1–11.
- 367 44. Lee SS, Baek YS, Eun CS, Yu MH, Baek NI, Chung DK, et al. Tricin derivatives as anti-  
368 inflammatory and anti-allergic constituents from the aerial part of *Zizania latifolia*.  
369 *BiosciBiotechnolBiochem*. 2015;79(5):700–6.
- 370 45. Seki N, Toh U, Kawaguchi K, Ninomiya M, Koketsu M, Watanabe K, et al. Tricin inhibits  
371 proliferation of human hepatic stellate cells in vitro by blocking tyrosine phosphorylation of  
372 PDGF receptor and its signaling pathways. *J Cell Biochem*. 2012;113(7):2346–55  
373