

Original Research Article

Sensitivity and resistance patterns of gram-negative uropathogens isolated from the urine of patients with upper/lower urinary obstruction in Nigeria.

## **Abstract**

**Background:** Urinary tract infection is a cause of significant morbidity and potential mortality in patients. Urine microscopy culture and sensitivity enable the isolation of the incriminating microbes. The sensitivity and resistance of the various microorganisms are invaluable in the effective management of UTIs and the associated adverse consequences. Gram-negative organisms are the usual pathogens responsible for most UTIs. The abuse of antibiotics can increase the prevalence of antimicrobial resistance. This leads to an increased cost of treatment, as more expensive higher-end antibiotics may become indicated. There is also the risk of spreading multidrug-resistant infections to the community.

**Aims:** To evaluate the sensitivity and resistance patterns of commonly available antibiotics to uropathogens in the positive urine culture of patients who presented with upper and lower urinary obstruction.

**Methods:** This retrospective study was carried out on urine samples of patients with upper/lower urinary obstructive from two specialist urology referral hospitals with a positive culture and sensitivity tests between January 2011 and December 2020. The patients' case notes were retrieved, and their urine culture, sensitivity results and mode of treatment were analyzed. These data were collated using Microsoft Excel, and they were analysed using SPSS version 20.

**Results:** 314 urine samples had positive culture and sensitivity tests. All were Gram-negatives: *Klebsiella*, *Escherichia Coli*, *Pseudomonas*, *Proteus* and *Citrobacter spp.* in decreasing frequency. Among the quinolones, levofloxacin {56.7% (178)} had the highest moderate-high

(M-H) sensitivity to the Gram-negative uropathogens; followed by Ciprofloxacin {46.2 % (145)} and Ofloxacin {19.1% (60)}.

The gram-negatives were most sensitive to streptomycin {75.5% (237)} and gentamicin 62.4% (196)} and also least resistant to them. (Streptomycin 11.1%; gentamicin 21.0%) The highest resistance was to Nalidixic acid {90.1%, (225)}, peflacin {76.1% (239)}, Augmentin {73.6% (231)} and Ampicillin {72% (226)}.

**Conclusion:** Among the commonly available antibiotics in our study, the gram-negative uropathogens are the most sensitive and least resistant to streptomycin, gentamicin and levofloxacin. Levofloxacin had the best sensitivity and lowest quinolones resistance compared to ciprofloxacin and ofloxacin. There is very low sensitivity and high resistance to nalidixic acid, ampicillin, Augmentin, Septrin and Peflacin.

**Keywords:** Bacteria, gram-negative, sensitivity, resistance, UTI

## Introduction

Urinary Tract Infection (UTI) is the inflammatory response of the urothelium to microbial invasion.<sup>1</sup> UTIs are quite common and affect men, women, young, old, immunocompetent and immunocompromised. The urinary tract should usually be free of microorganisms. Bacteria can ascend from the perineum and lead to inoculation, adherence, colonization and infection.<sup>2</sup> These processes are more likely to occur when host defence mechanisms are reduced or the virulence of the organisms increases. UTIs can also happen following haematogenous spread.<sup>3</sup>

The infection may be asymptomatic or symptomatic. In symptomatic individuals, it can cause storage symptoms, painful voiding and severe life-threatening pyelonephritis associated with pyrexia, nausea, vomiting, and rigours. Renal abscess, perinephric abscess and urosepsis can also occur following UTI. These can lead to significant morbidity, may progress to renal scarring and end-stage renal failure.<sup>4</sup>

The common organisms that cause UTIs include *Escherichia coli*, *Klebsiella pneumoniae*, *Proteus mirabilis*, *Enterococcus faecalis*, *Citrobacter* spp and *Staphylococcus saprophyticus*. Effective treatment requires evaluation with a careful history, examination, urine culture and sensitivity, and identifying the risk factor for urinary obstruction.<sup>5</sup> This ensures that an appropriate antibiotic is utilized to treat the cultured bacteria and prevent the development of resistant strains.<sup>6</sup>

Antibacterial resistance is known to increase morbidity, mortality, and cost of treatment.<sup>7,8,9</sup> As observed in our environment, indiscriminate use and abuse of antibiotics can lead to an increased prevalence of antimicrobial resistance. This increases the cost of treatment, as more expensive higher-end antibiotics may become indicated. There is also the risk of spreading multidrug-resistant infections to the community.<sup>9</sup> We aim to evaluate the sensitivity and resistance patterns to the commonly available antibiotic by uropathogenic bacteria in the urine culture of patients who presented with urine stasis.

## **Materials and Methods**

This retrospective study was carried out on urine samples of patients with upper/lower urinary obstructive from two specialist urology referral hospitals with a positive culture and sensitivity tests between January 2011 and December 2020. The patients presented to the Urology

Department University of Port Harcourt Teaching Hospital, and Rosivylle Clinic and Urology Centre, Port Harcourt, River, Nigeria. All who had a positive culture with clinical or imaging features of upper and lower urinary tract obstruction and stasis were included in the study. The folders were retrieved, and their age, sex, urine culture and sensitivity results, and mode of treatment were analyzed. The culture and sensitivity tests were carried out using gram-negative and gram-positive culture discs. The degree of sensitivity is quantified as +1= low sensitivity; +2= moderate sensitivity; +3= high sensitivity; Mild to Moderate sensitivity = M-M and Moderate to High sensitivity = M-H. Patients with incomplete records and without sensitivity reports were excluded from the study. These data were collated using Microsoft Excel version 2016, and they were analyzed using SPSS version 20.

## Results

Three hundred and fourteen patients had uropathogens cultured from their urine samples. The organisms were all gram-negative: *Escherichia coli*, *Klebsiella sp.*, *E. Coli*, *Pseudomonas spp*, *Proteus spp* and *Citrobacter spp*. in decreasing order of frequency. The gram-negative uropathogens had the highest activity and lowest resistance to streptomycin, gentamicin and rifampicin. The lowest sensitivity and highest resistance were observed with nalidixic acid and the Penicillin -Ampicillin and Augmentin.

The antibiotic with the highest sensitivity was streptomycin, with the cultured organisms expressing M-H in 75.5% (237) and resistance of 11.1%(35 ). M-H sensitivity to gentamicin was noted in 62.4% (196), and resistance was observed in 21.0% (66).

Levofloxacin had the best activity on the gram-negative organisms of the quinolones, with 56.7% (178) M-H sensitivity and 20.4% (64) resistance. Ciprofloxacin had M-H sensitivity to the uropathogens and was observed in only 46.2% (145), with resistance seen in 27.7% (87)

10.9% (34) of the gram-negative organism had M-H sensitivity to ampicillin; 25.5% (80) had M-H sensitivity to Amoxicillin, and 9.6% (30) were M-H sensitive to Augmentin.

Nalidixic acid had the least sensitivity, and the uropathogens all showed the highest resistance against it. The M-H sensitivity to nalidixic acid was only 4.1% (13) of the cultured uropathogens. 90.1% (283) of the gram-negative organism were resistant to nalidixic acid.

Table 1. Combined sensitivity and resistance pattern of uropathogens to common antibiotics.

Sensitivity and resistance pattern to various antibiotics. (+1 = *Mild sensitivity*; 2+ = *Moderate sensitivity*; 3+ = *Highly sensitive*)

SENSITIVITY/RESISTANCE	1+	2+	3+	Resistance
CIPROFLOXACIN	82(26.1)	95 (30.3)	50 (15.9)	87 (27.7)
NORFLOXACIN	52(16.6)	43 (13.7)	12 (3.8)	207 (65.9)
GENTAMICIN	52(16.6)	153 (48.7)	43 (13.7)	66 (21.0)
AMOXICILLIN	53(16.9)	59 (18.8)	21 (6.7)	181 (57.6)

STREPTOMYCIN	42(13.4)	145 (46.2)	92 (29.3)	35 (11.1)
PEFLACINE	70(22.3)	52 (16.6)	13 (4.1)	179 (57.0)
RIFAMPICIN	96(30.6)	58 (18.5)	82 (26.1)	78 (24.8)
ERYTHROMYCIN	95(30.3)	70 (22.3)	25 (8.0)	124 (39.5)
CHLORAMPHENICOL	64(20.4)	102 (32.5)	49 (15.6)	99 (31.5)
AMPICLOX	72(22.9)	51 (16.2)	11 (3.5)	180 (57.3)
LEVOFLOXACIN	72(22.9)	120 (38.2)	58 (18.5)	64 (20.4)
TARIVID	10(33.4)	47 (15.0)	13 (4.1)	149 (47.5)
REFLACINE	57(18.2)	15 (4.8)	3 (1.0)	239 (76.1)
AUGMENTIN	53(16.9)	25 (8.0)	5 (1.6)	231 (73.6)
NALIDIXIC ACID	18(5.7)	11 (3.5)	2 (.6)	283 (90.1)
SEPTRIN	51(16.2)	34 (10.8)	4 (1.3)	225 (71.7)
AMPICILLIN	54(17.2)	31 (9.9)	3 (1.0)	226 (72.0)

---

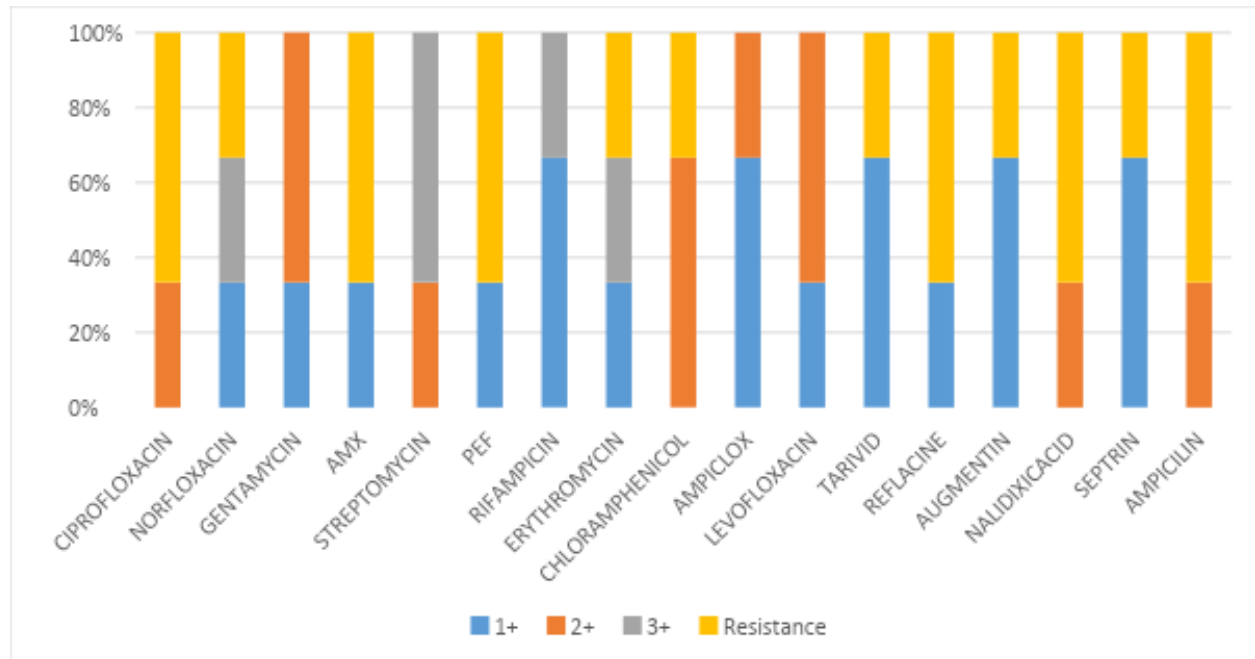


Figure 1. Sensitivity and resistance pattern of *Citrobacter spp.* to antibiotics. (+1 = Mild sensitivity; 2+ = Moderate sensitivity; 3+ = Highly sensitive)



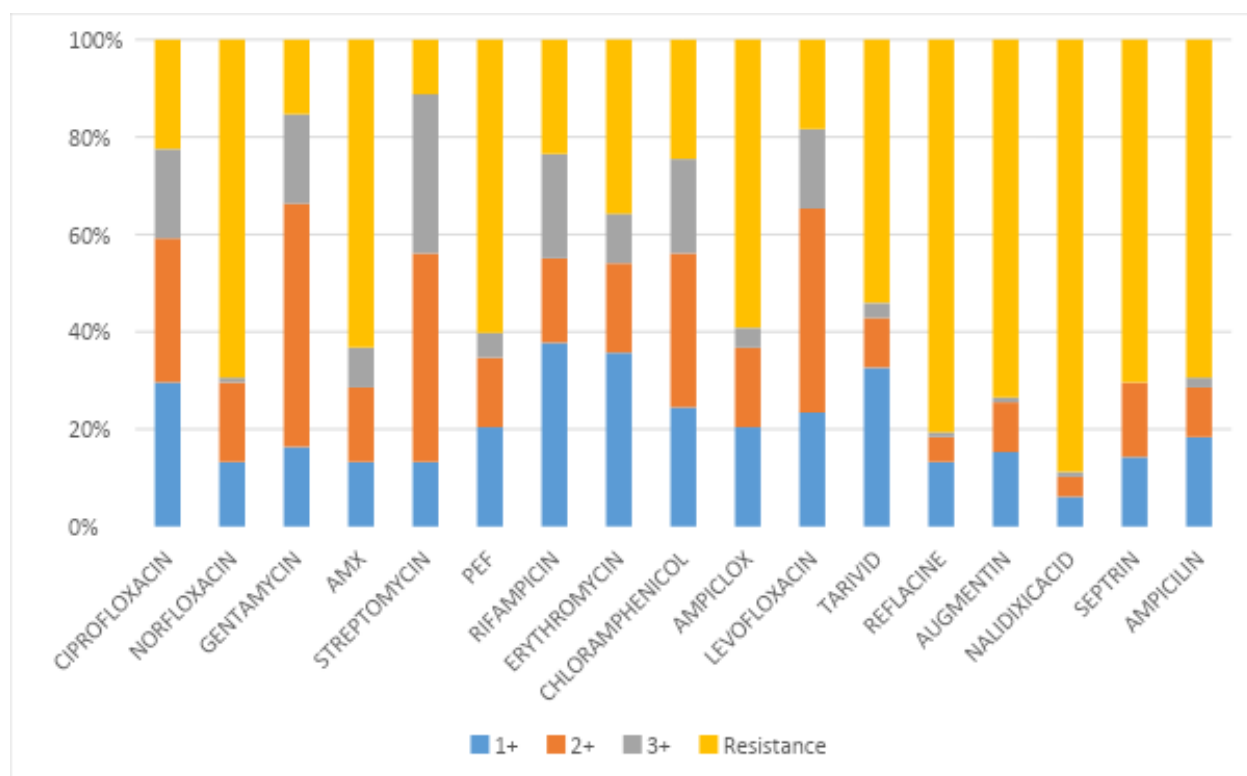


Figure 2. Sensitivity and resistance pattern of *Escherichia Coli* to common antibiotics. (+1 = Low sensitivity; 2+ = Moderate sensitivity; 3+ = Highly sensitive)

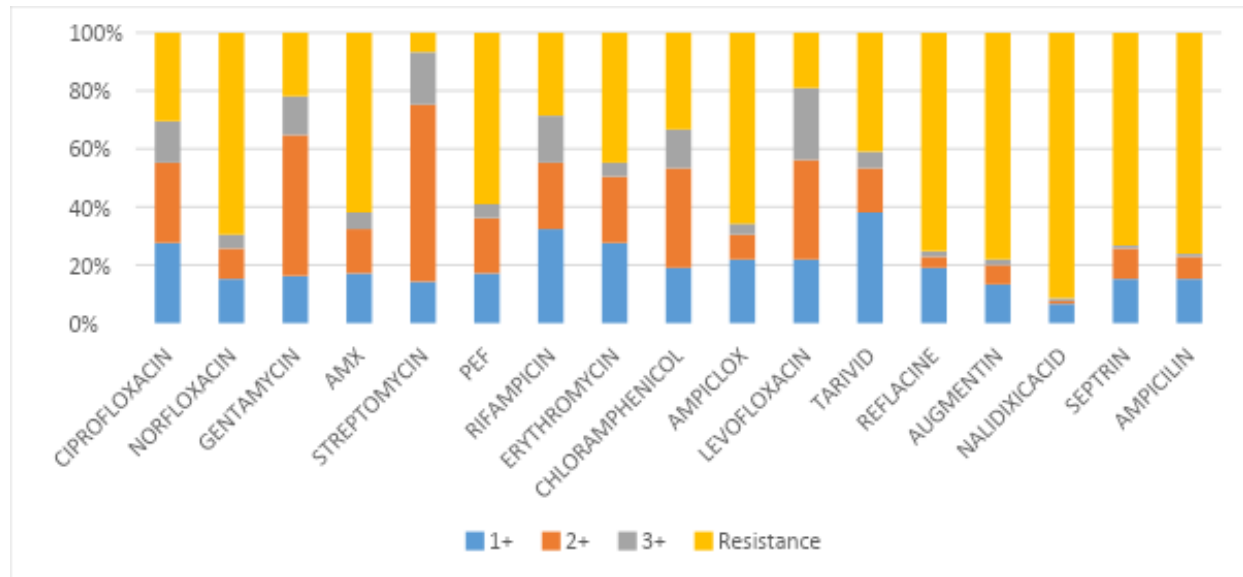


Figure 3. Sensitivity and resistance pattern of *Klebsiella sp.* to common antibiotics. (+1 = Low sensitivity; 2+ = Moderate sensitivity; 3+ = Highly sensitive)

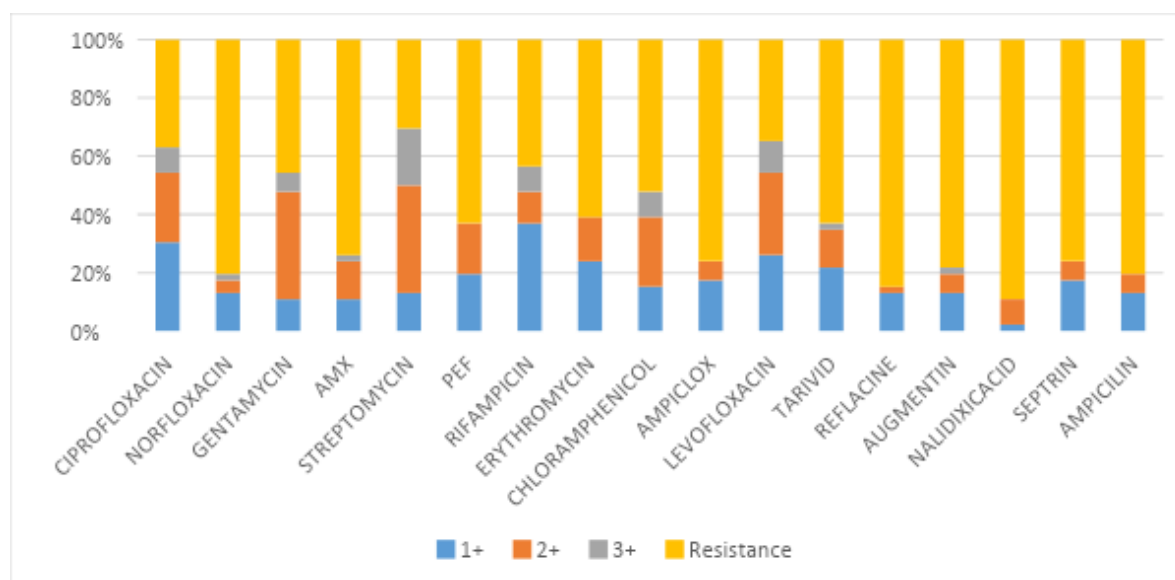


Figure 4. Sensitivity and resistance pattern of *Pseudomonas sp.* to common antibiotics. (+1 = Low sensitivity; 2+ = Moderate sensitivity; 3+ = Highly sensitive)

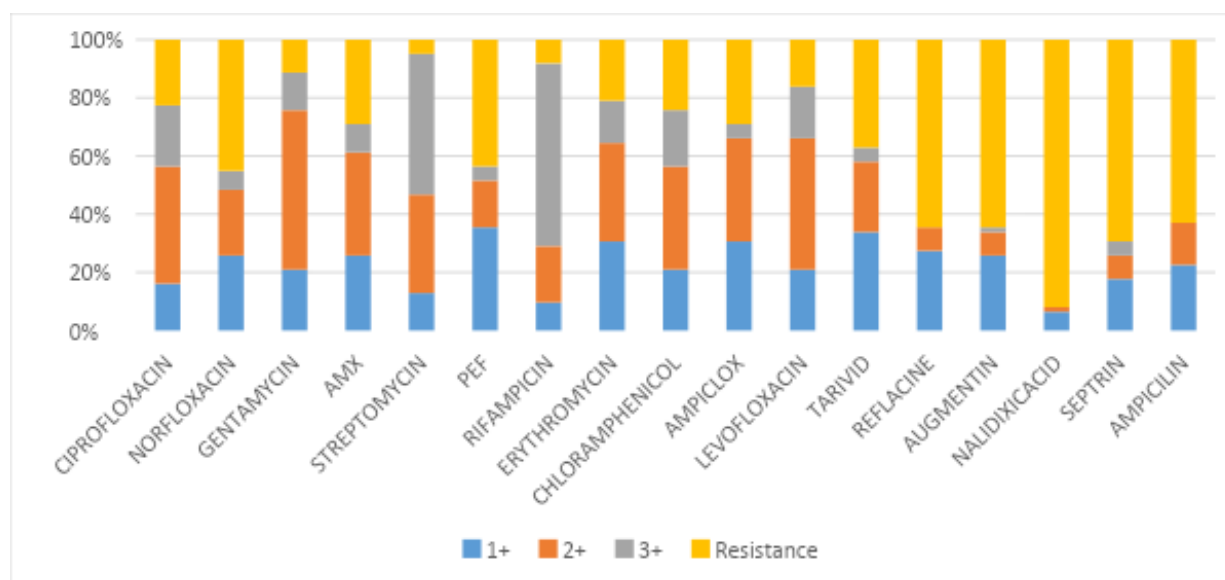


Figure 5. Sensitivity and resistance pattern of *Proteus sp.* to antibiotics. (+1= *Low sensitivity*; 2+= *Moderate sensitivity*; 3+ = *Highly sensitive*)

## Discussion

The treatment objective of UTIs is essentially to eliminate proliferating bacteria in the urinary tract, which usually occurs within hours of administering the appropriate antibiotic. This underscores the invaluable premium and critical importance of using the right antibiotics during antimicrobial therapy. It should be excreted in the urine for the antibiotic to be effective. The level should be above the minimum inhibitory concentration (MIC) for the infecting organism.<sup>10</sup>

The activity of antimicrobial agents, besides the side effect profiles, is the most crucial consideration in managing UTIs.

Gram-negative organisms are the commonest organisms cultured in the urine from most studies worldwide in both sexes.<sup>11-17</sup> The route of infection is ascending from the perineum, from its situation near the anus. Uropathogens use different mechanisms for survival once in the urinary tract in response to stresses in the bladder, such as starvation and immune responses. By forming biofilms and undergoing morphological changes, uropathogens can persist and cause recurrent infection.<sup>18,19</sup>

Streptomycin is the first discovered aminoglycoside antibiotic, originally isolated from the bacteria *Streptomyces griseus*.<sup>20</sup> It is now used mainly in the treatment of tuberculosis. It has additional activity against gram-negative organisms hence its sensitivity to uropathogens.<sup>21</sup> The primary mechanism of action is inhibition of protein synthesis.<sup>22</sup> In this study, streptomycin was found to have the highest sensitivity and least resistance to the uropathogenic gram-negative organisms. (Tables 1 and Figures 1-5) The drug is administered via the parenteral route, and abuse is seldom. It is also ototoxic and nephrotoxic and should be used with caution, especially with other aminoglycosides. It is essential in tuberculosis treatment, and hence routine use for the treatment of UTIs may not be advisable. Such use can lead to resistance to uropathogens and increase the prevalence of multidrug drug-resistant tuberculosis. A common mechanism of bacterial resistance is via downregulation of drug uptake and modification of enzymes expressed by the bacteria.<sup>23</sup> A possible reason for the high sensitivity and low resistance of streptomycin among the gram-negative organisms is the restrictive or near-exclusive use for tuberculosis treatment. Also, abuse is expected to be less since it is a parenteral medication and is less utilized than readily available oral medications.

In our study, gentamicin had the second-best activity on the uropathogens, with an M-H sensitivity of 75.5% (237) and a low resistance of 13.7% (43). It is also used parenterally only, and hence it is less likely to be abused. Its mechanism of action, side effects, and development of resistance are similar to rifampicin, the second most sensitive antibiotic in this study.

Rifampicin was discovered in 1965 by Professor Piero Sensi.<sup>24</sup> It is on the World Health Organization's list of essential medicines. It is made by the soil bacterium *Amycolatopsis rifamycinia*.<sup>25</sup> The primary mechanism of action of gentamycin and rifampicin is the inhibition of bacterial DNA-dependant RNA polymerase.<sup>24</sup> The drug is used mainly in treating tuberculosis but can also treat leprosy, legionnaires and uropathogens in urine.<sup>26</sup> Rifampicin can cause hepatotoxicity leading to elevation of liver enzymes. It turns urine, sweat and tears red or orange. Rifampicin is intrinsically resistant to Enterobacteriaceae and pseudomonas specie,<sup>27</sup> However, we found the activity of rifampicin against the uropathogens and resistance of 24.8% (78) to be relatively better than many of the other antibiotics in our study, likely due to its restricted use.

Levofloxacin is a broad-spectrum antibiotic that belongs to the drug class fluoroquinolone.<sup>28</sup> It is a left-handed isomer of the medication ofloxacin.<sup>29</sup> It is used to treat many bacterial infections, including UTIs. Its primary mechanism of action is the inhibition of DNA gyrase.<sup>23</sup> The main side effects include dizziness, gastrointestinal symptoms, myalgia and tendon rupture.<sup>28</sup> It is not routinely indicated in children because of premature fusion of the growth plate and cartilage problems. Levofloxacin is the third most sensitive antibiotic in this study, with an M-H sensitivity of 56.7% (178). Resistance was noted in 20.4% (64) and was the lowest among our study's oral antibiotics. Its mechanism of developing resistance is via active efflux of the drug, mutation in DNA gyrase binding site and alteration of cell wall porins.

Ofloxacin, pefloxacin and Norfloxacin (other fluoroquinolones) were found not to be as sensitive as levofloxacin and with the gram-negative organism showing high resistance as indicated in *Table 1, Figures 1-5*.

Nalidixic acid is also a synthetic quinolone and had the least sensitivity to the gram-negative microbes, with an M-H of only 4.1%, with 90.1% of the organisms resistant in our study. It is frequently used as a urinary antiseptic. Circumspection should be exercised with its use, given the observed present level of resistance in our environment.

Ampicillin is a Beta-lactam antibiotic used to manage and treat certain bacterial infections. It is in the aminopenicillin class of medications. Its mechanism of action is via inhibition of cell wall synthesis, and it causes cell wall lysis and death.<sup>30</sup> It can be administered through the oral, intramuscular and intravenous routes. Resistance is through the production of  $\beta$ -lactamase, changes in cell wall porin size and alteration of the penicillin-binding protein.<sup>23</sup> In this study, ampicillin was the second least sensitive antibiotic. Several other studies have noted antimicrobial resistance to ampicillin.<sup>31,32</sup> In our environment, ampicillin is readily bought over the counter, and it is taken orally in most cases. These may account for the low activity and high resistance rate of uropathogenic bacteria.

Besides the biological activity of the antibiotics, it appears from our study that oral antibiotics that are frequently used in the treatment of upper respiratory tract infections, such as the penicillin, augmentin, ampiclox, and pefloxacin, display low activity and high resistance to gram-negative organisms compared to the less frequent utilized medication like streptomycin, rifampicin, and gentamycin that are given parenterally. This emphasizes the importance of enforcing and strengthening the relevant regulatory bodies to help curtail the indiscriminate use and abuse of antibiotics to combat antibiotic resistance.

## Conclusion

Among the commonly available antibiotics in our study, the gram-negative uropathogens are most sensitive and least resistant to streptomycin, gentamycin and levofloxacin. Levofloxacin had the best sensitivity and lowest quinolones resistance compared to ciprofloxacin and ofloxacin. There is very low sensitivity and high resistance to nalidixic acid, ampicillin, augmentin, septrin and peflacin. Active joint institutional and governmental effort is needed to combat the abuse of antibiotics that fosters resistance.

## References

1. Reynard J, Brewster S, Biers S. Oxford Handbook of Urology. Third edition. UK: Oxford University Press, 2013: 176-177.
2. Flores-Mireles AL, Walker JN, Caparon M, Hultgren SJ. Urinary tract infections: epidemiology, mechanisms of infection and treatment options. Nature Reviews Microbiology. 2015 May; 13(5):269-84.
3. Van Schoor J. Urinary tract infections in women. South African Family Practice. 2016 Mar 17; 58:6-10.
4. Raphael JE, Udo K. Bacteriological Spectrum of Urine Culture in patients with Obstructive Uropathy. American Journal of Medical Sciences and Medicine. 2022;10(1): 8-22.
5. Najeeb S, Munir T, Rehman S, Hafiz A, Gilani M, Latif M. Comparison of urine dipstick test with conventional urine culture in the diagnosis of urinary tract infection. J Coll Physicians Surg Pak. 2015 Feb 1; 25(2):108-114.



6. Price TK, Dune T, Hilt EE, Thomas-White KJ, Kliethermes S, Brincat C, Brubaker L, Wolfe AJ, Mueller ER, Schreckenberger PC. The clinical urine culture: enhanced techniques improve detection of clinically relevant microorganisms. *Journal of clinical microbiology*. 2016 May;54(5):1216-1222.
7. Alanazi MQ. Clinical Efficacy and Cost Analysis of Antibiotics for Treatment of Uncomplicated Urinary Tract Infections in the Emergency Department of a Tertiary Hospital in Saudi Arabia. *Therapeutics and Clinical Risk Management*. 2021;17:1209-1217.
8. Vallejo-Torres L, Pujol M, Shaw E, Wiegand I, Vigo JM, Stoddart M, Grier S et al. Cost of hospitalised patients due to complicated urinary tract infections: a retrospective observational study in countries with a high prevalence of multidrug-resistant Gram-negative bacteria: the COMBACTE-MAGNET, RESCUING study. *BMJ Open*. 2018 Apr 1;8(4): e020251.
9. Kariuki S, Dougan G. Antibacterial resistance in sub-Saharan Africa: an underestimated Open emergency. *Annals of the New York Academy of Sciences*. 2014 Sep;1323(1):43-55.
10. Hooton TM, Stamm WE. Management of acute uncomplicated urinary tract infection in adults. *Med Clin North Am* 1991;75:339–57.
11. Odoki M, Almustapha Aliero A, Tibyangye J, et al. Prevalence of Bacterial Urinary Tract Infections and Associated Factors among Patients Attending Hospitals in Bushenyi District, Uganda. *Int J Microbiol*. 2019; 2019:4246780.
12. Kayima JK, Otieno LS, Tahir A. et al., “Asymptomatic bacteriuria among diabetics attending Kenyatta National Hospital,” *East Afr Med J*.1996;73(8): 524–526.

13. Moges AF, Genetu A, Mengistu G. Antibiotic sensitivities of common bacterial pathogens in urinary tract infections in Gondar Hospital, Ethiopia. *East Afr Med J.* 2002;79 (3,):140–142.
14. J. Wanyama, “Prevalence, bacteriology and microbial sensitivity patterns among pregnant women with clinically diagnosed urinary tract infections in Mulago Hospital Labour Ward,” Makerere University, Kampala, Uganda, 2003, M.Ed. dissertation of Wanyama. View at: Google Scholar
15. R. Mayanja, C. Kiggundu, D. Kaddu-Mulindwa et al., “The prevalence of asymptomatic bacteriuria and associated factors among women attending antenatal clinics in lower Mulago Hospital,” Makerere University, Kampala, Uganda, 2005, M.Ed. Dissertation of Mayanja. View at: Google Scholar.
16. Oladeinde BH, Omoregie R, Olley M, Anunibe JA. Urinary tract infection in a rural community of Nigeria. *North American Journal of medical sciences.* 2011 Feb;3(2):75-90.
17. Abdulhadi SK, Yashua AH, Uba A. Organisms causing urinary tract infection in paediatric patients at Murtala Muhammad Specialist Hospital, Kano, Nigeria. *International Journal of Biomedical and Health Sciences.* 2021;10; 4.
18. Thanassi DG, Saulino ET, Hultgren SJ. The chaperone/usher pathway: a major terminal branch of the general secretory pathway. *Curr Opin Microbiol.* 1998;1:223–231.
19. Piatek R et al. Pilicides inhibit the FGL chaperone/usher assisted biogenesis of the Dr fimbrial polyadhesin from uropathogenic *Escherichia coli*. *BMC Microbiol.* 2013;13:131.
20. Ohnishi Y, Ishikawa J, Hara H, et al. Genome sequence of the streptomycin-producing microorganism *Streptomyces griseus* IFO 13350. *J Bacteriol.* 2008;190(11):4050-4060.

21. Petroff BP, Lucas FV. Streptomycin in urinary infections. *Annals of surgery*. 1946;123(5):808.
22. Springer B, Kidan YG, Prammananan T, Ellrott K, Böttger EC, Sander P. Mechanisms of streptomycin resistance: selection of mutations in the 16S rRNA gene conferring resistance. *Antimicrobial agents and chemotherapy*. 2001 Oct 1;45(10):2877-84.
23. Reygaert WC. An overview of the antimicrobial resistance mechanisms of bacteria. *AIMS Microbiol*. 2018;4(3):482-501.
24. Goldstein BP. Resistance to rifampicin: a review. *The Journal of antibiotics*. 2014 Sep;67(9):625-30.
25. Verma E, Chakraborty S, Tiwari B, Mishra AK. Antimicrobial compounds from actinobacteria: synthetic pathways and applications. In *New and Future Developments in Microbial Biotechnology and Bioengineering* 2018 Jan 1 (pp. 277-295).
26. Graham DB, Tripp J. Ofloxacin. InStatPearls [Internet] 2021 Sep 14. StatPearls Publishing.
27. Goldstein, B. Resistance to rifampicin: a review. *J Antibiot*. 2014;67: 625–630.
28. Bientinesi R, Murri R, Sacco E. Efficacy and safety of levofloxacin as a treatment for complicated urinary tract infections and pyelonephritis. *Expert Opinion on Pharmacotherapy*. 2020 Apr 12;21(6):637-44.
29. Bradley JS, Kauffman RE, Balis DA, Duffy CM, Gerbino PG, Maldonado SD, Noel GJ. Assessment of musculoskeletal toxicity 5 years after therapy with levofloxacin. *Paediatrics*. 2014;134(1):146-53.

30. Ghooi RB, Thatte SM. Inhibition of cell wall synthesis--is this the mechanism of action of penicillins?. *Med Hypotheses*. 1995;44(2):127-131.
31. Hrbacek J, Cermak P, Zachoval R. Current antibiotic resistance trends of uropathogens in Central Europe: Survey from a Tertiary hospital urology department 2011–2019. *Antibiotics*. 2020 Sep;9(9):630.
32. Nguyen SN, Thi Le HT, Tran TD, Vu LT, Ho TH. Clinical Epidemiology Characteristics and Antibiotic Resistance Associated with Urinary Tract Infections Caused by E. coli. *International Journal of Nephrology*. 2022 Feb 28;2022.