Original Research Article

Impact of soil biological parameters on soil health in the intensively cultivated deltaic Inceptisol of Thanjavur, Tamil Nadu

ABSTRACT

An experiment was conducted during 2021-22 at Agricultural Research Station, Kattuthottam, Thanjavur, Tamil Nadu to identify the impact of biological parameters on different cropping sequences. Three soil samples were randomly taken from each replication of every treatment making a total of 90 samples in each season from different cropping sequences during active vegetative stage. The size of each plot is $40m^2$. Samples were analysed for soil biological parameters *viz.*, Urease, Acid phosphatase, Dehydrogenase, Soil microbial count (Bacteria, fungi and actinomycetes) and Microbial Biomass Carbon. Different cropping sequences showed their effect as variations in soil biological properties. The cropping sequence T_4 , sunhemp-rice+dhaincha(10:1)-green gram showed more biological activity with urease activity of $40.6 \text{ NH}_4^+ \text{ µg/g/h}$, acid phosphatase activity of 43.1 P-NP µg/g/h, dehydrogenase activity (137.9 TPF µg/g/day), microbial biomass carbon value (307 mg kg⁻¹), bacterial count (55.6 cfu g⁻¹ soil), fungal count (23.5 cfu g⁻¹ soil) and actinomycetes count (41.2 cfu g⁻¹ soil). Rice-rice-sesame sequence was observed to have less biological activity than other cropping sequences.

Keywords: Biological properties, cropping sequence and soil health

1. INTRODUCTION

'Soil health' is described as a condition of dynamic balance between flora and fauna and their surrounding soil environment, in which all the former's metabolic functions run smoothly without any interference, impedance from the latter [1]. Soil health is the ability of soil to continue and operate as a vibrant living ecosystem that supports plants, animals and humans. Soil health is a word used frequently in conversations about sustainable agriculture to indicate the overall state and quality of the soil resource. We have clean air and water, abundant crops and forests, productive grazing grounds, diversified fauna and attractive vistas because of healthy soil. All agricultural systems require management to function properly. Despite this, there is evidence of extensive damage of the environment due to erosion and degradation of organic matter in agricultural soils. Pollution, compaction and salinity increasing as well as other negative consequences. Plant cover affects soil biological properties via quality of root exudates and decomposition of litter. The release of root exudates into the rhizosphere is known to enhance soil biological activity and alter microbial community structure. Addition of labile root exudate components to the rhizosphere induced a small but significant increase on litter decomposition but that the magnitude of this effect was regulated by temperature.

Rice (*Oryza sativa*) is a component of widely varying cropping sequences. Rice-based cropping sequences form an integral part of agriculture in Tamil Nadu. Several intense rice-based cropping sequences have been noticed and are being practiced by the farmers. While intensive agriculture, those involving exhaustive high yielding varieties of rice and other crops, has led to heavy withdrawal of nutrients from the soil by crops, imbalanced and indiscriminate use of chemical fertilizers has resulted in deterioration and degradation of soil health [2]. According to the FAO, global rice demand in 2020 would be 800 MT, whereas current output is 600 MT. To satisfy future requirements, an extra 200 million tonnes must be generated by boosting productivity per hectare [3]. Even when rice is planted with appropriate N, P and K, the productivity of the crop is dropping. Previously conducted experiment in the same established field comprising of seven different crop rotations *viz.*, rice-black gram, rice-sesame, rice+dhaincha-maize+green gram, rice+dhaincha-bhendi, rice+dhaincha-varagu, rice+dhaincha-fodder cowpea with first crop of green manure in all rotations. The results revealed that crop rotations had a significant impact on soil quality by altering their properties [4]. Suitable

rice-based cropping sequence has to be evaluated, to assess the stability in production. Hence in this context, a research work was carried out to evaluate the soil biological properties in different cropping sequences in order to ascertain the health of cultivated soil.

2. MATERIALS AND METHODS

2.1. Study site

The present investigation was carried out at Agricultural Research Station, Thanjavur. The latitude was 10°45′ north, the longitude was 79° east, and the elevation was 50 meters above mean sea level. For this study, three soil samples were collected from each plot making a total of 90 samples in each season at Agricultural Research Station, Thanjavur which is 272 km away from the Coimbatore in the eastern direction. The mean of three samples taken from each plot is considered as that replication value. The analysis was carried out in the Department of Soil Science and Agricultural Chemistry, Tamil Nadu Agricultural University, Coimbatore. There are 10 cropping sequences replicated thrice in Randomized Block Design. The area of each plot is about 40 m². The soil initial analysis showed it was sandy loam soil. Samples were collected at active vegetative stage from the root zone at 0-15 cm which were considered as surface samples, during *kharif*, *rabi* and summer seasons of 2021-22. The cropping sequences were established with recommended dosage of fertilizer. The details of the cropping sequences are as follows (Table 1)

Table 1. List of treatments in cropping sequence

Treatments	Kharif	Rabi	Summer
T ₁	Rice (Co 51)	Rice (ADT 46)	Black gram (ADT 5)
<mark>T</mark> 2	Rice (Co 51)	Rice (ADT 46)	Sesame (VRI 2)
T ₃	<mark>Dhaincha</mark>	Rice+dhaincha (5:1)	Black gram (ADT 5)
		(ADT 46)	
T_4	Sun hemp	Rice+dhaincha (10:1)	Green gram (Co 6)
		(ADT 46)	
<mark>Т₅ Т₆</mark>	Black gram (ADT 5)	Rice (ADT 46)	Groundnut (TMV 13)
T ₆	Green gram (Co 6)	Rice (Bio fortified-CR	Sesame (VRI 2)
		Dhan45)	
T ₇	Fodder Cowpea Co(FC) 9	Rice (ADT 46)	Fodder Maize (African tall)
T ₈ T ₉	Fodder Sorghum Co(FS) 29	Rice (Co 52)	Fodder Cowpea Co(FC) 9
T ₉	Maize hybrid (Co 6)	Rice (Seeraga samba)	Bhendi (Co 4)+black gram (ADT
			5) (5:1)
T ₁₀	Ragi (Co 15)	Rice (Navara)	Cluster bean (MDU1)+black gram
			(ADT 5) (5:1)

T₁ – Rice-rice-black gram; T₂ – Rice-rice-sesame; T₃ – Dhaincha-rice+dhaincha (5:1)-black gram;

2.2. Soil analysis

The soil analysis was carried out at the laboratory of Department of Soil Science and Agricultural Chemistry in Tamil Nadu Agricultural University, Coimbatore. Urease was determined by calculating μg of NH_4^+ released /g/hr [5]. Acid Phosphatase enzyme activity was determined using p-nitro phenyl phosphate as substrate colorimetrically at 410 nm [6]. The Dehydrogenase of the soil was determined as μg of TPF /g/day [7]. Microbial population (bacteria, fungi and actinomycetes) of soil were determined by [8] [9] [10] and [11] in cfu g^{-1} soil. Microbial biomass carbon of the soil sample was analysed by using fumigation extraction technique [12].

3. RESULTS AND DISCUSSION

3.1. Urease and Acid Phosphatase

From Table 2, the values of urease activity in the soil ranged from 18.6 to 39.8 NH₄ μ g/g/h with a mean value of 29.8 NH₄ μ g/g/h during the *kharif* season, the higher value was recorded under T₄ cropping sequence and the lower value was recorded under T₉ cropping sequence. In *rabi* season, the values of urease activity of the soil ranged from 22.9 to 40.6 NH₄ μ g/g/h with a mean value of 30.4 NH₄ μ g/g/h. The higher value of urease activity present under T₄ cropping sequence and the lower present under T₉ cropping sequence. During summer season values of urease activity of the soil

 T_4 – Sun hemp-rice+dhaincha-green gram; T_5 – Black gram-rice-groundnut; T_6 – Green gram-rice-sesame; T_7 – Fodder cowpea-rice-fodder maize; T_8 – Fodder sorghum-rice-fodder cowpea; T_9 – Maize hybrid-rice-bhendi+black gram; T_{10} – Ragi-rice-cluster bean+black gram

ranged from 16.0 to 31.1 NH $_4$ µg/g/h with a mean value of 24.5 NH $_4$ µg/g/h. The highest urease value was recorded in T $_4$ cropping sequence whereas the lowest value was found in T $_2$ cropping sequence. The inclusion of green manure crop and legume black gram in the T $_4$ cropping sequence may have caused the greatest urease activity to be detected. This crop may have fixed more N than in the T $_2$ crop [13]. The soil of the experiment was acidic in nature. Higher urease activity in an acidic soil indicated soil N gain. This is mainly due to the inclusion of readily decomposable organic materials which did not hinder abiotic urease's capacity to hydrolyze urea, and thus considerably enhanced the microbiological production of urease by soil microorganisms [14].

Acid phosphatase activity of the soil samples ranged from 21.4 to 27.5 P-NP μ g/g/h with a mean value of 25.1 P-NP μ g/g/h during the *kharif* season, the higher value was recorded under T₂ cropping sequence. In *rabi* season, the values of acid phosphatase activity of the soil ranged from 19.2 to 33.6 NH₄ P-NP μ g/g/h with a mean value of 26.3 NH₄ P-NP μ g/g/h. The higher value of acid phosphatase activity present under T₄ cropping sequence and the lower present under T₂ cropping sequence. During summer season, values of acid phosphatase activity of the soil ranged from 25.5 NH₄ to 43.6 NH₄ P-NP μ g/g/h with a mean value of 33.5 NH₄ P-NP μ g/g/h. The highest acid phosphatase activity was recorded in T₄ cropping sequence whereas the lowest value was found in T₉ cropping sequence. Legumes in agricultural rotations may have generated more acid phosphatase enzymes than non-legume crops. Cereals have symbiotic associations with root fungi known as arbuscular mycorrhizal fungi for the purpose of supplying phosphorus. Furthermore, compared to legumes and green manure crops, cereals do not require as much P for the symbiotic N fixation process [15] [16].

Table 2. Effect of various cropping sequence on Urease and Phosphatase

	<mark>Urease (NH₄ μg/g/h)</mark>		Acid Phosphatase (P-NP µg/g/h)			
Cropping sequence	Kharif	Rabi	Summer	Kharif	Rabi	Summer
<mark>T₁</mark>	<mark>39.8</mark>	<mark>40.6</mark>	31.1	<mark>24.2</mark>	30.1	<mark>39.3</mark>
T_2	<mark>19.9</mark>	<mark>24.0</mark>	<mark>16.0</mark>	<mark>21.4</mark>	<mark>19.2</mark>	<mark>27.8</mark>
T ₃	<mark>35.3</mark>	<mark>35.1</mark>	<mark>29.5</mark>	<mark>25.6</mark>	<mark>32.4</mark>	<mark>41.9</mark>
T ₄	<mark>38.4</mark>	<mark>38.7</mark>	<mark>30.2</mark>	<mark>27.5</mark>	<mark>33.6</mark>	<mark>43.6</mark>
${\sf T_5}$	<mark>32.1</mark>	<mark>27.5</mark>	<mark>28.3</mark>	<mark>26.9</mark>	<mark>22.5</mark>	<mark>37.4</mark>
T ₆	<mark>31.7</mark>	<mark>29.2</mark>	<mark>26.9</mark>	<mark>24.7</mark>	<mark>21.7</mark>	<mark>36.1</mark>
T ₇	<mark>29.2</mark>	<mark>30.4</mark>	<mark>19.7</mark>	<mark>24.5</mark>	<mark>29.8</mark>	<mark>35.5</mark>
T ₈	<mark>28.0</mark>	<mark>30.8</mark>	<mark>23.6</mark>	<mark>26.7</mark>	<mark>27.3</mark>	<mark>29.0</mark>
T ₉	<mark>18.6</mark>	<mark>22.9</mark>	18.1	<mark>23.9</mark>	<mark>20.8</mark>	<mark>25.5</mark>
T ₁₀	<mark>25.6</mark>	<mark>25.6</mark>	<mark>21.8</mark>	<mark>25.1</mark>	<mark>25.9</mark>	<mark>27.4</mark>
Mean	<mark>29.8</mark>	30.4	<mark>24.5</mark>	<mark>25.1</mark>	<mark>26.3</mark>	<mark>33.5</mark>
SEd	0.72	0.61	<mark>0.59</mark>	<mark>0.62</mark>	<mark>0.63</mark>	<mark>0.62</mark>
CD (0.05)	<mark>1.52</mark>	1.29	<mark>1.24</mark>	<mark>1.32</mark>	<mark>1.33</mark>	<mark>1.31</mark>

3.2. Dehydrogenase and Microbial biomass carbon

From Table 3, the values of dehydrogenase enzyme of the soil ranged from 21.1 to 33.6 TPF μ g/g/day with a mean value of 27.9 NH₄ TPF μ g/g/day during the *kharif* season, the higher value was recorded under T₄ cropping sequence and the lower value was recorded under T₈ cropping sequence. In *rabi* season, the values of dehydrogenase in the soil ranged from 42.7 to 137.9 TPF μ g/g/day with a mean value of 81.3 TPF μ g/g/day. The higher value of dehydrogenase activity present under T₄ cropping sequence and the lower value is present under T₂ cropping sequence. During summer season, values of dehydrogenase value in the soil ranged from 23.2 to 36.4 TPF μ g/g/day with a mean value of 29.3 TPF μ g/g/day. The highest dehydrogenase activity was recorded in T₄ cropping sequence whereas the lowest value was found in T₁₀ cropping sequence. There is a direct relationship between soil moisture and dehydrogenase activity. Soil organic matter that increases soil moisture retention will increase soil dehydrogenase activity. The activity was brought about by the presence of legume and green manure crop in the cropping sequence, which improved organic matter addition and root exudation [17]. *Rabi* season dehydrogenase levels were greater, which may be related to the increased organic matter storage caused by the cooler temperatures and favourable dehydrogenase activity-promoting conditions. Additionally, [18] noted that autumn (wet) had greater dehydrogenase activity than dry season.

In *kharif* season, microbial biomass carbon of the soil samples ranged from 223 to 279 mg kg⁻¹ with a mean value of 250 mg kg⁻¹, the higher value was recorded under T₄ cropping sequence and the lower value was recorded under T₂ cropping sequence. In *rabi* season, the values of microbial biomass carbon of the soil ranged from 245 to 307 mg kg⁻¹ with a mean value of 273 mg kg⁻¹. The higher value of microbial biomass carbon present under T₄ cropping sequence and the lowest present under T₂ cropping sequence. During summer season, values of microbial biomass carbon of the soil ranged from 152 to 269 mg kg⁻¹ with a mean value of 213 mg kg⁻¹. The highest microbial biomass carbon was recorded in

 T_4 cropping sequence whereas the lowest value was found in T_9 cropping sequence. This is due to higher microbial activity that will eventually lower C:N ratio and increase the availability of Nitrogen to plants. Compared to other fractions of organic matter the turnover rate of soil microbial biomass carbon is rapid and higher due to microbial activity [19]. Cropping sequence had significant impact on soil microbial carbon.

Table 3. Effect of various cropping sequences on soil dehydrogenase activity and soil microbial biomass carbon

	Dehydrogenase (TPF μg/g/day)			Microbial biomass carbon (mg kg ⁻¹)		
Cropping sequence	Kharif	<u>Rabi</u>	Summer	<u>Kharif</u>	<u>Rabi</u>	Summer
T ₁	<mark>23.5</mark>	<mark>50.8</mark>	<mark>24.6</mark>	<mark>237</mark>	<mark>251</mark>	<mark>189</mark>
T_2	<mark>30.4</mark>	<mark>42.7</mark>	34.6	<mark>223</mark>	<mark>245</mark>	<mark>169</mark>
T ₃	<mark>32.8</mark>	112.0	33.1	<mark>241</mark>	<mark>295</mark>	<mark>254</mark>
T_4	<mark>33.6</mark>	137.0	<mark>36.4</mark>	<mark>279</mark>	<mark>307</mark>	<mark>269</mark>
<mark>T</mark> ₅	<mark>28.3</mark>	<mark>74.2</mark>	32.0	<mark>265</mark>	<mark>287</mark>	<mark>237</mark>
T ₆	<mark>26.2</mark>	<mark>92.4</mark>	<mark>30.1</mark>	<mark>267</mark>	<mark>278</mark>	<mark>234</mark>
T ₇	<mark>29.6</mark>	<mark>89.3</mark>	<mark>27.7</mark>	<mark>259</mark>	<mark>274</mark>	<mark>222</mark>
T ₈	21.1	<mark>63.0</mark>	<mark>26.9</mark>	234	<mark>256</mark>	<mark>217</mark>
T ₉	<mark>24.8</mark>	<mark>44.6</mark>	<mark>25.3</mark>	243	<mark>263</mark>	<mark>152</mark>
T ₁₀	<mark>28.9</mark>	106.0	<mark>23.2</mark>	253	<mark>280</mark>	<mark>195</mark>
<mark>Mean</mark>	<mark>27.9</mark>	<mark>81.3</mark>	<mark>29.3</mark>	<mark>250.1</mark>	273.6	<mark>213.8</mark>
SEd	0.59	<mark>1.34</mark>	0.45	<u>5.15</u>	<mark>6.09</mark>	<mark>5.44</mark>
CD (0.05)	<mark>1.24</mark>	<mark>2.83</mark>	0.96	<mark>10.81</mark>	<mark>12.79</mark>	<mark>11.42</mark>

3.4. Soil microbial population (Bacteria, fungi and actinomycetes)

From fig. 1, the bacterial population in the soil ranged from 41.1 to 50.1 cfu g^{-1} soil with a mean value of 45 cfu g^{-1} soil during the *kharif* season, the higher value was recorded under T_4 and the lower value was recorded under T_2 . In *rabi* season, the values of bacterial population of the soil ranged from 43.4 to 55.6 cfu g^{-1} soil with a mean value of 49 cfu g^{-1} . The higher value of bacterial population is present under T_4 cropping sequence and the lower value present under T_2 cropping sequence. During summer season, values of bacterial population of the soil ranged from 39.2 to 48.7 cfu g^{-1} with a mean value of 44.8 cfu g^{-1} . The highest bacterial population was recorded in T_4 cropping sequence whereas the lowest value was found in T_8 cropping sequence.

Fungal population of the soil samples ranged from 10.8 to 19.6 cfu g^{-1} with a mean value of 15.1 cfu g^{-1} during the *kharif* season, the higher value was recorded under T_4 cropping sequence and the lower value was recorded under T_2 cropping sequence. In *rabi* season, the values of fungal population of the soil ranged from 12.3 to 23.5 cfu g^{-1} with a mean value of 17.7 cfu g^{-1} . The higher value of fungal population is present under T_4 cropping sequence and the lower value present under T_2 cropping sequence. During summer season, values of fungal population of the soil ranged from 9.2 to 16.1 cfu g^{-1} with a mean value of 12.6 cfu g^{-1} . The highest fungal population was recorded in T_4 cropping sequence whereas the lowest value was found in T_9 cropping sequence.

Actinomycetes population of the soil samples ranged from 33.7 to 38.5 cfu g^{-1} with a mean value of 36 cfu g^{-1} during the *kharif* season, the higher value was recorded under T_4 cropping sequence and the lower value was recorded under T_2 cropping sequence. In *rabi* season, the values of actinomycetes population of the soil ranged from 36.9 to 41.2 cfu g^{-1} with a mean value of 39 cfu g^{-1} . The higher value of actinomycetes population present under T_4 cropping sequence and the lower value is present under T_2 cropping sequence. During summer season, values of actinomycetes population of the soil ranged from 31.1 to 35.4 cfu g^{-1} with a mean value of 33.7 cfu g^{-1} . The highest actinomycetes value was recorded in T_4 cropping sequence whereas the lowest value was found in T_9 cropping sequence.

In this study, the legume-dominated sequence with green manure crop had a higher microbial population than the cereal-dominated system because the former system was better for the growth and development of the soil microorganisms. This could be because bacteria and the nodules found in the roots of the leguminous crops have a symbiotic interaction. Bacteria can use 2 sources of carbon: organic and inorganic carbon. on the other hand, fungi and actinomycetes can use only organic carbon source. Bacteria are also more saprophytically more competent than fungi and actinomycetes in that sequence. Any cropping sequence that has the highest population of fungi is an indication that it has more organic than inorganic carbon and that it is a source of not easily decomposable organic material. Conversely, cropping sequence with higher bacteria population shows that it is a rich source of easily decomposable organic material. Due to their structure and C:N ratio between 7:1 and 25:1, fungi need a greater amount of carbon to grow and reproduce and will therefore collect the required from the soil organic matter. Bacteria, which have a smaller C:N ratio than fungi (between 5:1 and 7:1), need food rich in nitrogen like green manure, legume residues etc because they have higher nitrogen requirement and take more nitrogen from the soil for their own requirements. Legumes may also

enhance SOC, which is good for the growth of microorganisms, and their roots produce sugar-like compounds that aid in the growth of soil bacteria in the rhizosphere [20] [21]. Legume-based cropping methods boost microbiological and enzymatic activity because they supply high-quality biomass (low C:N ratio) to the soil through active root development and exudation, nodule degeneration and leaf shedding. Different environmental conditions in the root zone are brought about by the legume systems, which have an impact on nutrient uptake and carbon exudation [22].

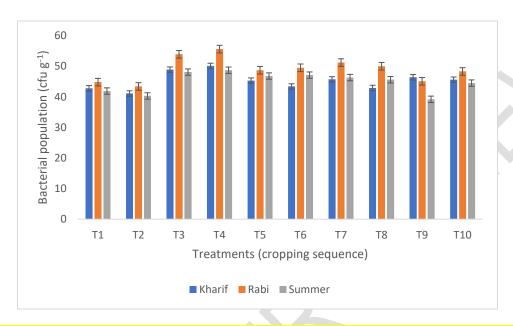


Fig. 1. Effect of various cropping sequences on bacterial count in different seasons

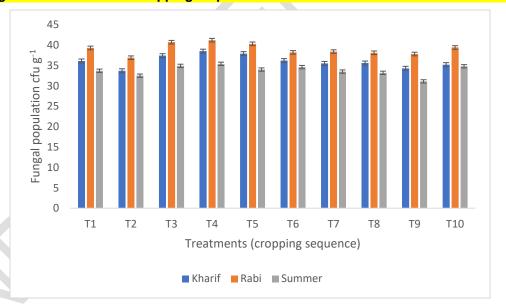


Fig. 2. Effect of various cropping sequences on fungal count in different seasons

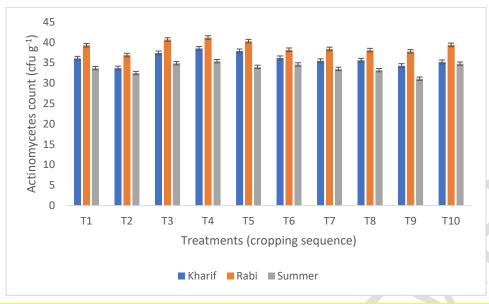


Fig. 3. Effect of various cropping sequences on actinomycetes count in different seasons

4. CONCLUSION

From the investigation, it was observed that the soil biological parameters differ significantly due to the effect of various cropping sequences. It was observed that, most of the parameters show higher values in *rabi* season than *kharif* and summer. Under various cropping sequences taken for research work, sunhemp-rice+dhaincha(10:1)-green gram cropping sequence shows higher fertility status than other cropping sequences as it contains higher biological activity and better soil properties than others due to inclusion of green manure and legume crop. The cropping system rice-rice-sesame sequence shows minimum value in most of the soil biological properties than other cropping sequence.

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