Original Research Article

Effect of fertility levels and biofertilizers on growth parameters, root architecture and quality of chickpea (*Cicer arietinum* L.)

Abstract:

The present field experiment was conducted during Rabi season of 2017–18 at the Student's Instructional Farm of Chandra Shekhar Azad University of Agriculture and Technology, Kanpur, Uttar Pradesh to evaluate the effect of different fertility levels and biofertilizers on growth parameters, root architecture and quality of late sown chickpea (*Cicer arietinum* L.). The experiment comprised of 12 treatment combinations in split plot design which comprised 4 treatments [F_1 (control), F_2 (RDF 100%), F_3 (75% RDF), F_4 (50% RDF)] in main plot and 3 treatments [B_1 (*Rhizobium* + PSB), B_2 (*Rhizobium* + PGPR) and B_3 (*Rhizobium* + PSB + PGPR)] in sub plots with three replications. Results showed that among the different fertility levels, application of 100% RDF significantly enhanced growth parameters, root architecture and protein yield over the control. Among the different biofertilizers treatments application of *Rhizobium* + PSB + PGPR had significantly improved growth parameters, root architecture and protein yield as compared to *Rhizobium* + PGPR. The combined application of 100% RDF with *Rhizobium* + PSB + PGPR resulted in significantly higher growth parameters, root architecture and protein yield of late sown chickpea during winter (*Rabi*).

Key Words:- Biofertilizers, Chickpea, Fertility Levels, Protein, Root Architecture and Yield

Introduction

"Pulses play a pivotal role and occupy a unique position in Indian agriculture by virtue of their inherent capacity to grow on marginal lands. It is an easily available source of protein in the rural heart of India. Pulses provide significant nutritional and health benefits and are known to reduce several non- communicable diseases such as colon cancer and cardio-vascular diseases" (**Jukanti** *et al.*, **2012**). India is the largest producer and consumer of pulses in the world. Major pulses grown in India include chickpea, pigeonpea, lentil, urd bean, mung bean, pea, lablab bean, moth bean, horse bean. Among the pulses, chickpea is the most important grown in every part of India. "It is largest produced food legume in South Asia. Chickpea (*Cicer arietinum* L.) is a major legume crop cultivated for its edible seeds legume of the genus *Cicer*, Tribe *Cicereae*, family *Fabaceae* (*leguminaceae*), and subfamily

Papilionaceae. It provide protein rich diet to the vegetarian of the Indian and complement the stable cereals in the diets with proteins, essential amino acids, vitamins and minerals" (Pingoliva et al., 2013). "Pulses play an equally important role in rainfed and irrigated agriculture by improving physical, chemical and biological properties of soil and are considered excellent crop for natural resource management, environmental security, crop diversification and consequently for viable agriculture" (Ali and Kumar, 2006). Globally, India is recognized as, a major player in pulses contributing 25% global production, (4-6 mt.) and consumer (26-27mt). Import duty on chickpea has been fixed at 60%. The year 2017-2018 had, witnessed record production in pulses (25.23mt). In India, Madhya Pradesh is the largest pulse producing state, which accounts for 23% of total pulse production. It covers 32.97% area of chickpea in country. Chickpea is the King of pulses consist of more than 1/3 of area and 40% total pulse production. In Asian region chickpea is the premier pulse crop of Indian sub-continent. India is the largest producer as well as consumer of chickpea in the world. It is grown in area of 6.3million hectare with production of 5.1 mt. The average yield of chickpea is 806kg/hec. (FAOSTAT 2017-18). "It is an important source of energy, protein, soluble and insoluble fibre. Mature chickpea grains contain 60-65% Carbohydrates, 6% Fat and 12-25% Protein higher than any other pulse crop. Through symbiotic Nitrogen fixation, the crop meet up to 80% of soil nitrogen needs, so farmers have to apply less N fertilizers". (Das et al., 2012). Legumes are heavy feeder of phosphorus and less responsive to nitrogen because of their capacity to meet their own nitrogen requirement through symbiotic fixation (Kumar et al., 2016). "Phosphorus is connected with some particular plant growth factors that are root development, vigorous stem, enhanced flower formation and seed production, earlier and more uniform crop maturity, increase nitrogen fixing capacity of legumes, improvement in crop quality and resistance to plant diseases" (Rehan et al., 2018). "It is required for higher and sustainable production of grain legumes. Generally, legumes have higher P requisites due to more consumption of energy in the process of symbiotic nitrogen fixation" (Islam et al., 2012). "Biofertilizers promote plant growth and development also reduce the cost of production as they tend to decrease the doses of chemical fertilizers used. These can be used for fodder, food, vegetables and leguminous crops. Commonly used microorganisms as biofertilizers are Rhizobia, Phosphate Solubilizing Bacteria (PSB) and Plant Growth Promoting Rhizobacteria (PGPR). Seed inoculation with Rhizobium increases the nodulation through better root development and improves nutrient availability which is beneficial in improving the grain yield" (Ali et al., 2004). Inoculation of chickpea with Rhizobium significantly increased the nodulation and its dry weight, plant

height, pods plant⁻¹, 1000-grain weight, root length, root dry weight and grain yield (**Akhtar** *et al.*, **2009**). "Rhizobium and phosphate solubilizing bacteria (PSB) assume a great importance on account of their vital role in N2 fixation and P solubilizations. Use of Rhizobium and PSB had shown advantage in enhancing chickpea productivity" (**Rudresh** *et al.* **2005**).

Materials and Methods

Experimental site and Climate

The experiment was laid out in at the SIF Farm of CSAUA&T, Kanpur, Uttar Pradesh. It is located on 25°18′ N latitude, 83°03′ E longitude and at an altitude of 80.71 meters above mean sea level. Experimental site area, Kanpur is situated in the central part of U.P. and have sub-tropical climate, characterized by hot summer and cool winters. Total rainfall received during the crop growing period was 15.90 mm.

Soil analysis

The experimental field is sandy clay loam in texture, neutral in reaction (pH 7.6), EC (0.11 dSm⁻¹), low in organic carbon (0.30%), available N (188 kgha⁻¹), medium in available P (13.4 kg ha⁻¹) and available K (173.3 kgha⁻¹).

Table 1: Mechanical and chemical of soil analyses of the experimental field

Particular	Values	Method employed						
Mechanical Analysis								
Soil separates (%)								
i) Coarse sand	8.48							
ii) Fine sand	53.44	Hydromotor mothod (Douyouana						
iii) Silt	19.46	Hydrometer method (Bouyoucos, 1927)						
iv) Clay	17.85	1927)						
Textural class	Sandy clay loam							
Chemical Analyses								
i) Soil reaction (pH)	7.6	Glass electrode pH meter						
(1: 2.5 soil and water suspension)		(Jackson, 1973)						
ii) Electrical conductivity	0.11	Sytronics electrical conductivity						
(dSm ⁻¹ at 25°C)		meter (Jackson, 1973)						
iii) Organic carbon (%)	0.30	Walkley and Black's Method, 1947						
iv) Available N (kg ha ⁻¹)	188	Alakaline permanganate (Subbiah and						
		Asija, 1956)						
v) Available P (kg ha ⁻¹)	13.42	0.5 N NaHCO ₃ extractable						
		(Olsen et al, 1954)						
vi) Available K (kg ha ⁻¹)	173.30	Ammonium acetate extractable flame						
		photometer (Toth and Price, 1949)						

Treatments of Investigation

The experiment was consists of 12 treatment combinations and laid out in split plot design assigning four treatments in main plot *viz*. F1- Control, F2- RDF 100%, F3- RDF 75%, F4- RDF 50% and three treatments in sub plot *viz*. B1- Rhizobium+PSB, B2- Rhizobium+PGPR, B3- Rhizobium+PSB+PGPR with three replications. Each treatment was randomly allocated with in them.

Application of Fertilizers

The crop was fertilized with a recommended dose of @ 20-60-20-20 kg nitrogen, phosphorus potassium and sulphur/ha, respectively. Urea DAP, MOP and gandhak powder were used as the source of nitrogen, phosphorus, potassium and Sulphur respectively.

Seed Treatment

Culture of biofertilizers i.e. Rhizobium, PGPR and PSB, each packet has 200g weight and used for seed treatment at the rate of 20g/kg seed. Seeds were treated with *biofertilizers* (20 g per kg of seed) as per standard procedure and were sown after drying for six hours under shade.

Sowing of Seed

Chickpea seeds were sown at 75 kg ha⁻¹ in the furrows opened by the kudal by manual labours at 40 cm row to row spacing and 10 cm plant to plant spacing.

Protein Analysis

The average nitrogen (N) content of proteins was found to be about 16 percent, which led to use of the calculation N x 6.25 (1/0.16 = 6.25) to convert nitrogen content into protein content. (Mariotti et al., 2008)

Protein (%) =
$$N$$
 (%) x factor 6.25

The protein yield (kg ha⁻¹) was obtained by the following formula:

Statistical analysis

The data recorded during the course of investigation was subjected to statistical analysis by "Analysis of variance technique". The significant and non-significant treatment effects were judged with the help of 'F' (variance ratio) table. The significant differences between the means were tested against the critical difference at 5% probability level (**Chandel**, 1998).

Result and Discussion

Growth Parameters

"The data revealed that maximum plant height at 60 DAS was found with the application of 100% RDF which was statistically at par with 75% RDF and significantly higher than 50% RDF and control treatment. The results of present investigation are also in agreement with the findings of" (**Fatima** *et al.*, 2008). "At 60 DAS and at 90 DAS found maximum plant dry matter accumulation with the application of 100% RDF which was significantly higher than 50% RDF and control treatment". (**Jat and Ahalawat, 2004**)

Table 2. Effect of fertility levels and Biofertilizers on plant population, plant height and dry matter accumulation

Treatments	•	Plant population (running meter)		Plant height(cm)			Dry matter accumulation (gram/plant)		
	Initial	Harves	30DA	60	Harves	30	60DAS	90 DAS	
		t	S	DAS	t	DAS			
Fertility levels		I.							
F1	16.26	16.01	8.50	47.91	50.92	2.40	14.04	20.22	
F2	16.76	16.55	10.49	58.36	61.99	3.01	17.57	25.31	
F3	16.63	16.48	9.95	55.94	58.92	2.86	16.68	24.02	
F4	16.58	16.41	9.41	52.83	55.93	2.70	15.76	22.71	
SEm±	0.28	0.29	0.19	0.69	1.09	0.07	0.27	0.38	
CD (P= 0.05)	NS	NS	0.68	2.39	3.77	0.25	0.96	1.31	
Biofertilizers				·					
B1	16.57	16.45	9.59	53.87	56.99	2.75	16.08	23.16	
B2	16.38	16.13	9.10	50.84	53.96	2.58	15.07	21.71	
В3	16.72	16.51	10.07	56.56	59.86	2.89	16.88	24.32	
SEm±	0.27	0.26	0.18	0.89	0.95	0.07	0.25	0.32	
CD (P= 0.05)	NS	NS	0.53	2.69	2.85	0.25	0.75	0.96	
FXB	NS	NS	NS	NS	NS	NS	NS	NS	

Where F1-Control, F2- RDF 100%, F3- RDF 75%, F4- RDF 50%, B1- Rhizobium + PSB, B2- Rhizobium + PGPR, B3 - Rhizobium + PSB + PGPR

Root Parameters

All the root parameters i.e. root dry weight/plant, number of nodules per plant and nodule dry weight/plant influenced significantly due to different Fertility levels at all the stages of crop growth except root dry weight/plant at 30 DAS. However, higher values of root parameters i.e. root dry weight/plant, number of nodules per plant and nodule dry weight/plant with the application of 100% RDF which was statistically at par with 75 % and 50% RDF and significantly higher than control treatment. "Favourable effect on plant growth with different nutrient levels over control treatment may be attributed to better nutrient availability and number of metabolic processes taking place in the plant body, which in turn are affected by a variety of inherent and environmental factors to which plant is exposed that results more root dry weight, number of nodules per plant and nodule dry weight/plant" (Gray and Bahar, 2013; Egamberdieva et al., 2015). Biofertilizers found significant effect on all the root parameters at all the stages of crop growth except root dry weight/plant at 30 DAS. Biofertilizers treatments resulted higher values of root parameters in chickpea with the application of Rhizobium + PSB + PGPR (B₃) followed by 75% RDF (F₃) treatment. "The probable reasons for such results could be the growth promoting substances secreted by the microbial inoculants, which in turn might have led to better root development, better transpiration of water and enhanced uptake of nutrients that results more root dry weight/plant, number of nodules per plant and nodule dry weight/plant. These results were in accordance with works of" (Triphati et al., 2015; Singh and Prasd, 2008 and Gupta, 2004).

Table 3. Effect of fertility levels and biofertilizers on Root parameters of chickpea

	Root dry weight			Number of nodules			Nodules dry weight		
Treatments	(gram/plant)		/plant			(mg/plant)			
	30DA	60	Harvest	30	60	90	30	60DA	90
	S	DAS		DAS	DAS	DAS	DAS	S	DAS
Fertility levels	Fertility levels							1	
F1	0.17	0.56	0.63	12.26	15.43	17.50	15.71	38.86	40.73
F2	0.21	0.70	0.78	15.16	19.08	21.65	19.07	47.19	49.46
F3	0.20	0.65	0.73	14.13	17.77	20.16	17.77	43.95	46.07
F4	0.19	0.62	0.69	13.42	16.89	19.16	16.88	41.76	43.77
SEm±	0.01	0.01	0.02	0.30	0.39	0.39	0.36	0.75	0.91

CD (P= 0.05)	NS	0.05	0.06	1.06	1.35	1.36	1.24	2.60	3.16
Biofertilizers									
B1	0.19	0.63	0.71	13.79	17.35	19.69	17.35	42.92	44.99
B2	0.18	0.59	0.66	12.95	16.26	18.45	16.46	40.72	42.69
В3	0.20	0.67	0.75	14.52	18.27	20.18	18.26	45.18	47.35
SEm±	0.008	0.02	0.02	0.25	0.34	0.33	0.32	0.64	0.84
CD (P= 0.05)	NS	0.05	0.06	0.77	1.02	1.02	0.94	1.94	2.51
FXB	0.19	0.63	0.71	13.79	17.35	19.69	17.35	42.92	44.99

Where F1-Control, F2- RDF 100%, F3- RDF 75%, F4- RDF 50%, B1- Rhizobium + PSB , B2- Rhizobium + PGPR , B3 - Rhizobium + PSB + PGPR

Quality Traits

However, maximum protein content was observed with the application of control treatment and lowest protein content was found with the application of 100% RDF. The increase in fertility levels that results higher nitrogen content in 100% fertilized plot which ultimately results low protein content in seeds (Singh et al., 2004). Maximum protein content in seed was recorded with the application of B3 treatment and lowest protein content was found B2 treatment. Application of biofertilizers increase the protein content in seeds because biofertilizers enhance the nutrient uptake and plant use nutrients rapidly and efficiently that results more protein content in seeds (Singh and Prasad 2008). Protein yield influenced significantly by different fertility levels. Maximum protein yield was recorded with the application of 100% RDF which was statistically at par with 75% RDF and 50% RDF but significantly higher than control treatment. Increasing the seed yield increased the protein yield. These results are in tune with (Meena et al., 2005). Protein yield influenced significantly by different biofertilizers treatments. However, maximum protein yield was recorded with the application of B3 treatment which was significantly higher than other treatments. Increasing the seed yield increased the protein yield.

Table 4. Effect of fertility levels and biofertilizers on quality of chickpea

Quality parameter							
Treatment	Treatment Protein Content (%) Protein yield (q/ha)						
Fertility levels							
F ₁	21.22	427.63					

F2	23.11	572.31					
F ₃	22.10	523.34					
F4	21.87	485.17					
SEm ±	0.051	11.71					
CD (P = 0.05)	0.17	40.42					
Biofertilizers	Biofertilizers						
B1	21.81	500.68					
B2	21.67	462.74					
В3	22.70	542.91					
SE± m	0.039	8.98					
CD (P = 0.05)	0.117	26.94					
FXB	NS	NS					

Where F1-Control, F2- RDF 100%, F3- RDF 75%, F4- RDF 50%,B1- Rhizobium + PSB ,B2- Rhizobium + PGPR ,B3 - Rhizobium + PSB + PGPR

Conclusion

Based on the finding of the present study, it can be inferred that application of 100% RDF (F_2) with Rhizobium + PSB + PGPR (B_3) resulted maximum growth and root parameters which results ultimate more protein yield of late sown chickpea crop during rabi season in central zone of Uttar Pradesh.

References

Ali, H., Khan, M. A., & Randhawa, S. A. (2004). Interactive effect of seed inoculation and phosphorus application on growth and yield of chickpea (Cicer arietinum L.). *International journal of Agriculture and Biology*, 6(1), 110-112.

Akhtar, M., & Siddiqui, Z. (2009). Effects of phosphate solubilizing microorganisms and Rhizobium sp. on the growth, nodulation, yield and root-rot disease complex of chickpea under field condition. *African Journal of Biotechnology*, 8(15).

Ali, A., M.S. Zia, Rahmatullah, A. Shah and M. Yasin (1998). Nodulation in Sesbania bispinosa as affected by nitrogen application. Pakistan Journal of Soil Science 15: 183-185.

Bouyoucos, **G.J.** (1927). The hydrometer as a new method for the mechanical analysis of soil. *Soil Sci.* 23343-353.

Chandel SRS. (1998) Advance agriculture statics, 2nd Edition, Kalyani Publication, and New Delhi 1998.

Das, S., Pareek, N., Raverkar, K. P., Chandra, R., & Kaustav, A. (2012). Effectiveness of micronutrient application and Rhizobium inoculation on growth and yield of Chickpea. *International Journal of Agriculture, Environment and Biotechnology*, 5(4), 445-452.

Egamberdieva, D., Abdiev, A., Khaitov, B., (2015). Synergistic interactions among root-associated bacteria, rhizobia and chickpea under stress conditions. In: Plant Environment Interaction: Responses and Approaches to Mitigate Stress, M.M. Azooz, P. Ahmad (Eds.), John Wiley & Sons, Ltd., pp.250-261.

FAOSTAT (2018), Food and agriculture organization of the United Nations, 2017-2018

Fatima, Z., Bano, A., Sial, R., Aslam, M., (2008). Response of chickpea to plant growth regulators on nitrogen fixation and yield. *Pakistan Journal of Botany* 40(5): 2005-2013.

Gray, N., Bahar, N., (2013). Role of arbuscular mycorrhizal symbiosis in proline biosynthesis and metabolism of *Cicer arietinum* L. (chickpea) genotypes under salt stress. *Journal of Plant Growth Regulation* 32: 767–778.

Gupta, **S.C** (2004). Response of gram (Cicer arietinum L.) to types and methods of microbial inoculation. Indian J. of Agric. Sci. 74 (2): 73-75.

Islam M, Mohsan S, Ali S, Khalid R & Afzal S (2012). Response of chickpea to various levels of phosphorus and sulphur under rainfed conditions in Pakistan. *Romanian Agric Res.* **29**: 175-183.

Jackson, M. L. (1973). Soil Chemical Analysis, *Prentice Hall of India Private Limited*, *New Delhi*.

Jat, R. S. and Ahlawat, I. P. S. (2004). Effect of vermicompost, biofertilizer and phosphorus on growth, yield and nutrient uptake by gram (*Cicer arietinum* L.) and their residual effect on fodder maize (*Zea mays* L.). *Indian Journal of Agricultural Sciences* 74 (7): 359-361.

Jukanti, A.K., Gaur, P.M., Gowda, C.L.L. and Chibbar, R.N. (2012). Nutritional quality and health benefits of chickpea (*Cicer arietinum* L.). *A Review British J. Nutr.* 108:S11-S26.

Kumar, S., Tripathi, D.K., Bharose, R., Kumar, M. and Kumar, R. (2016). Effect of different fertility level and micronutrients on nodulation and nutrient uptake by chickpea. *An Asian Journal of Soil Sci.*11 (1): 62-66.

Meena KN, Pareek RG, Jat RS (2005). Effect of phosphorus and biofertilizers on yield and quality of chickpea. An. Agric. Res. New Series Vol. 2005; 22(3):388-390.

Mariotti, F., Tomé, D., & Mirand, P. P. (2008). Converting nitrogen into protein—beyond 6.25 and Jones' factors. *Critical reviews in food science and nutrition*, 48(2), 177-184.

Olsen, S.R., Cole, C.V., Watnahe, F.S. and Dean, L. A. (1954). Estimation of available phosphorous in soil by extraction with sodium bicarbonate U.S. Dept. *Agr. Cric.* 939.

Pingoliya, K, K., Dotaniya, M.L., Mathur, A, K. (2013). Role of phosphorus and iron in chickpea (Cicer arietinum L.). *Lap Lambert Academic Publisher, Germany*.

Rehan, W., Jan, A., Liaqat, W., Jan, F.M., Ahmadzai, M.D., Ahmad, H., Haroon, J., Anjum, M.M. and Ali, N. (2018). Effect of phosphorous, rhizobium inoculation and residue types on chickpea productivity. *Pure Appl. Biol.*, **7**(4): 1203-1213.

Rudresh D L, Shivaprakasha M K, Prasad R D (2005) Effect of combined application of *Rhizobium*, phosphate solubilizing bacterium and Trichoderma spp. on growth, nutrient uptake and yield of chickpea (*Cicer aritenium* L.). *App Soil Ecol* 28: 139-46.

Singh R, Prasad K. (2008) Effect of vermicompost, Rhizobium and DAP on growth, yield and nutrient uptake by chickpea. J. Food legumes. 2008; 21(2):112-114.

Toth, S. J. and Prince, A. L. (1949). Estimation of cation exchange capacity and exchangeable Ca K and Na Content of Soil by Flame photometer technique. *Soil Sci.* 67 439-445.

Tripathi, L.K., Thomas, T., Singh, V.J., Gampala, S., Kumar, R., (2015). Effect of nitrogen and phosphorus application on soil nutrient balance in chickpea (*Cicer arietinum* L.) cultivation.

Singh, S., Saini, S.S. and Singh, B.P. (2004). Effect of irrigation, sulphur and seed inoculation on growth, yield and sulphur uptake of chickpea (*Cicer arietinum*) under late sown conditions. *Indian J. Agron.*, 49 (1): 57-59.

Subbiah, B. V. and Asija, C. L. (1956). A rapid procedure for the estimation of available nitrogen in soil, *Current Sci.* 25. 259-260.

Walkley, A. (1947). Critical examination of rapid method for determining organic carbon in soil, effect of variation in digestion condition and of inorganic soil constitutes. *Soil Sci.* 632-251.