

## Original Research Article

# Comparative Repellency of Selected Plants to Adult *Anopheles gambiae* complex (Gilles) in the Laboratory

### ABSTRACT

**Aims:** The study evaluated the repellency effects of some tropical plants and shrubs found in semi-rural communities of Badagry Area of Lagos state; which are acclaimed to have the potentials of repelling mosquitoes from human dwellings. The repellency effects of *Moringa oleifera*, *Morinda lucida*, *Magnifera indica* and *Phyllanthus muellerianus* to adult *Anopheles gambiae* was evaluated in the Laboratory.

**Study Design:** The study was carried out at Central Research Laboratory of Lagos State University, Ojo, Lagos, Nigeria and Central Research Laboratory of University of Lagos, Akoka, Lagos, Nigeria. Powdered of dried test plants were prepared and admix with coconut husk as inert, different concentrations were rubbed on the forearm of volunteers and repellency to blood starved female *Anopheles* mosquitoes was observed.

**Methodology:** Test plants were collected from Badagry area of Lagos State, they were identified at University of Lagos Herbarium and given numbers. They were dried between 10 and 14 days at temperature of 25-27°C and powdered. Different concentrations of the powder mixed with powdered coconut husk was used to treat volunteers forearms and they were exposed to 0-2 two day old adult unfed mosquitoes in an aluminum glass cage fitted with net as arm entrance and repellency was observed for a period of 180 minutes, with landing counts taken every 30 minutes. The test plants were also subjected to qualitative and quantitative phytochemical analysis at University of Lagos Central Research Laboratory.

**Results:** Results showed that all test plants were able to repel *Anopheles* mosquitoes in the study, repellency was shown in descending order *Moringa oleifera* with 88%, *Magnifera indica* 83%, *Phyllanthus muellerianuss* 80% and *Morinda lucida* 72%. There was no statistical significance in percentage repellency at 95% CL. The result of phytochemical screening of the test plants showed that only *M. indica* indicated presence of saponing (36.99%). While *M.oleifera* has highest phenol content (45.6%3), Alkaloid (38.68%), steroid (24.89%) and Tannin (33.19%). Flavonoid and reducing sugar quantity was highest in *M. indica* (39.39%) and (55.18%) respectively.

**Conclusion:** The plants were able to show repellency to *Anopheles gambiae* a nuisance malaria vector of serious medical importance. These plants are available in all tropical areas of Africa, they can therefore be used to prevent nuisance and painful mosquito bites which could be a sustainable way to prevent mosquito vectored diseases.

**Keywords:** *Anopheles gambiae*, *Moringa oleifera*, *Morinda lucida*, *Magnifera indica*, *Phyllanthus muellrianus* Repellency Badagry Phytochemical flavonoid phenol tannin alkaloid steroid saponin

## 1. INTRODUCTION

Mosquitoes are the most important single group of insects in terms of public health importance because they are vectors of serious human diseases such as malaria, filariasis, Japanese encephalitis, dengue fever, and yellow fever. Mosquito-transmitted diseases are a major cause of illness and death in the world, particularly in tropical and subtropical Africa [1]. They are considered the deadliest insects in the world and are major human and animal health problem as they transmit diseases to more than 700 million people annually [2,3]. World Health Organization [4] estimated that more than a million people die annually from malaria and other mosquito borne diseases with 300 to 500 million people being affected. It is also estimated that 100 children under one year of age and 200 children under five years out of 1000 respectively die annually of malaria in Nigeria [5].

Mosquito control in Africa and its environs is becoming a great challenge because oftentimes synthetic chemical insecticides use for it control are harmful to man and animals, non-degradable and non-selective and are considered to be environmental pollutants and bio accumulates both in the body and build up in the food chain has discouraged their use and hence some of them have been banned in developed Countries [6, 7].

The problems associated with the use of chemical insecticides have challenged researchers to explore traditional non chemical control methods that were practiced before the advent of chemical insecticides. Such as mosquito avoidance practices like burning of plant parts, rubbing of ash and oils to ward off mosquitoes and consequently avoid bites. Plant botanicals have been used from time immemorial because they are cheap, safe and effective means of mosquito vectors control [8].

The aim of the research is to evaluate the repellency effect of four tropical plants namely: *Moringa oleifera*, *Magnifera indica*, *Morinda lucida* and *Phyllanthus muellerianus* against *Anopheles gambiae*.

## 2. MATERIAL AND METHODS

### 2.1 COLLECTION AND PREPARATION OF TEST PLANTS

Matured whole plants of *Moringa oleifera*, *Magnifera indica*, *Phyllanthus mullerianuis*, and *Morinda lucida* were collected from local villages around Badagry Area of Lagos state.

The Plants were taken to University of Lagos herbarium for identification and deposit.

Respective voucher numbers were given: *Magnifera indica* (8591), *Phyllanthus mullerianuis* (8590), *Morinda lucida* (8593) and *Moringa oleifera* (8589). The collected plants were washed under running water and transported to the laboratory and dried on the laboratory work tables at  $26 \pm 2^{\circ}\text{C}$  for about 10 days until fully dried. The dried leaves were grinded into fine powder with JŘ 200<sup>R</sup> grinding machine. Each powder form of plant leaves was kept separately in an airtight containers and stored in refrigerator until needed.

The stored leaf powders were admixed with powdered coconut husk as inert material at different concentrations. The repellent study methodology of World Health Organization (1996) arm in cage testing method was adopted with slight modifications. The repellent study was carried out in the laboratory using 40 by 40 netted aluminum cages with arm entry; twenty 0-2 day old starved female *Anopheles gambiae* mosquitoes were introduced into the cages. The volunteers hands were washed with mild soap rinsed with water and rubbed with plant powder that had been mixed with water to form a paste and the arm was put in the cage that houses the mosquitoes. The powder paste was applied at 10%, 20%, 30% to 100%. 1g of each concentration was used to rub the exposed area of the fore arm. Only powder coconut husk served as control. The control and treated arms were introduced simultaneously into the mosquito cage, and gentle tapping of the sides of the experimental cages is to activate the mosquitoes. Each test concentration was repeated three times. The volunteers conducted their test of each concentration by inserting the treated and control arms in to the same cage every 30 minutes for 1800 minutes. The mosquitoes that landed and bites received on the hands were recorded after every 30 minutes and then shaken off making out a 180 minute protection. The percentage of repellency was calculated with the formula: % Repellency =  $[(T_a - T_b)/T_a] \times 100$ , where  $T_a$  is the number of mosquitoes in the control group and  $T_b$  is the number of mosquitoes in the treated group.

## 2.2 PREPARATION OF PLANTS FOR PHYTOCHEMICAL ANALYSIS

### 2.2.1 Qualitative Tests

#### **2.2.1.1 Test for Tannins**

About 0.5 g of the sample was boiled in 20 ml of water in a test tube and then filtered. A few drops of 0.1% ferric chloride was added and observed for brownish green or a blue-black colouration.

#### **2.2.1.2 Test for Phlobatannins**

0.5g extract of each plant sample was boiled with 2mL of 1% aqueous hydrochloric acid for 10 minutes. Deposition of a red precipitate indicates the presence of phlobatannin.

#### **2.2.1.3 Test for Saponin**

About 2 g of the sample was boiled in 20 ml of distilled water in a water bath and filtered. 10ml of the filtrate was mixed with 5 ml of distilled water and shaken vigorously for a stable persistent froth. The frothing was mixed with 3 drops of olive oil and shaken vigorously, formation of emulsion was observed.

#### **2.2.1.4 Test for Flavonoids**

5 ml of dilute ammonia solution were added to a portion of the aqueous filtrate of each plant extract followed by addition of concentrated tetra oxosulphate  $\text{VI}$  acid.

#### **2.2.1.5 Test for Steroids**

2 ml of acetic anhydride was added to 0.5 g ethanolic extract of each sample with 2 ml tetra oxosulphate  $\text{VI}$  acid. The colour changed from violet to blue or green in some samples indicating the presence of steroids.

#### **2.2.1.6 Test for Terpenoids (Salkowski test)**

5 ml of each extract was mixed in 2 ml of chloroform, and concentrated sulfuric acid (3 ml) was carefully added to form a layer. A reddish brown colouration of the inter face was formed to show positive results for the presence of terpenoids.

#### **2.2.1.7 Test for Cardiac Glycosides (Keller-Killani test)**

Five ml of each extracts was treated with 2 ml of glacial acetic acid containing one drop of ferric chloride solution. This was underlayered with 1 ml of concentrated sulphuric acid. A brown ring of the interface indicates a deoxysugar characteristic of cardenolides.

#### **2.2.1.8 Test for Alkaloid**

A 5 mg sample of the extract dissolved in 3 ml of acidified ethanol was warmed slightly and then filtered. Few drops of Mayer's reagent and 1 ml of Dragendorff's reagent were added to 1 ml of the filtrate and turbidity was observed

### **2.2.2 Quantitative Analysis Tests**

#### **2.2.2.1 Estimation of Tannins**

500 mg sample of the concentrate was dissolved in 50 ml of distilled water, and shake for one hour. A 5 ml aliquot of the filtrate was mixed with 2 ml of 0.1 M  $\text{FeCl}_3$  in 0.1 N HCl and 0.008 M potassium ferrocyanide. The absorbance was measured at 720 nm within 10 minutes.

#### **2.2.2.2 Estimation of Total Phenolic Compound**

0.5g sample of extract was dissolved in 50 ml of water. 0.5 ml was added to 0.1 ml of Folin-Ciocalteu reagent (0.5 N) mix and incubate at room temperature for 15 minutes. After this, add 2.5 ml sodium carbonate solution (7.5% w/v) and further incubated for 30 minutes at room temperature. The absorbance of the solution was measured at 760 nm. The concentration of total phenol was expressed as gallic acid equivalent (GAE) (mg/g of dry mass) which is a commonly used reference value.

#### **2.2.2.3 Total Flavonoid Content Estimation**

1 ml of sample solution (100 $\mu\text{g}$ / ml) was mixed with 3 ml of methanol, 0.2 ml of 10% Aluminum chloride, 0.2 ml of 1 M potassium acetate and 5.6 ml of distilled water. The resulting mixture was incubated at room temperature for 30 minutes and the absorbance of the reaction mixture was measured at 415 nm. The calibration curve was prepared by preparing quercetin solutions at various concentrations in methanol.

#### **2.2.2.4 Total Antioxidant Capacity Determination**

solution of the samples extract (1ml) was mixed with 3 ml of reagent solution (0.6 M sulphuric acid, 28 mM sodium phosphate and 4 mM ammonium molybdate). The tubes were capped and incubated in a boiling water bath at 95 °C for 90 minutes. After the samples had cooled to room temperature, the absorbance of the aqueous solution of each was measured at 695 nm. The total antioxidant capacity was expressed as equivalent of ascorbic

#### **2.2.2.5 DPPH Radical Scavenging Activity Assay**

An aliquot of 0.5 ml of extract in ethanol (95%) at different concentrations (25, 50, 75, 100 µg/ml) was mixed with 2.0 ml of reagent solution (0.004 g of DPPH in 100 ml methanol). The control contained only DPPH solution in place of the sample while methanol was used as the blank. The mixture was vigorously shaken and left to stand at room temperature. After 30 minutes the decrease in absorbance of test mixture (due to quenching of DPPH free radicals) was read at 517 nm. The scavenging effect was calculated using the expression  $\% \text{ inhibition} = [A_0 - A_1] \times 100 / A_0$ . Where  $A_0$  is the absorption of the blank sample and  $A_1$  is the absorption of the extract

#### **2.2.2.6 In vitro Lipid Peroxidation Assay**

The reaction mixture containing liver homogenate (0.2ml), Tris-HCl buffer (20 mM pH 7.0, 0.1ml),  $\text{FeCl}_2$  (2 mM, 0.1ml), ascorbic acid (10 mM, 0.1 ml), and 0.5 ml plant extract (25–100 µg/ml) in a final volume of 1 ml. The reaction mixture was incubated at 37 °C for 1 hour. Lipid peroxidation was measured as malondialdehyde (MDA) using trichloro acetic acid (TCA), thiobarbituric acid (TBA) and HCl (TBA-TCA reagent: 0.375 % w/v TBA; 15 % w/v TCA and 0.25 N HCl). The incubated reaction mixture was mixed with 2 mL of TBA-TCA reagent and heated in a boiling water bath for 15 minutes. After cooling, the flocculent precipitate was removed by centrifugation at 10,000 g for 5 minutes. Finally, malondialdehyde concentration in the supernatant fraction was determined spectrophotometrically at 535 nm. Ascorbic acid was used as standard.

#### **2.2.2.7 Nitric Oxide Scavenging Activity Assay**

4 ml sample of plant extract or standard solution of different concentrations (25, 50, 75, 100 µg/ml) were taken in different test tubes and 1 ml of Sodium nitroprusside (5 mM in phosphate buffered saline) solution was added into the test tubes. They were incubated for 2 hours at 30 °C to complete the reaction. A 2 ml sample was withdrawn from the mixture and mixed with 1.2 ml of Griess reagent (1% Sulphanilamide, 0.1% naphthylethylenediaminedihydrochloride in 2% H<sub>3</sub>PO<sub>4</sub>). The absorbance of the chromophore formed during diazotization of nitrite with sulphanilamide and its subsequent coupling with naphthylethylenediamine was measured at 550 nm [9]. Ascorbic acid was used as standard. The percentage (%) inhibition activity was calculated from the following equation:

$$[(A_0 - A_1)/A_0] \times 100.$$

Where, A<sub>0</sub> is the absorbance of the Control and A<sub>1</sub> is the absorbance of the extract or standard

#### 2.2.2.8 Reducing power assay

Various concentrations of the extracts (20 to 100 µg/ml) in 1.0 ml of deionized water were mixed with phosphate buffer (2.5 ml) and potassium ferricyanide (2.5 ml). The mixture was incubated at 50°C for 20 min. Aliquots of trichloroacetic acid (2.5 ml) were added to the mixture, which was then centrifuged at 3000 rpm for 10 min. the upper layer of solution (2.5 ml) was mixed with distilled water (2.5 ml) and a freshly prepared ferric chloride solution (0.5 ml). The absorbance was measured at 700 nm. A blank was prepared without adding extract. Ascorbic acid at various concentrations (1 to 16 µg/ml) was used as standard.

### 3. RESULTS

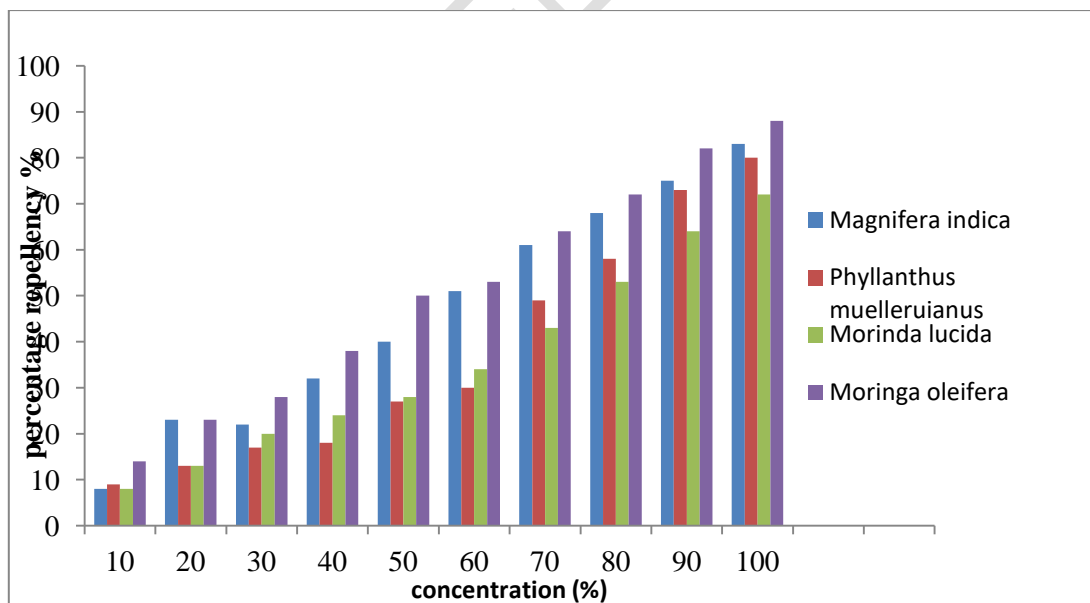
The result showed that repellency is dose dependent; *M. oleifera* showed highest percentage of repellency of 88% at for 100% powder while there was no repellency observed in controls as all starved mosquitoes became activated and landed on the untreated forearm. *Morinda lucida* also showed repellency of 86% at 100% concentration, *Magnifera indica* had repellency of 83% while repellency of 80% was shown by *Phythallatus muellerianus* 100% concentration which is the least. However, there was no statistical significance among the four test plants (<0.05).

**Table 1. Quantitative Phytochemical Screening Of Test Plants**

	Saponin	Phenol	Flavonoid	Alkaloid	Steroid	Reducing Sugar	Tannin
<i>Moringa.oleifera</i>		45.63	31.02	38.63	24.89	43.09	33.19
<i>Mag indica</i>	36.99	39.39	22.53	30.05	14.89	55.18	28.65
<i>Morinda. lucida</i>		27.01	33.47	24.51	19.98	39.99	19.65
<i>P.muellerianus</i>		29.66	57.31	26.28	23.77	45.27	21.58

**Table 2. Qualitative Phytochemical Screening Of Test Plants**

Sample	Saponin	Phenol	Flavonoid	Tannin	Alkaloid	Reducing Sugar	Phlobatannin	Steroid	Terpenoid
<i>Moring oleifera</i>	-	+	+	+	+	+	-	+	+
<i>Mag. indica</i>	+	+	+	+	+	+	-	+	+
<i>Morinda lucida</i>	-	+	+	+	+	+	-	+	+
<i>P.muellerianus</i>	-	+	+	+	+	+	-	+	+



**Fig. 1. Chart of Plants showing comparative percentage repellency against concentration.**

#### **4. DISCUSSION AND CONCLUSION**



The present research evaluated repellency effect of *Moringa oleifera*, *Magnifera indica*, *Morinda lucida* and *Phyllanthus muellerianus* to adult *Anopheles gambiae* in the Laboratory. The result of the research showed that all plants are potent and were able to repel mosquitoes at concentration 0.6 to 1.0g where over 50% population of exposed insects were repelled. *Moringa oleifera* showed highest repellency to *Anopheles gambiae* with repellency of 88% at 1.0g. *Magnifera indica* conferring 83% repellency, *Phyllanthus muellerianus* conferring 80% and *Morinda lucida* conferring 72% repellency on volunteer after 60 minutes of exposure.

The result equally showed that leaf powder of *Moringa oleifera* is highly repellent to *Anopheles gambiae* adult and it shows a very strong indication of its potential as a repellent. The result from the study is in agreement with Prabhu *et al* [11] that evaluated the repellent potential of *M. oleifera* against adult mosquito; the result showed that repellency is dose dependent, 90.41% repellency was achieved at 100% concentration; while 23% was recorded at 20% concentration. The use of plants to ward off nuisance insects is an age long practice. The phytochemical analysis equally showed that *M. oleifera* exhibited highest presence of phenol, alkaloid, steroid and tannin. These phytochemical may be the compound that is making it show higher repellency. However, there is need to authenticate this observation.

The result justifies studies on botanicals against insects pests as shown by several authors; and not just adherence to traditional beliefs of hanging plants on door and bed post. The insecticidal properties of 10 mosquito repellants plants were well documented; as shown by Egunyomi *et al.* [10]. The methanol and hexane extracts of the plants were investigated for phytochemical compounds with repellent activities against *A.stephensi*. The test plants contained phenols and stereroids. The active extracts are promising ethnobotanical repellent at 2mg/ml against *A. stephensi* and could be sources of new natural repellent compounds.

The plants produce certain compounds that repel and ward off insects pests from attacking them and these has provide a basis of use of the plants as repellents. Most branded

repellents are made from harmful chemicals; and there has been reports of undesirable side effects to their use; also some of them are non-effective but a way to make consumers part with their hard earned money.

This result agrees with work of Denloye et al. [8], who reported that some plants such as Citrus species, whose peels were dried and burnt to repel mosquitoes in Nigeria and have been found to be toxic to some other insects. These plants therefore can serve as very potent replacement for the chemical insecticides used in mosquito coils and other repellents, thereby reducing the burden of respiratory disorders.

### **Consent:**

Ten volunteers that gave consent were used for the study.

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