Phytochemcial Assessment of *Persea americana* Powdered Leaves and Its Potency in Protecting *Clarias gariepinus* against *Klebsiella pneumonia*

1. ABSTRACT

The phytochemcial content of P.. americana powdered leaves was assessed using ethanol extracts. The potency of the powdered leaves was assessed by exposing C.gariepinus fed different levels of the dietary powdered leaves (Do: 0% leaves inclusion; D1: 3% leaves inclusion; D2: 6% leaves inclusion; D3: 9% leaves inclusion and D4: 12% leaves inclusion diets) for eight weeks to K. pneumonia via intrapretoneal injection. The fish were injected after the 8 weeks feeding at day 1, 7, 14 and 21. The haematological parameters and microbial load (antibacterial activities) of the infected fish blood and organs were assessed 28 days after the first injection. The ethanol extracts of the P. americana leaves powder revealed the following phytochemicals: phenol, carotenoids, alkaloids, flavonoids, saponin, steroids, coumarine and proanthocyanidin in varying quantities. The haematological parameters in the infected fish compared to the control Do indicated that there were improved Pack cell volume (PCV), Haemoglobin (Hb) and Red blood cells (RBC) in fish fed D2-D4 compared to the fish fed Do (control). The white blood cells (WBC), the lymphocytes, Monocytes and Eosinophils were higher (P>0.05) in the fish fed Do compared to the fish fed D1-D4. The Red blood indices (MCH, MCHC and MCV) fluctuated in values across the diets, but were higher in fish fed D1. The microbial load was higher (P<0.05) in the organs (Kidney, spleen, liver and Blood) of the fish fed Do compared to the rest, and the effect was dose dependent. The results showed that P. americana powdered leaves improved the haematological and antibacterial activities of the experimental fish.

Keywords: Acquaculture, fish health, *Persea americana*, *Klebisiella pneumonia*, *C. gariepinus*

2. INTRODUCTION

Aquaculture have contributed to the protein availability for human consumption despite age, class or sex [1]. Fish and fish products emanating from aquaculture practice surpasses that of the marine capture fisheries with over 10,000,000 tonnes [2]. Essential vitamins, fatty acids, minerals, lipids etc are contained in aquacultural products, and if consumed appropriately prevents the occurrence of certain diseases such as cancers, eye defects, cardiovascular disorders etc [3]. Pollution of our environment have contributed to decline in the fish availability [4], and aquaculturist and fish farmers must adopt a practice of not only improving the growth rate of fish, but also its survival. [5] observed that the type of feed administered in aquaculture does not only determines the growth rate of fish but also the survival rate.

Herbs and herbal products have contributed to the growth and immunostimulant enhancement in aquaculture [6]. Some of the herbal products that have contributed to aquaculture practice includes: Avacado pear leaves aqueous extracts [7] Moringa leaves [8] and *Carica papaya* root extracts [9].

Clarias gariepinus is an economically important freshwater fish with outstanding qualities such as fast growth, diseases resistance, good flesh qualities among others [10].

Avocado pear (*P. americana*) leaves plants have been reported to be medicinal [11]. The introduction of avocado pear leaves in aquaculture practice has led to improvement in growth [12] and immunostimulants enhancement [13].

The purpose of this research work is to assess the medicinal phytochemical in the ethanolic extracts of the *P. americana* powdered leaves and it's importance in the health of *Clarias* gariepinus.

3. MATERIALS AND METHODS

3.1 Experimental fish

The experimental fish was procured in Idi-onyana farms along Abua – Ahoada road in Abua/Odual Local Government Area of Rivers State.

3.2 Experimental diets

Persea americana leaves were harvest in Port Harcourt, Rivers State. They were air dried to constant weight, grounded to it's powdered form, sieved and stored. The sieved powdered leaves were added to 38.35 crude protein formulated diet (Do) at 3% (D1), 6%(D2), 9% (D3) and 12% (D4) respectively following the method of [14].

3.3 Phytochemistry of Powdered *P. americana* leaf ethanol extracts

Quantitative analysis of powdered *P. americana* leaves was carried out in the Department of Chemistry, Faculty of Science, Rivers State University, in accordance with ISO 179245 using UV 250 visible.

3.4 Experimental procedure

150 catfish of 117.8± 1.11g and 25.88± 0.14cm weight and length respectively were stocked in the experimental tanks at 10 fish per tanks in triplicates, and feeding commenced 24 hours after stocking. The fish were fed the experimental diets according to the designated tanks (Do-D4) at 5% body weight per day, 2 times daily. After eight weeks of feeding, the fish were injected three consecutive times (at day 1, 7 and 14) intraperitoneally with 1.5ml of 1.9 x 10⁵cfu/ml of over night grown *K. pneumonia*. After 21 days from the first infection, blood samples were collected

for haematological analysis, and the organs such as blood, liver, kidney and spleen were collected for microbial (*K. pneumonia*) presence analysis.

3.5 Analysis of Plasma Haematological Parameters

Blood samples were collected from the experimental fish after day 28 days post infection via kidney puncture using an injection needle and 5ml syringe. Samples were taking to the laboratory and analysed for packed cell volume (PCV), Haemoglobin (Hb), Red blood cells count (RBC), Thrombocytes (TCT) and White blood cell (WBC) using auto haematological analyzer model MY-BOO2B, while white blood differential and red blood indices were analysed using the methods of [15].

3.6 Microbial Load (Anti-bacterial)

Approximately one gram of each organ was weighed independently and macerated in a universal container 10ml of normal saline was added to the macerated samples. 10ml of each diluted organs were cultured on TSA and inoculated at 37°C for 24 hours. The number of bacteria were determined by counting the colonies grown on the agar plate.

3.7 Data Analysis:

The collected data were analysed using SPSS statistics software 17.0 for windows. A one way analysis of variance (ANOVA) was employed to reveal significant difference between control and treated groups. Turkey's multiple comparison test was applied to separate treatments with significant difference [16].

4. RESULTS

Table 1 shows the phytochemical analysis of the powdered *P. Americana* leaves, using ethanol as solvent, it reveals the presence of Phenol, Carotenoids, Alkaloid, Flavonoid, Saponin, Steriod, Coumarine and Proanthocyaniden at varying quantities. The result of the haematological parameters is in Table 2, the PCV, Hb, RBC and Neut were lower in the control group compared to the treated group. Basophil was zero across the experimental groups, the TCT fluctuated among the groups, but higher in D1-D3, but the WBC, Lymp, Eos, Mono, MCH, MCHC and MCV were higher in the control group compare to the treated groups. The result for the microbial load (Tables 3) reveals, that *K. pneumonia* presence was higher in the organs of the control group compared to the treated groups, while Plates 1 – 3 shows *P. americana* leaves at the fresh dry and powdered leaves respectively.

Table 1: Quantitative screening for Phyto-chemical Components of ethanol extract of Dry Avocado Pear Powdered Leaves (Mean ±SD)

COMPONENT	Yield (mg/dl)
Phenol	43.1 ± 0.64
Carotenoids	0.851 ± 0.20
Alkaloid	0.50 ± 0.31
Flavonoid	3.60 ± 0.61
Saponin	1.30 ± 0.10
Tannin	0.00
Quinon	0.00
Steroid	1.31 ± 0.13
Triterpenoid	0.00
Coumarin	0.24 ± 0.12
Cyanogenic glycosides	0.00
Proanthocyanidin	0.93 ± 0.50
Phytocobillins	0.00
Pycocyanin	0.00

Data are mean ± Standard deviation of triplicate determination

Table 2: Haematological Parameters in Plasma of C. gariepinus fed Avocado Pear leaf Supplemented diets challenged with $Klebsiella\ pneumoniae\ (Mean \pm SD)$

Parameters	Diets				
	D0	D 1	D2	D3	D4
Packed Cell Volume (%) (PCV)	33.00±2.00 ^b	40.00±1.00 ^a	43.00±2.00 a	44.00±1.00°	44.00±1.00 a
Haemoglobin (g/dl) (Hb)	12.00±1.00 ^a	13.50±0.20 ^a	14.40±0.10 ^a	14.53±0.21 ^b	13.50±0.20 ^a
Red Blood Cell (cells x 10 ¹²) (RBC)	5.80 ± 0.10^{a}	4.60±0.10 ^b	6.47±0.15 ^a	6.47±0.15 ^a	6.00 ± 0.10^{c}
White Blood Cell (cells x 10 ⁹ /l) (WBC)	16.40±0.10 ^a	10.00±0.20 ^b	7.60±0.20 °	7.53±0.06 °	7.13±0.38 °
Neutrophils (%) (Neut)	27.00±1.00°	38.00±1.00 ^b	42.33±0.58 ^a	46.00±1.00 a	45.00±1.00 a
Lymphocytes (%) (Lymph)	60.00±1.00 ^a	52.33±2.52 ^b	48.67±1.15 °	50.00±1.00 ^b	48.00±1.00 °
Eosinophils (%) (Eos)	4.00 ± 1.00^{a}	4.00±1.00 a	2.67 ± 0.58^{b}	$2.87\pm1.00^{\ b}$	3.00 ± 1.00^{b}
Monocytes (%) (Mono)	10.00±1.00 ^a	6.67±0.58 ^b	7.33±0.58 ^b	5.00±1.00 b	5.33±0.58 ^b
Thrombocytes (%) (TCT)	215.00±1.00 ^b	217.00 ± 2.00^{b}	238.00±1.00°a	236.00±1.00 a	178.00±1.00°
Basophils (%) (Base)	0.00 ± 0.00^{a}	0.00 ± 0.00^{a}	$0.00\pm0.00^{\rm \ a}$	0.00 ± 0.00^{a}	0.00 ± 0.00^{a}
Mean Corpuscular Haemoglobin (pg)	36.52±4.72 ^a	33.77±1.19 ^a	33.04±0.32 a	33.04±0.32 a	33.77±1.19 ^a
(MCH)					
Mean Corpusc. Haemoglobin Conc. (g/dl)	26.08±1.97 ^a	23.28±0.66 ^a	22.27±0.38 a	22.48±0.22 a	22.50±0.04 ^a
(MCHC)					
Mean Corpuscular Volume (FL) (MCV)	71.73 ± 3.80^{a}	68.96±0.54 ^b	66.53±3.85 ^b	68.04±0.31 ^b	66.69±2.40 ^b

Means within the same roll with different superscript are significantly different (p<0.05)

Table 3: Microbial Load (cfu/ml) in Different Organs of Clarias gariepinus fed Avocado pear leaf Supplemented diets and infected with Klebsiella pneumoniae (Mean ±SD)

Organs	Diets							
	D0	D1	D2	D3	D4			
Kidney	56.33±7.57 ^a	52.67±2.08 ^a	43.67±1.52 ^b	37.00±4.58°	11.33±3.51 ^e			
Spleen	42.00±2.65 ^a	34.67±3.51 ^b	21.33±3.01°	15.00±2.65 ^d	4.00±1.00 ^e			
Liver	53.00±2.65 ^a	35.33±2.52 ^b	32.33±2.08 ^b	28.33±3.06 ^c	27.00±4.59°			
Blood	90.00±4.36 ^a	33.33±3.22 ^b	33.33±2.08 ^b	31.00±6.00 ^b	19.67±5.03°			

Means within the same roll with different superscript are significantly different (p<0.05)



Plate 1: Freshly harvested *P. Americana* leaves



Plate 2: P. Americana leaves air-dried to constant weight



Plate 3: P. Americana powdered leaves (grinded and filter)

5. Discussion

5.1 Phytochemicals of Avocado Pear (P. americana) Powered Leaf

The ethanol extracts of the *P. americana* powdered leaves showed the presence of the following phytochemicals in varying quantities: phenol, saponin, carotenoids, alkaloids, flavonoid, coumarin, steride and proanthocyanidin. This result is in agreement with the findings of [17] and [18] who reported similar phytochemicals in the ethanolic extract of powdered *P. ameriana* leaves, but slightly different from the result of [19] who reported the absence of coumarin and alkaloids in the ethanolic extract of the dry Avocado leaves powder, probably due to experimental procedure.

Phytochemicals have been reported to have inhibitory and eliminating effects on series of microorganisms that are life threatening to both humans, terrestrial and aquatic animals [20; 11]. The phytochemicals in powered *P. americana* leaves extracts are strong antioxidants that can prevent oxidative stress [21; 18] and modulate the metabolism of cholesterol in the body [22; 23], although saponins interrupt the sucking up of minerals and vitamins in the body [24], the extracts of Avocado pear leaves have been observed to increase plasma enzymes and total cholesterol [25]. There may be more valuable and health benefiting phytochemicals in the avocado pear powdered leaves that were not identified by this quantitative screening, since studies have shown that the presence of phytochemicals in any extraction process depends also on the type of solvent used [18] and that ethanol is less effective in screening phytochemicals [26]. Methanol and water are most efficient solvents for the extraction of antioxidants [27]. The benefit of applying the powdered leaves in this research work is that all the phytochemicals in the leaves were completely utilized.

5.2 Effect of *P. americana* powered leaves on Haemotology of *K. pneumonia* C. gariepinus

The presence of bacterial infection alters the haematological paramaters of fish, and reveals, the health status of fish [12; 28]. After infecting *C. gariepinus* with *K. pneumonia* in this research work, the result indicated that the values of the PCV, Hb and RBC increased in the fish fed D1-D4 compared to the fish fed D0 (control). This result is similar to the result obtained by [29] when *C. gariepinus* was experimentally challenged with *Escherichra coli* and *Vibrio fischeri*. The reduction in the PCV, Hb and RBC in the plasma of fish fed Do is an indication that the production and distribution of blood cells is negatively altered, and this could lead to low oxygen circulation in the fish [30]. The results agrees with the report of [31] who reported similar result when tilapia was naturally infected with *F. Colunaris* and [32] who reported similar result when *channa punctatus* was challenged with *A. hydrophila*. The result of these parameters in the fish D1-D4 (Supplemented diets) after the infection shows that there were improved antibody production which promotes better oxygen circulation and blood cells formation [33; 34].

The WBC, lymp, Neut and Eos increased in the plasma of the fish fed Do compared to the fish fed D1-D4, this is an indication that the fish fed Do is under pathogen induced stress [35; 36], since these parameters are responsible for the protection of the fish against pollution, intoxicants, pathogen and other infections [37; 38]. The lower values of the WBC, lymp, Neut, Eos could be as a result of the presence of phytochemicals such as alkaloids, flavonoids, phenol etc. that are present in the *P. americana* powdered leaves diets (D1-D4), which have been reported to be bactericidal [11]. It could also be that the supplemented diets (D1-D4) reduced the pathogenicity of the *K. pneumonia* by boosting the immune systems of the experimental fish [39]. The fish fed

D1-D3 had higher thrombocytes (TCT) values compared to fish fed Do after the infection with *K. pneumonia*. This result is in agreement with the result obtained by [28] when *C. gariepinus* fed dietary mango back was infected by *Pseudomonas geruginosa*. The improved TCT production in the fish fed D1-D4 could be as result of the phytochemicals in the supplemented diets (D1-D4) [40], the resultant effect could be the protection of fish against skin ulceration by bacterial infection [28; 41], since increase in thrombocytes prevents injuries and leads to quick recovery from external wounds [42].

Basophils were completely absent in all the experimental fish, this result is similar to the results reported by [31].

5.3 Antibacterial activities of *P. americana* powered leaves

There were reduction in microbial load of the experimental fish as the percentage of *P. aeruginosa* powdered leaves increases in diets (Do-D4). The microbial loads were higher in fish fed Do compared to fish fed D1-D4. This result is in agreement with the one reported by [43] who reported reduction in the microbial load of *Vibrio harveyi* infection when diets supplemented with herbs were administered to juvenile greasy groupers; and [44] who reported decrease in bacterial load in treated groups compared to the control when *C. gariepinus* fed various levels of onion bulb and walnut supplemented diets were exposed to *P. aeruginosa*. The reduction in the bacterial load in the fish fed D1-D4 could be as a result of the phytochemicals such as alkaloids, flavonoids, tannins, saponins present in the *P. americana* powdered leaves which have been reported to be immune-stimulant and anti-pathogenic [45; 11]. These phytochemical have also been reported to be bacteriostatic and bactericidal [46; 47].

Conclusion

The presence of *Klebsiella pneumonia* in our fish farms can cause serious economic loses as a result of fish mortality. The presence of this harmful pathogenic bacteria (*K. pneumonia*) can be detected via haematological analysis of the blood, determination of microbial load in the organs (Kidney, Spleen liver and blood and liver histopathology. *Persea american* powdered leaves has commendable quantities of Phenol Flavanoid, Carotenoids, Steriods etc that are antitbacterial, anti-inflammatory and immunostimulant.

The result of this work dietary *P. americana* powdered leaves is a good prophylactic to fresh water pathogens such as *Klebsiella pneumonia*.

References

- 1. Food and Agriculture Organization (FAO). (2016). State of World Fisheries and Aquaculture 2016. Contributing to Food Security and Nutrition for all. Vialedelle Terme di Caracalla, Rome, Italy: Food and Agriculture Organization
- 2. Food and Agriculture Organization (FAO), (2014). The state of the World Fisheries and Aquaculture, 2014. Food and Agriculture Organization of the United Nations, Vialedelle Terme di Caracalla. 00153 *Rome*, Italy.
- 3. Ukwe, O. I. K., Edun, O. M. & Akinrotimi, O. A. (2018a). Aquaculture and Fisheries: A Recipy for Job creation and Health challenges. *International Journal of Research under literal Access* 1 (4), 12-20.
- 4. Otene, B. B, & Ukwe, O. I. K. (2018). Evolution of Heavy metals Accumulation in water and sediment from Elechi Creek, Port Harcourt, Nigeria. *International Journal of Geography and Environmental Management*, 4(1): 1-9.
- 5. Ukwe, O. I. K., Edun, O. M. & Akinrotimi, O. A. (2018b). Growth and Microbial indices in African Catfish (*Clarias gariepinus*) larva fed formulated and commercial diets. *Journal of Fisheries and Science*, 12(2), 001-008.
- 6. Ukwe, I. O. K. & Gabriel, U.U. (2019). Herbs and Herbal supplements: Key to a productive, Healthy and Eco-friendly aquaculture. *Delta Agriculturist*, 11(1/1), 55 67.
- 7. Ukwe, O. I. K., Abey, I. S. & Akinrotimi, O. A. (2021). Behavioural and Haematological changes in *Pseudomonas aeruginosa* Infected *Clarias gariepinus* Exposed to Carica Papaya Roots Extracts. Hematology Asian Research Journal, 5(1): 20-33.
- 8. Puycha. K., Yuangsoi, B., Charoenwanttanasak, S., Wongmaneeprateep, S., Niamphithak, P. & Wiriyapattanasub (2017). Effect of Moringa (*Moringa Oleifera*) leaf supplementation on growth performance and feed utilization of Bocourti catfish (*Pangasiusocourtis*). Agriculture and Natural Resources, 23, 1-6.
- 9. Ukwe, I. O. K. & Vopnu, F. B. (2021). Disease Resistance and Enzymatic changes in *Pseudomonas aeruginosa* infected *Clarias gariepinus* treated with *carica papaya* root extracts. *Journal of Medicial care Research and Review* 1-6; 2589-8949/(P) 2589 8930.
- 10. Jamabo, N. A. & Dienye, H. E. (2017). Growth and Performance of *Clarias* gariepinus fed different commercial feeds. *Journal of Natural Science Research*, 7(6): 125-129.

- 11. Ogundare, A. O. & Oladejo, B. O. (2014). Antibacterial Activities of the leaf and Bark Extract of *Persea Americana*. *American Journal of Ethnomedicine*, 1(1), 64-71.
- 12. Ukwe, I. O. K., Gabriel, U. U. & Abu, O. M. G. (2020a). Periodical Evaluation of the Haematology of African Catfish (*Clarias gariepinus*) fed Dietary Powdered Avocado Peer (*Persea americana*) leaves *Global Scientific Journals*, 8(9): 1627-1641
- 13. Ukwe, I. O. K., Gabriel, U. U. & Deekae (2020b). Assessment of Dietary Powdered Avocado Peer (*Persea americana*) leaves on Growth Performance and Survival of African Catfish (*Clarias gariepinus*). *Global Scienfic Journals*, 8(8), 1612 1626.
- 14. Lee, D. H., Ra, C. S., Song, Y. H., Sung, K. I. & Kim, J. D. (2012). Effects of dietary garlic extract on growth, feed utilization and Whole body composition of juvenile starlet sturgeon (Acipenser ruthenus). *Asian Australian Journal of anima Science*, 25(4): 577 583.
- 15. Asadi, M. S., Mirvaghefei, A. R., Nematollahi, M. A., Banace, M. & Ahmadi, K. (2012). Effects of Watercress (*Nasturtium nasturtium*) extract on selected immunological parameters of rainbow trout (*Oncorhynchus mykss*) *Open Veterinary Journal*, 2, 32-39.
- 16. Wahua, T. A. T. (1999). Applied statistics for scientific studies. Africa links books. Aba, Nigeria, 365pp.
- 17. Lucky, E. & Jonathan, I. (2017). Antibacterial Activity of *Persia Americana* Leaf Extracts Against Multidrug Resistant Bacterial Isolates. *American Association of Science and Technology. Journal of Biosciences*, 3(4), 29-34.
- 18. Rahman, N., Dewi, N. U. & Bohari (2018). Phytochemical and Antioxidant Activity of Avocado Leaf Extract (*P. Americana mill*) Asian Journal of Scientific Research, 11(3), 357-366.
- 19. Kumala, S., Utami, H. & Sari, W. K. (2013). The effect of Avocado (*P. americana mill*) leaves Extract Towards the Mouse's Blood Glucose Decrease with the Glucose Tolerance Method. *International Journal of Pharmaceutical Sciences and Research*, 4(2), 661 665.
- 20. Gomez-Flores, R., ARzate-Quintana, C., Quintanilla-Licea, R., Ramez-Guerra, P., Tamez-Guerra, R., Monreal-Cuevas, E. & Rodriguez-Padilla, C. (2008). Antimicrobial Activity of *P. americana* mill (Lauraceae) (Avocado) and *Gymnosperma glutinosum* (Spreng). Less (Asteraceae) leaf Extracts and Active Fractors Against *Mycobacterium tuberculosis*. *American-Eurasian Journal of Science Research*, 3(2), 188-194.

- 21. Altemimi, A., Lakhssassi, N., Baharlouei, A., Wastom, D. G. & Lightfoot, D. A. (2017). Phytochemicals: Extraction, Isolation and Identification of Bioactive compounds from plant Extracts. *Baseel Journal*, 6(4), 42 44.
- 22. Visavadiya, N. P. & Narasimhacharya, P.A. (2009). *Asparagus* root regulates cholesterol metabolism and improves antioxidant status in hypercholesteremic rats Evidence Based Complement. *Alternative Medicine*, 6, 219-226.
- 23. Aggarwal, B. B., Piasad, S., Reuter, S., Kannuopan, R. & Yadav, V. R. (2011). Identification of novel anti-inflammatory agents from Ayurvedic medicine for prevention of chronic diseases: "Reverse pharamacology" and "bedside to bench" approach *Current Drug Targets*, 12, 1595-1653.
- 24. Southern, S., Wright, A. J. A., Price, K. R., Fairweather-Tait, S. J. & Fenwick, G. R. (1988). The effect of three types of saponin on iron and zink absortioin from a sing mill in rat *Brazikan Journal of Nutrition*, 59, 389-396.
- 25. Olaniyan, M. F. (2014). Effect of liquid Extract of Pear Avocado Leaf (*Persea americana*) on Plasma levels of Aminotransferases, cholesterol and Total Bile Acids in Hypertensive Patients. *American Journal of Medicine and Medical Science*, 4 (3), 87-91.
- 26. Boeing, J. S., Barizao, E. O., Esilva, B. C., Montanher, P. F., Almedia, V. D. & Visentainer, J. V. (2014). Evaluation of Solvent effect on the extraction of phenolic compounds and antioxidant capacities from berriers: Application of principal component analysis. *Chemistry Central Journal*, 8(1), 1-9.
- 27. Snyder, L. R., Kirkland, J. J. & Glajch, J. L. (1997). Pracical HPLC method Development. 2nd Edition, John Willey and Sons, Hoboken, New Jersey, U.S.A.
- 28. Ukwe, I.O.K., & Jamabo, N. A., (2020). Effect of dietary mango barks on *Clarias gariepinus* (Burchell, 1822) Infected with *Psuedomonas aeruginosa*. World Journal of Fish and Marine Science. 12 (3): 74-80.
- 29. Adeyemi, J. A. (2014). Oxidative stress and antioxidant enzymes activities in the African catfish. *Clarias gariepinus*, experimentally challenged with *Eshenchina coliarod* vibro fischeri. *Fish Physiology and Biochemistry*, 40, 347-354.
- 30. Yu, J. H., Han, J. J. & Park, S. W. (2010). Haematological and biochemical alterations in Korean catfish *Sihirus asotus*, experimentally infected with *Edwardsiella tarda*. *Aquaculture Research*, 41(2), 295 302.
- 31. Sebastiao, F. a., Nomura, D., Sakabe, R. & Pilarski, F. 92011). Hematology and Productive Performance of Nile tilapia (*O. niloticus*) naturally infected with Flavobacterius Columnare. *Brazilian Journal of Microbiology*, 42:282 289.

- 32. Rajendiran, A., Natarajan, E. & Subramania, P. (2008). Control of *A hydrophila* infection in spoted snake head, *Channa punctatus* by *Solanumnigrum* L. a medicinal plant. *Journal of the World Aquacultural Society*, 39(3): 375 382.
- 33. Sivagurunathan. A., Innocent, B. X., Saraswathi, S. G. & Mariappan, A (2012). Immunostimulatory Potential Dietary Amila (*Phyllanthus emblica*) in Growth and Haematology of Tilapia Mossambicus Challenged with *Pseudomonas aeruginosa*. *International Research Journal of Pharmacy*, 3(7), 165-168.
- 34. Joshi, P. K., Harish, D., & Bose, M. (2002). Effect of Lindane and Malathione exposure to certain blood parameters in a fresh water teleost fish clarias batrachus. *Pollution Research*, 21(1), 55 57.
- 35. Awe, F. A., Hammed, A. M., Akinyemi, A. A., Whenu, O. O. & Olanloye, O. A. (2019). Antibacterial Activities of Mango leaf (Manifera indica) Extracts on catfish clarias gariepinus (Burchell, 1822) infected with *Pseudomonas aeruginosa*. Asian Journal of Agricultural Research, 13(1), 28-36.
- 36. Fadi, S. E., Barak, M. & Elogohari, M. (2013). Biochemical studying of Anabaena (Cyanobacteria) on Nile Tilapia. *Alexandria Journal of Veterinary Sciences*, 39, 91 104.
- 37. Souza, C. F., Baldissera, M. D., Vaucher, R. A., Lopes, L. Q. S. & Vizzotto, B. S. (2016). *In vivo* bactericidal effect of *Melaleuca alternifolia* essential oil against *Aeromonas hydrophila*. Silver catfish (*Rhamdia quelen*) as an experimental model. *Microbiology and Pathology*, 98, 82-87.
- 38. Innocent, B. X., Fatima, M. S. A. & Dhanalakshmi (2011). Studies on the Immunostimulant activity of Coriandrum Sativum and resistance to Aeromonas hydrophila in catla. *Journal Applied Pharmaceutical Science*, 1(7): 132 135.
- 39. Sales, C. H., Souza, P. R., Peghini, B. C., Da Silver, J. S. & Cardoso, C. R. (2013). An overview of the modulatory effects of Oleic acid in health and diseases. *Minireviews in Medicinal Chemistry*, 13, 1-11.
- 40. Oyeyemi, A. O. & Oyeyemi, R. B. (2015). Effect of Aqueous of the leaves and seeds of Avocado pear (*Persea Americana*) on some Marker Enzymes and Cholesterol in Albino Rat Tissues. *Journal of Environmental Science, Toxicology and food Technology*, 9 (3), 15-18.
- 41. Jagadeeswaram, P., Gregory, M., Day, K., Cykowski, M. & Thattaliyath, B. (2005). Zebrafish: a genetic model for hemostasis and thrombosis. *Journal of Thrombosis and Heamostasis*, 3(1), 46-53.

- 42. Etim, L., Ekanem, S. B. & Utim, A. (1999). Haematological profile of two species of catfish, *Chrystcthys nigrodigitatus* and *Chrysicthys furcatus* from the great Kwa River, Nigeria. *Global Journal of Pure and Applied Science*, 5(1): 1 8.
- 43. Sivaram, V., Babu, M. M. Citarasu, T., Immanuel, G., Murugadass, S. & Mariam, M. P. (2004). Growth and Immune response of juvenile greasy groupers (*Epinephelus tavivina*) fed with herbal antibacterial active principle supplemented diets against *Vibro harveyi* infections. *Aquaculture*, 237, 9-20.
- 44. Bello, O. S., Olaifa, F. E., Emikpe, B. O. & Ogunbanwo, S. T. (2012). The effect of walnut (*Tetracarpidium conopolhorum*) leaf and onion (*Allium cepa*) bulb reduces on tissue bacteriological changes of *Clarias gariepinus* juviniles. *Bulletin of Animal Health and Protection in Africa*, 60(2), 205-212.
- 45. Reverter, M., Bontemps, N., Lecchini, D., Banaigs, B., and Sasal, P. (2014). Use of plant extracts in fish aquaculture as alternative to chemotherapy: Current status and future perspective. *Aquaculture*, 433: 50 61.
- 46. Ardo, L., Yin, G., Xu, P., Varadi, L., Szigeti, G. and Jeney, Z. (2008). Chinese herbs (*Astragalus membranaeus & lonicera japonica*) and bofon enhance the non-specific immune response of Nile tilapia (*O. niloticus*) and resistance agains *A. hydrophila*. *Aquaculture*, 275:26-33.
- 47. Tewary, A. & Patra, B. C. (2008). Use of vitamin C as an immunostimulant effect on growth, nutrititional quality and immune response of *labeo rohita (Ham.) Fish Physiology and Biochemistry*, 34, 251-259.