

Heat treated African kudzu (*pueraria phaseoloides, roxb. benth.*) flour influenced lipid peroxidation, liver enzymes and organ histology of wistar rats

## Abstract

Scarcity of protein-rich foods has been the bane of developing countries with its attendant nutritional problems. Research efforts have been directed towards finding solutions to this problem. This include investigation into the nutritional potentials of some lesser known legumes and grains. This paper looked at the effect of processed kudzu flours on the liver enzymes, alkaline phosphatase (ALP), aspartate amino transferase (AST) and alanine amino transferase (ALT); lipid peroxidation of the liver and pancreatic tissues as well as the liver and pancreatic architecture of wistar rats. Raw, cooked and time-controlled autoclaved (20mins, 40mins, 60mins) kudzu seeds were made into flours and compounded into 8 diets. Diet 1 contained the raw kudzu flour, diet 2 had the cooked kudzu flour and diets 3-5 had time-controlled autoclaved (20mins, 40mins, 60mins) kudzu flour respectively. Diet 6 was the negative control diet while diet 8 was the positive control. Diet 8 had lower ALP value ( $51.6 \pm 6.5$ iu) than the other groups though not statistically ( $p \leq 0.05$ ) different. Diet 3 had the highest AST ( $51.0 \pm 0.1$ iu) which was statistically ( $p \leq 0.05$ ) different from the other groups and ALT ( $48.0 \pm 0.1$ iu) that was statistically ( $p \leq 0.05$ ) not different with only group 4. The level of malondialdehyde in the liver was lowest in Diet 2 ( $2.75 \times 10^{-5} \pm 0.07 \times 10^{-5}$ MDA unit) followed by Diets 6 and 8. While the level in the pancreatic tissues was lowest in Diet 8 ( $83.25 \times 10^{-5} \pm 0.07 \times 10^{-5}$ MDA unit). However, the variations in the level of MDA in both organs in all the groups were not statistically ( $p \leq 0.05$ ) different. Histopathological examination of the liver tissues showed various degrees of abnormalities in Diets (1, 3, 4, 5 and 6) while Diets 2, 7 and 8 had normal cell architecture. The pancreatic tissues showed normal architecture. The results therefore suggest that kudzu when processed poses no toxicological danger to animal and by extension man.

Key words: African kudzu, liver enzymes, lipid peroxidation, organ histology.

## Introduction

The worsening food insecurity globally, exacerbated by climate change [1], economic downturn of many nations, artificial and man-made disasters, various forms of regional and territorial armed conflicts [2,3] and recently a global disease pandemic [4] has necessitated the quest for multidimensional approach in dealing with the situation. Several approaches have been adopted while some are being worked out such as probing into the nutritional potentials of some food items and so encourage their exploitation for immediate and future food security, the others aim to make food production more environmentally friendly and more resilient to climate shocks [5] while others concentrate on improving the food

environment and advocacy for policy briefs in food and nutrition issues.

Probing into the nutritional or functional properties of food items has been a common practice right from the early times. Our Ancestors through 'crude methods' or intuitions were able to decipher what food was 'good' for them. Indeed, the process was the basis for identifying edible and isolating the inedible food items long before the advent of civilizations. Subsequently, scientific technologies were developed and the investigations continued, providing more reliable information and data. Therefore, several authors have provided nutritional, phytochemical, therapeutic and toxicological information on different foodstuff including vegetables [6,7,8,9,10,11]; fruits [12,13,14,15]; cereals [16,17,18]; legumes [19,20,21]; roots and tubers [22,23,24], nuts and seeds [25,26,27] etc. The immediate and remote implications of food insecurity are burdensome. They include high food prices and attendant lack of access to food, food insecurity predisposes individuals to increased risk to a variety of diseases and other health challenges [28,29,30,31,32,33]. Food insecurity is a major factor in the cause of different forms of crime and criminality. Food security index of a country depicts the country's poverty and consequently her development index. Thus, food security is central for the achievement of the some of the Sustainable development goals (SDGs) particularly goals 1-3. Therefore, any contribution towards alleviating food insecurity will help in the development agenda of any nation. This study looked at the effect of some processing methods on selected parameters of wistar rats fed with diets formulated from an unutilized legume-*kudzu*. The broad objective is to popularize its use either for human or animal nutrition.

## Materials and Methods

Kudzu seeds were obtained from International Institute for Tropical Agriculture, (IITA) Onne, Rivers State while maize (*Zea mays*) grains and red palm oil were purchased from the Mile 3 Market, Port Harcourt, Rivers State. Granulated pure cane sugar (sucrose) and Nutrend were purchased from Sunrise Supermarket, University of Port Harcourt. Vitamins and mineral mixtures were bought at Raf - Veterinary Store, Rumuigbo Port Harcourt, Rivers State. One kilogram (1kg) of the seeds was not given any heat treatment while same quantity was cooked; autoclaved at 121°C (15psi) for 20 minutes, 40 minutes and 60 minutes respectively. They were later ground into fine powder (710mm sieve) using an electric mill and individually packed in air-tight bags and stored in the refrigerator at 4°C until used for analysis.

Seven (8) diets were formulated using corn starch, kudzu flour (raw and processed), sucrose, non-nutritive cellulose, palm oil and vitamin/mineral mixture. Diet 1 contained the raw kudzu, diet 2 cooked kudzu, diet 3 kudzu autoclaved at 121°C (15psi) for 20 minutes, diet 4 kudzu autoclave at 121°C 15psi for 40 minutes, diet (5) kudzu autoclave at 121°C (15psi) for 60 minutes. The sixth diet (6) was the basal diet (protein free diet) while diet seven (7) was made up of 100% cooked kudzu and diet eight (8) was the reference diet (Nutrend). The diets were formulated to provide 16% protein.

A total of 40 wistar rats aged between 24-28 days, obtained from the Biochemistry Department, University of Port-Harcourt were used for the study. All experimental procedures herein were carried out in accordance with the ethical guidelines for laboratory animals and complied with the guide for the care and use of laboratory animals (National Research Council, 2011). The animals were weighed and divided into eight groups of five (5) rats each on the basis of their body weights. They were singly housed in perforated Perspex wire bottom cages with facilities for food and water. The rats were acclimatized for seven days and fed with the experimental diet and water *adlibitum* for 28 days. They were then sacrificed by cervical dislocation on the 28<sup>th</sup> day. Their blood collected for estimation of liver enzyme. The animals were dissected, their liver and pancreas excised, inspected for any pathological abnormalities and later histological procedure carried out on them according to Baker and Silverton [34] method. MDA estimation was done on the tissues using Gutteridge and Wilkins [35] method modified from Hunter et al., [36].

The results were analyzed statistically by the use of one way analysis of variance (ANOVA) to determine the differences between the mean values at  $P < 0.05$  level [37]

## Results and Discussion

The results of the lipid peroxidation of the liver and pancreatic tissues as well as the liver enzyme assay of the experimental animals are shown in Table 1. The values for malondialdehyde (MDA) in the liver tissues was highest among the animals on diet 1 ( $32 \times 10^{-5} \pm 0.0$  MDA unit) and lowest in animals on diet 4 ( $2.7 \times 10^{-5} \pm 0.14$  MDA unit). The values were however not statistically ( $P < 0.05$ ) different from the other groups. There were also no statistical difference in the MDA values for pancreatic tissues. MDA is a by-product of lipid peroxidation. Lipid peroxidation occurs in tissues when tissues are exposed to adverse environment including toxicants. It is an oxidative damage that affects cellular membranes, lipoproteins, and other molecules that contain lipids. Cellular membrane lipids represent most often substrates of oxidative attack [38]. Lipid peroxidation is a chain reaction and is created by free radicals influencing unsaturated fatty acids in cell membranes, leading to their damage. Free radicals are initiators and terminators of lipid peroxidation processes. Once activated, reaction continues autocatalytically; it has a progressive course, and its final result is structural and functional changes of substrate [39]; [40]. Thus, high level of MDA has been implicated in several human diseases. For instance, corneas of patients suffering from keratoconus and bullous keratopathy have increased levels of malondialdehyde [41] and MDA also can be found in tissue sections of joints from patients with osteoarthritis. [42].

Levels of malondialdehyde is also used to assess the membrane damage in spermatozoa; this is crucial because oxidative stress affects sperm function by altering membrane fluidity, permeability and impairing sperm functional competence. [43] The comparable values of MDA in all the groups in this study may suggest that the toxic phytochemicals in the legume had no negative effect on the lipid-containing structural

components of the experimental animals.

The values for the liver enzyme, alkaline phosphatase (ALP) showed that diet 2 had the lowest value of  $49.67 \pm 9.29$  IU followed by diet 7 ( $50.7 \pm 5.0$  IU) and diet 8 ( $51.60 \pm 5.5$  IU) but the values were not significantly ( $P < 0.05$ ) different from others. Aspartate amino transferase (AST) was least in diet 8 ( $39.2 \pm 6.83$  IU) and statistically lower than only diet 3 ( $51.0 \pm 2.16$  IU). Alanine amino transferase (ALT) had the lowest value in diet 7 ( $36.0 \pm 7.62$  IU) followed by diet 8 ( $37.0 \pm 0.0$  IU) and the values were statistically different from those of diet 3 ( $48.0 \pm 1.10$  IU) and 4 ( $46.67 \pm 3.51$  IU). Enzymes are membrane-bound and damage to the cells and membrane of the liver may result in their release with other intracellular constituents into the blood plasma [44,45]. Increases in the activities of these enzymes following exposure to toxicants have been reported [46]. The serum levels of these enzymes reflect the physiological state of the liver. Since the values of these liver enzymes in the experimental animals fed with the cooked diets were always comparable with those of the reference diet, it can be inferred that the result was due to the lowering effect of wet heat on some toxic phytochemicals [47,48].

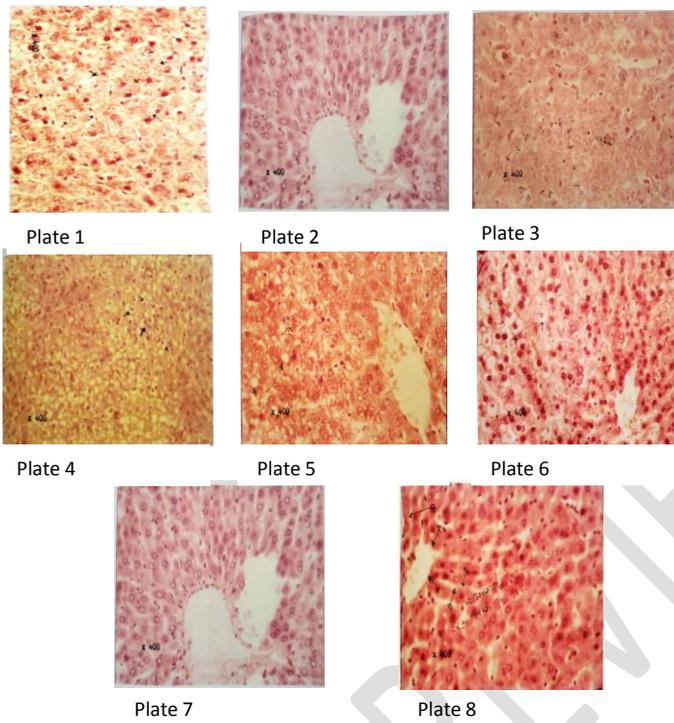
The microphotographs of the liver and the pancreas are shown on plates 1-16. The pictures for the two organs in diets 2, 7 and 8 were essentially normal while others showed varying degrees of abnormalities which may be attributed to the presence of toxic phytochemicals. However, since the MDA values for the organs across the groups were comparable as well as the liver enzyme activities, it may be inferred that the observed abnormalities in the organs may not be physiologically detrimental.

**Table 1 Lipid Peroxidation (MDA unit) and Enzyme Assay (UI)**

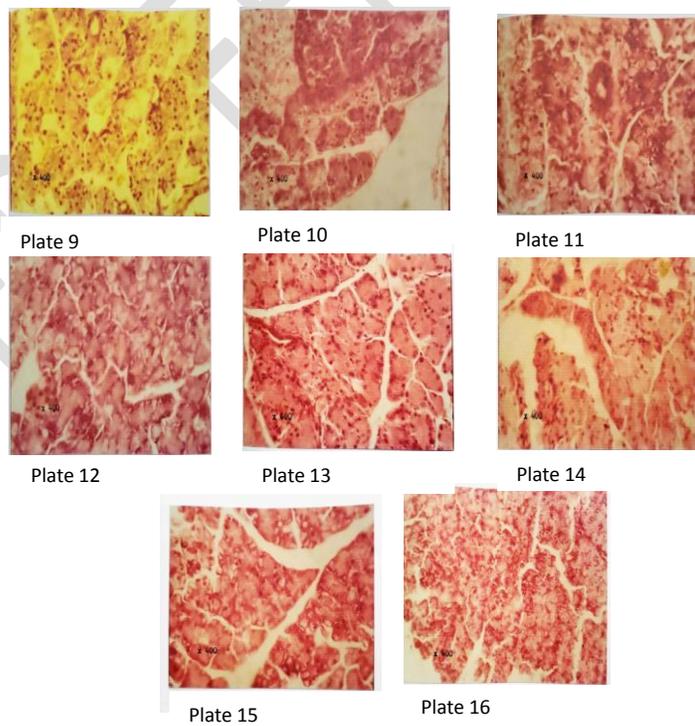
Diet	Liver	Pancreas	ALP	AST	ALT
1	3.2×10 <sup>-5a</sup> ± 0.0	99.35×10 <sup>-5a</sup> ± 46×10 <sup>-5</sup>	52.25 <sup>a</sup> ± 4.43	47.25 <sup>a</sup> ± 3.3	43.25 <sup>a</sup> ± 3.86
2	2.75×10 <sup>-5a</sup> ± 0.21×10 <sup>-5</sup>	89.7×10 <sup>-5a</sup> ± 9.05×10 <sup>-5</sup>	49.679 <sup>a</sup> ± 9.29	45.67 <sup>a</sup> ± 6.11	39.0 <sup>a</sup> ± 0.0
3	3.1×10 <sup>-5a</sup> ± 0.0	96.1×10 <sup>-5a</sup> ± 0.0	54.25 <sup>a</sup> ± 1.71	51.0 <sup>b</sup> ± 2.16	48.0 <sup>b</sup> ± 1.16
4	2.7×10 <sup>-5a</sup> ± 0.14×10 <sup>-5</sup>	92.9×10 <sup>-5a</sup> ± 4.53×10 <sup>-5</sup>	55.33 <sup>a</sup> ± 6.51	49.67 <sup>a</sup> ± 4.16	46.67 <sup>b</sup> ± 3.51
5	2.90×10 <sup>-5a</sup> ± 0.21×10 <sup>-a</sup>	92.95×10 <sup>-5a</sup> ± 13.65×10 <sup>-5</sup>	54.0 <sup>a</sup> ± 9.54	46.35 <sup>a</sup> ± 14.36	41.0 <sup>a</sup> ± 13.08
6	2.85×10 <sup>-5a</sup> ± 0.35×10 <sup>-5</sup>	92.95×10 <sup>-5a</sup> ± 13.65×10 <sup>-5</sup>	55.0 <sup>a</sup> ± 6.68	44.0 <sup>a</sup> ± 9.5 <sup>a</sup>	41.5 <sup>a</sup> ± 10.34
7	3.05×10 <sup>-5a</sup> ± 0.21×10 <sup>-5</sup>	89.7×10 <sup>-5a</sup> ± 0.00	50.5 <sup>a</sup> ± 5.0	41.75 <sup>a</sup> ± 7.5	36.0 <sup>a</sup> ± 7.64
8	2.85×10 <sup>-5a</sup> ± 36×10 <sup>-5</sup>	83.25×10 <sup>-5a</sup> ± 0.07×10 <sup>-5</sup>	51.60 <sup>a</sup> ± 6.5	39.2 <sup>a</sup> ± 6.83	37.0 <sup>a</sup> ± 6.63

Values are mean± SD for 5 rats per diet group (n=5). Means in the same column with similar superscript letters are not significantly different at 5% level (p<0.05).

Legend:MDA-Malondiadehyde,ALP-Alkaline phosphotase,ALT-Alanine amino trananferase,AST-Aspartate amino transferase.



**Figure 1: (Plates 1-8) Histological examination of the liver tissue magnification  $\times 400$**



**Figure 2: (Plates 9-16) Histological examination of the pancreas tissue magnification  $\times 400$**

## Conclusion

This study has revealed that, African *kudzu* when processed by cooking posed no obvious toxicologically threat to the experimental animals. The legume could there be used for either of human or animal feed.

## COMPETING INTERESTS DISCLAIMER:

Authors have declared that no competing interests exist. The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

## Reference.

1. Parodi A, Leip A, De Boer IJM, Slegers PM, Ziegler F, Temme EH, Herrero M, Tuomisto H, Valin H, Van Middelaar CE, Van Loon JJA, Van Zanten HHE. The potential of future foods for sustainable and healthy diets. *Nature Sustainability*. 2018; 1: 782.
2. Ujunwa A, Okoyeuzu C, Kalu EU. "Armed Conflict and Food Security in West Africa: Socioeconomic Perspective. *International Journal of Social Economics*. (2019);46(2):182-198. <https://doi.org/10.1108/IJSE-11-2017-0538>
3. Van Weezel S. Food Security and Armed Conflict: A Cross-Country Analysis. FAO Agricultural Development Economics Working Paper 18-03, 2018. Rome.
4. The World Bank Food Security and COVID19. 2021. <https://www.worldbank.org/en/topic/agriculture/brief/food-security-and-covid-19>
5. Beddington JR, Asaduzzaman M, Clark ME. The role for scientists in tackling food insecurity and climate change. *Agriculture and Food Security*. 2012; 1: 10 <https://doi.org/10.1186/2048-7010-1-10>
6. García-Herrera P, Morales P, Cámara M, Fernández-Ruiz V, Tardío J, Sánchez-Mata MC. Nutritional and phytochemical composition of mediterranean wild vegetables after culinary treatment. *Foods*. 2020; 9: 1761; doi:10.3390/foods9121761]
7. Zhan LJ, Pang LY, Ma YD, Zhang CC. Thermal processing affecting phytochemical contents and total antioxidant capacity in broccoli (*Brassica oleracea* L.). *Journal of Food Processing and Preservation*. . 2018;42:1045
8. Danowska-Oziewicz M, Narwojsz A, Draszanowska A, Marat N. The effects of cooking method on selected quality traits of broccoli and green asparagus. *International Journal of Food Science and Technology*. 2020; 55: 127–135.
9. Ifeanacho MO, Ogunwa SC. Nutritional and bioactive potentials of an underutilized vegetable- *Vitex doniana*. *Food and Nutrition Sciences*. .2021;12:975-995. doi:10.4236/fns.2021.1210072.

10. Ikewuchi JC, Ikewuchi CC, Ifeanacho MO. Nutrient and bioactive compounds composition of leaves and stems of *Pandiaka heudelotti*: A Wide vegetable. *Heliyon*. 2019;5(4):e01501
11. Ifeanacho MO, Oshotse RB. Evaluation of the biochemical impact of varied mixtures of extracts of *vernonia amygdalina* (bitter leaf) and *gnetum africanum* (okazi leaf) on alloxan induced diabetic wistar rats. *Sciencia Africana*. 2020;20(1): 17-30.
12. Edeke A, Uchendu N, Omeje K, Odiba AS. Nutritional and Pharmacological Potentials of *Solanum melongena* and *Solanum aethiopicum* Fruits. *Journal of Phytopharmacology*. 2021;10(1):61-67
13. Olufunmilayo DA. Mini review on two species of garden egg (*S. aethiopicum L.* and *S. macrocarpon L.*) found in Nigeria. *Journal of Analytical and Pharmaceutical research*. 2018; 7(2):237–8.
14. Daagema AA, Orafa PN, Igbua FZ. Nutritional potentials and uses of pawpaw (*Carica papaya*): A Review. *European Journal of Nutrition and Food Safety*. 2020;12(3):52-66. <https://doi.org/10.9734/ejnfs/2020/v12i330209>
15. Ifeanacho MO, Ogunka-Nnoka CU. Evaluation of nutritional value of black plum (*vitex Doniana*): An attempt to broaden our food base via an unconventional source. *Proceedings of International Conference on Science and Sustainable Development*. 2011; 1(1): 124-130
16. Ma M, Sun QJ, Li M, Zhu KX. Deterioration mechanisms of high-moisture wheat-based food – A review from physicochemical, structural, and molecular perspectives. *Food Chemistry*. 2020; 318:126495. <https://doi.org/10.1016/j.foodchem..126495>
17. Saini A, Panwar D, Panesar PS, Bera MB. Bioactive compounds from cereal and pulse processing by-products and their potential health benefits. *Austin Journal of Nutrition and Metabolism*. 2019; 6(2): 1068.
18. Elijah H K, Smith GN, Emmanuel OA. Extrusion and nixtamalization conditions influence the magnitude of change in the nutrients and bioactive components of cereals and legumes. *Food Science and Nutrition*. 2020; 8 (4): 1753-1765, <https://doi.org/10.1002/fsn3.1473>
19. Ifeanacho MO, Ezecheta CC. Effect of domestic food processing methods on anti nutrients, some mineral content and functional properties of mungbean (*Vigna radiata*) flours. *Journal of Dieticians Association of Nigeria*. 2020;11(1):45-52
20. Ifeanacho MO, Abbey BW, Ayalogu EO. Effects heat treatments on the African Kudzu, (*Pueraria Phaseoloides*, Roxb. Benth) seeds. *Nigerian Journal of Nutritional Sciences*. 2008; 29(2): 318-329.
21. Agomuo EN, Amadi PU, Ogunka-Nnoka CU, Amadi BA, Ifeanacho MO, Njoku UC. Characterization of oils from *duranta repens* leaf and seed. *Oil Seeds and Fats, Crop and Lipids*. 2017;24(6):1-8. DOI:doi.org/10.105/oc/2017048.
22. Ogunjobi AA, Adebayo-Tayo BC, Ogunshe AA. Microbiological, proximate analysis and sensory evaluation of processed Irish potato fermented in brine solution. *African Journal Biotechnology*. 2005; 4(12):1409–1416

23. Obidiegwu JE, Lyons JB, Chilaka CA. The dioscorea genus (Yam)—An appraisal of nutritional and therapeutic potentials. *Foods*. 2020; 9:1304; doi:10.3390/foods9091304
24. Ifeanacho MO, Oloya CG, Essien EB. Nutritional evaluation of complementary foods based on yam, and orange fleshed sweet potato leaves. A paper presented at the annual national scientific conference of Association of Nigerian Dietitians, 2019, held at Asaba, Delta State, Nigeria.
25. Ballhorn DJ. Cyanogenic glycosides in nuts and seeds in health and disease prevention. Elsevier Inc. 2011;129–136, <http://dx.doi.org/10.1016/B978-0-12-375688-6.10014-3>
26. Chaouali N, Gana I, Dorra A, Khelifi F, Nouioui A, Masri W, Belwaer I, Ghorbel, H. Hedhili A. Potential Toxic Levels of Cyanide in Almonds ( *Prunus amygdalus* ), Apricot Kernels ( *Prunus armeniaca* ), and Almond Syrup. *Toxicology*. 2013:1–6
27. Ikewuchi CC, Ifeanacho MO, Ikewuchi JC. Fatty acids composition and oil characteristics of nut kernel oil of cashew (*anarcadium occidentale l*) from Nsukka, Nigeria. *Zimbabwe Journal of Science and Technology*. 13:40-46.
28. Holben DH, Pheley AM. Diabetes risk and obesity in food-insecure households in rural Appalachian Ohio. *Preventing Chronic Disease*. 2006;3(3). [http://www.cdc.gov/pcd/issues/2006/jul/05\\_0127.htm](http://www.cdc.gov/pcd/issues/2006/jul/05_0127.htm)
29. Seligman HK, Laraia BA, Kushel MB. Food insecurity is associated with chronic disease among low-income NHANES participants. *Journal of Nutrition*. 2010; 140(2), 304-310. <http://doi.org/10.3945/jn.109.112573>.
30. Gundersen C, Kreider B. Bounding the effects of food insecurity on children's health outcomes. *Journal of Health Economics*. 2009; 28(5):971-983.
31. Metallinos-Katsaras E, Must A, Gorman K. A longitudinal study of food insecurity on obesity in preschool children. *Journal of the Academy of Nutrition and Dietetics Journal of the Academy of Nutrition and Dietetics*. 2012;112(12):1949-58.
32. Cook, J.T (2013). Impacts of child food insecurity and hunger on health and development in children: Implications of measurement approach. In paper commissioned for the Workshop on Research Gaps and Opportunities on the Causes and Consequences of Child Hunger.
33. Burke MP, Martini LH, Çayır E, Hartline-Grafton HL, Meade RL. Severity of household food insecurity is positively associated with mental disorders among children and adolescents in the United States. *Journal of Nutrition*. 2016;146(10):2019-2026. doi: 10.3945/jn.116.232298.
34. Baker FJ, Silverton ER, Kilsnaw D. Introduction to medical laboratory technology. Butterworths, London. 1985;316-367
35. Guttridge JMC, Wilkins C. Copper dependent hydroxyl radical damage to ascorbic acid. Formation of a thiobarbituric acid reactive products. *FEBS letters*. 1992;137:327-3

36. Hunter FE, Gebicki JM, Hoffstein DE, Weinstein J, Scott A. Swelling and lysis of rat liver mitochondria induced by ferrous ions. *Journal of Biological Chemistry*. 1963;238:828-835.
37. Norusis MJ. One way analysis of variance. 1986.spp/pc for the Ibm Pc/XT/AT/SPSS Inc. Michigan
38. Nawrot T S, Van Hecke E, Thijs L. Cadmium related mortality and long-term secular trends in the cadmium body burden of an environmentally exposed population. *Environmental Health Perspective*. 2008; 116(12): 1620-8.
39. Cuypers A, Semane B, Dupae J, Noben J-P, Tuomainen M, Tervahauta A, Kärenlampi S, Van Belleghem F, Smeets K, Vangronsveld J. Leaf proteome responses of *Arabidopsis thaliana* exposed to mild cadmium stress. *Journal of Plant Physiology*. 2010 ; 167( 4):247-254.
40. Ognjanović BI, Marković SD, Pavlović SZ, Zikić RV, Stajin AS, Saicić ZS. Effect of chronic cadmium exposure on antioxidant defense system in some tissues of rats: Protective effect of selenium. *Physiology Research*. 2008; 57:403-411.
41. Buddi R, Lin B, Atilano SR, Zorapapel NC, Kenney MC, Brown DJ "Evidence of oxidative stress in human corneal diseases". *Journal of Histochemistry and cytochemistry*. 2002; 50 (3): 341–51. doi:10.1177/002215540205000306. PMID 11850437.
42. Tiku ML, Narla H, Jain M, Yalamanchili P. "Glucosamine prevents *in vitro* collagen degradation in chondrocytes by inhibiting advanced lipoxidation reactions and protein oxidation". *Arthritis Research and Therapy*. 2007; 9 (4): R76. doi:10.1186/ar2274. PMC 2206377. PMID 17686167.
43. Collodel G, Moretti E, Micheli L, Menchiari A, Moltoni L, Cerretani D. "Semen characteristics and malondialdehyde levels in men with different reproductive problems". *Andrology*. 2015; 3 (2): 280–286. doi:10.1111/andr.297. PMID 25331426. S2CID 28027300.
44. Wilkinson HJ. *Diagnostic Enzymology*, Edward Arnold, London. 1976;527-533
45. Gazuwa S Y, Dabak JD, Jaryum KH, Oluwa I. Relationship between Dose and Duration of Administration of Potassium Bromate on Selected Electrolytes and Hepatorenal Parameters in Male Albino Wistar Rats. *European Journal of Nutrition and Food Safety*. 2020; 11(4): 214-220.
46. Singh NS, Vats P, Suri S, Shyam R, Kumria MML, Ranganathan S, Sridharan K. Effect of an antidiabetic extract of *Catharanthus roseus* on enzymic activities in streptozotocin induced diabetic rats. *Journal of Ethnopharmacology*. 2001; 76:269–277
47. Ifeanacho MO, Abbey BW, Ayalogu EO. Effects heat treatments on the African *Kudzu*, (*Pueraria Phaseoloides*, Roxb. Benth) seeds. *Nigerian Journal of Nutritional Sciences*. 2008; 29(2): 318-329.
48. Simona M, Alessandro Dal B, Cesare C, Beatrice F, Valeria S, Ombretta M, Alice CM, Elisa C, Paolo B. Effect of heat- and freeze-drying treatments on phytochemical content and fatty acid

profile of alfalfa and flax sprouts. Journal of the Science of Food and Agriculture. 2019; 99  
(8):4029-4035

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