Original Research Article

THE SERO-EPIDEMIOLOGY AND RISK FACTORS OF *E.HISTOLYTICA* INFECTION IN CALABAR, NIGERIA

Abstract

Background: Amoebic infections is a prevalent infection in Nigeria._Currently, There is paucity of data on sero-epidemiology of *Entamoeba_histolytica_* in calabar. This study investigated the sero-epidemiology and risk-factors of *E_ntamoebahistolytica* infection among dysentery patients presenting at general hospital calabar, Cross river.

Methods: The Sero-epidemiology of *E. histolytica* were determined in three hundred and eighty-one subjects in calabar, Cross river state using an enzymelinked immunoassay. In addition, sero-prevalence association with the sociodemographic and risk factors of the subjects studied was investigated.

Results: Forty five (45) out of the three hundred and eight one (381) samples were positive for *E.histolytica*, 45(11.8%). Subjects in the age group 1-10 years had the highest prevalence (32.8%). The study also revealed that *E. histolytica* infections was associated with age, educational status, occupational status, source of water, toilet facility, Hand washing and contact with domestic animal/faecal matter(p<0.05). Females were more infected (13.5%) than male (9.9%) but the difference was not statistically significant (p> 0.05). The infection rate was higher in the wet season (18.1%) than the dry season (4.9%). However, *E. histolytica* infection was statistically associated with season (p<0.05).

Conclusions: The sero prevalence of *E. histolytica* infection found— in this study ismoderate compared with those reported in other Nigeria populations. The data of *E. histolytica* sero-positivity found in the present study may be useful for the planning of optimal preventive measures against *E. histolytica* infection.

Keywords: E. histolytica sero-positivity infection prevalence

INTRODUCTION

Entamoeba histolytica is a protozoan parasite that causes amoebiasis, High morbidity and mortality of_amoebisis have been reported worldwide. Infections with E. histolytica_are prevalent in the tropics and the major health problems in

developing countries_[3,4], Majority of the *E.histolytica* infection morbidity and mortality occurs in Africa, Central and South America and the indian subcontinent. The prevalence of amoebiasis due to *E. histolytica* has been difficult to establish because there is a probability tooverestimate it in endemic areas, where cases of dysentery or bloody diarrhea are often misdiagnosed as amebiasis due to the non-pathogenic *E.dispar*[6].

In non-endemic_areas with a low incidence of the disease, there is a tendency to underestimate, the prevalent of *E. histolytica* infection due other bacterial and viral pathogen causing dysentery and diarrhea [7]_Studies in parts of Africa reported prevalence rates of 22% and 21% in South Africa and Egypt, respectively [8]. In Nigeria, prevalence rates of 21.6% in Enugu [9]and 13.7% in Ilesha have been reported[10]. The rate of infection by *E. histolytica* differs among countries, socioeconomic and sanitary conditions and population's [8]. It is highly endemic throughout the tropics and subtropics [11]. Environmental, socio-economic, demographic and hygiene-related behavior is known to influence the transmission and distribution of intestinal parasitic infections. Humans are the host of *E. histolytica* and there are no other known animal reservoirs of this parasite [5]

Most persons infected with *E. histolytica* are carrier [13];_infection with *E. histolytica* is responsible for most cases of prolonged diarrhea in travelers_[14]. In addition, infection with *E. histolytica* may lead to the development of live-threateningabscess in liver, brain [15] or lungs_[5],_its becomes imperatives to investigate the sero-epidemiology of *E.histolytica* parasite in the population. Transmission of *E. histolytica* occurs in areas with poor sanitation by contamination of drinking water or food with human feces_[16]. Water-associated outbreaks of *E. histolytica* disease have been reported_[17].

There is -paucity of data on the sero-epidemiology of *E. histolytica* infection in calabar, Cross river state. Most of data are based on microscopic diagnosis of *E.histolytica* in stool which could be non-Pathogenic *E.dispar*[18]. Furthermore, socio-demographic and Risk factorsof the -subjects associated with *E. histolytica* sero-prevalence were investigated for public health intervention.

MATERIAL AND METHOD STUDY AREA

Cross river state -derives its name from the river which passes through the state-. It is a coastal -state located in the Niger Delta, and occupies 20,156 square kilometers. It shares boundaries with Benue state to the North, Ebonyi and Abia state to the west, its shares boundaries with Cameroun to the east and to the south west by Akwa Ibom State, Calabar is located in The South South geopolitical zone of Nigeria with a population of 3,737,517 inhabitants, Calabar is often described as the tourism capital of Nigeria(19).

Sample size

The minimum sample size was determined by using the formula described by Naing $et\ al$, [20], Therefore, to obtain a more reliable result, A total of -381 patients sample was collected.

Ethical Consideration

Informed consent was obtained from each of the patient or their parents. The work was also approved by the Ethical Committee of the Ministry of Health of -Cross River State. The general hospital Calabar used in this study.

Enrolment of patients

All patients presenting to the general hospital in -Calabar with acute and persistent diarrhoea or Dysentery within the 12 months (January to December,2013) -period of study was ere enlisted having consented to participate and fulfilled the inclusion criteria which included acute or persistent diarrhoea and dysenteric syndrome. Patients with diarrhoea or dysentery on antimicrobial agents were excluded. Patients visiting the hospital for reasons other than diarrhoea and had no diarrhoeal illness within the last 2 weeks were used as control.

Data collection

A well-Structured questionnaire was used to obtain information from each patients on the demographics and risk factors. Demographics include; Age,sex,Occupational status and education.Risk factors includes; Toilet facilities, Source of water and Contact with domestic animals/Faecal matter.

Specimen collection and processing

Three hundred and -eighty one_—samples (381) blood specimens were aseptically collected,_One hundred and eighty three_(183) samples were collected during the dry season (November-March)_and One hundred and ninety -eight samples_(198) collected during the wet season_(April-October)-,rRespectively. The sera obtained fromclotted blood and centrifuged to obtain the serum for each patient presenting with dysentery or diarrhoea at general hospital calabar,_Cross river State.

Analysis of samples

Detection of *E. Histolytica* antibody was carried out using the Enzyme linked Immunosorbent assay.

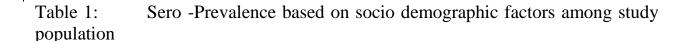
ELISA Antibody Detection Technique

The sera from the blood samples were analyzed using the Enyzme Linked Immunosorbent Assay (ELISA) made by TechLab (USA). The first well of the microplate was left blank (control) while 100ul of negative and positivecontrols were added to the second and third wells respectively. Two drops of the diluted test samples were added to the remaining wells and incubated at room temperature (15-

20°C) for 10 minutes. The contents were then shaken andwashed 3 times with diluted buffer. After washing, 2 drops of enzyme conjugate were added to each well and againincubated at room temperature for 5 minutes. This was followed by another shaking and washing again with buffer after which 2 drops of chromagen was added to each well and again incubated at room temperature. Finally, 2 drops of the stop solution were added to each welland mixed by tapping the strip holder. The results were read with a microplate reader machine set for biochromatic readings at 450/650-620nm. Positive and negative control sample was included for each batch of sample assayed.

Data analysis

The prevalence of *E. histolytica* was determined by the percentage of patients who tested positive, while the Chi-square was used to determine the association of between *E histolytica* and the selected variables. All statistical analyses were carried out using the SPSS statistical software.



Parameters/	Number	Number	x ² dfp.value
factors	Examined	Positive (%)	_
Age group			
1 -10	61	20(32.8)	33.5 5 0.0012
11 -20	83	10(12.1)	
21 -30	104	5(4.8)	
31 -40	63	6(9.5)	
41 -50	40	2(5.0)	
>50	30	2(6.7)	
Sex			
Male	180	18(9.9)	1.2 1 0.283
Female	200	27(13.5)	
Occupational status			
Students	112	5(4.5)	
Unemployed	54	7(13.0)	
Farmers	43	14(32.6)	
Civil servants	63	3(4.8)	
Artisans	25	6(24.0)	
Business	84	10(11.9)	
Educational status			
Not educated	68	15(22.1)	11.6 2 0.003
Primary	70	5(7.1)	
Secondary	158	12(7.6)	
Tertiary	85	8(9.4)	

Table 2: Sero--Prevalence of *E. histolytica* among the study population based on risk factors

Risk Factors	Number	Number	x ² dfp.value
	Examined	Examined	

Source of water			
Bore hole	201	28(13.9)	49.3 2 0.003
Tap water	164	7(4.3)	
Well	16	10(62.5)	
Toilet facility			
Pit	26	10(38.5)	19.0 1 0.0012
Water cistern	355	35(9.9)	
Hand washing /personal hygiene			
Washes hands	281	25(8.9)	8.7 1 0.002
with soap		(0.5)	3.332
Washes Hand without soap	100	20(20)	
		. ,	
Contact with domestic			
animal/fecal matter			
Yes	256	40(15.6)	10.9 1 0.001
No	125	5(4.0)	
Season			
Dry	198	36(18.1)	16.1 1 0.014
Wet	183	9(4.9)	

RESULT

Out of the 381 subjects studied,45(11.8%) were sero-positive for *E.histolytica* infection, Based on Age, The age group 1-10 years of the study population had the highest prevalence 32.8%, while those in age group 40-50 years had the lowest prevalence rate of 4.8%. Statistical analysis showed the difference was significant (P=0.0012)(Table 1).

E.histolytica infection among the subjects studied showed no significant association (P=0.283) with respect to genderin the study population. The female subjects had a prevalence rate of 13.5% and the male subjects a prevalence of 9.9% (Table 1). The subjects whose occupation were farmers, had the highest prevalence rate of *E.histolytica* infection of 32.6% while the subjects whose occupation were students had the lowest prevalence rate of 4.5%. This difference was statistically significant(P=0.002)(Table 1). Subjects not educated had the highest prevalence rate of *E.histolytica* infection of 22.1%, while subjects with primary education level had the lowest the lowest prevalence rate of 7.1%. *E.histolytica* infection was statistically associated with educational status (P=0.0014), (Table 1).

Subjects whose source of drinking water is wells had the highest prevalence rate of 62.5% of *E.histolytica* infection-, while subjects whose source of drinking water is tap water and bore hole had the prevalence rates of 4.3% and 13.9%,respectively. This difference was significant (P=0.003),(Table 2). The highest prevalence rate

of *E.histolytica* infection was recorded in subjects that use pit latrine. There was significant association between the type of toilet facility and *E.histolytica* infection (P=0.0012)(Table 2). Subject that washed their hands without soap had the highest prevalence rate of 20.0%, while those that washed their hands with soap had a prevalence rate of 8.9%. This difference was significant(P=0.002) (Table 2). *E.histolytica* infection was associated with contact of domestic animal/FaecalMatter (P=0.001)(Table 2). The prevalence of *E.histolytica* infection among Participant in the study population was highest in the rainy season 18.1% than in the dry season 4.9%. The association of *E.histolytica* infection with seasons was significant (P=0.0014). (Table 2).

Discussion

E. histolytica infection predominates in developing countries and represents a major public health problem in tropical countries [21]. This study established a low sero-prevalence rate of *E.histolytica* infection (11.8%–) in the study population when compared with 22% and 21% reported in south Africa and Egypt respectively. In Nigeria, 21.6% in Enugu 14.3%–, in Kaduna and 13.7% in Ilesa have been reported [8,9,10,22]. Although, slightly higher than 10% reported by World health organization in developing countries [23]. The reasons for the disparity in the variation of prevalence rate of *E.histolytica* infection could be attributed togeographical, study design, seasonal, diagnostic methods, Patients selection and behavioral factors in the different study population.

The –age group 1-10 years were the most infected with *E.ntamoebahistolytica* infection with a prevalence rate –of 32.8% in agreement with –Zahida *et al.* (2010)[24]. In their study, reported –that age is an important risk factor for many infectious diseases especially those —that are transmitted orally such as *E. histolytica*(24,25). The current study findings are also in consistence with previous studies done in Pakistan and Bangladesh which reported that infection with *E. histolytica* is most common among young children who are predisposed to contact with infected material as they crawl on the ground or play games outdoors [5,25–] In addition, Children are less acquainted with hygiene habits—which also makes them more vulnerable to infection [21]. The result from this study, found people from all age groups were infected with –*E.histolytica*, Although, there is variation of prevalence rate among the age groups. This –is consistent with the results by Zahida*et al.* (2010) [24] The association of *E.histolytica* infection with age is in agreement with Al-Harthi and Jamjoom, (2007) and Zahida *et al.*(2010) [5,7]. who reported similar results.

The lack of significant association of *E.histolytica* infection with gender observed in this study is consistent with Dawah*et al*,2010 [22], attributed to equal exposure of both sex in the study area to the risk factors of *E.histolytica* infection. Although, prevalence of 13.5 % of *E.histolytica* infection in females was higher than male ,it is inconsistence with other previous studies on amoebiasis. Jamaiah and Shekhar(1999) and Stauffer *et al*(2006)[8,27] reported higher prevalence in Males.

The high prevalence rate reported in female could be attributed to women being culturally expected to be involved -in more domestic chores than males, These may bring them into constant contact with contaminated fruits, vegetables and water which potentially promote oral transmission of the disease through contaminated hands. This might account for the slightly higher level of infection among the female population similar to report by Haque *et al.* (2006) [5].

Interestingly, occupation was associated with an increase risk increase of *E.histolytica* infection. Farmers had the highest prevalence rate_—32.6% of *E.histolytica* infection. This is attributed to the nature of their occupation were they expose themselves with human and animal excreta especially when using it as manure. This findings from this study, is in contrast with Pham duc *et al*,(2011)[28]. In their study of the risk factors of *E.histolytica* in agricultural communities in Vietnam.

The non-educated -subject -had highest number of -*E. histolytica* -prevalence -rate 22.1% when compared with primary, secondary and tertiary educated subjects, Education was significantly associated with *E.histolytica* infection from this study. Education is regarded as one of the parameter of determining personal well-being especially in improving hygiene. Ross *et al.* (2003) argued that people with higher levels of educations tend to be healthier than those of similar income who are less educated because they seek medical attention early[29,30,31] found ignorance, toilet habit anddegree of literacy as serious risk factors for amoebiasis. This is consistence with our findings in this study.

The result from this study shows significant association of *E.histolytica* infection with source of drinking water. The association of *E. histolytica* with the source of drinking water of the patients agreed with the findings of Cox (2001), Olsen *et al*,(2001), Ogunlesi_*et al*,(2005) and Rinne*et al*,(2005)[10,32,33,34], all reported source of water as a risk factor for amoebiasis. Most of the subjects in this study obtained water from bore hole which are less likely to be contaminated with *E.histolytica* parasites when compared with well water. The -subject who -use well water -as source of drinking water had the highest prevalence -rate of *E.histolytica* infection. Most of the wells in the study area were manually dug, uncemented with

no casing or covering. Sometimes, the well is contaminated with surface run-off which may be faecally contaminated.

There was significant association between personal hygiene and *E.histolytica* infection, this is attributed to the route of transmission of *E.histolytica* infection which is faecal –oral transmited. This is in agreement with Espinosa-Cantellano and Espinosa-Cantellano 2000; Ryan and Ray, 2004, which reported similar findings[30,35]. Close contact with domestic animals/faecal matter was associated with an important risk factor for *E.histolytica* infection in this study. The subject with the hisghest prevalence of 15.6% are respondent with contact domestic animals/faecal matter. it is well possible that cysts of *Entamoeba histolytica* deposited on the surface (fur) of the animals during close contact with humans and then later transmitted to a –next person and facal matter with the parasite is transmitted faecal-orally.

The seasonal variation was observed in this study, there was association of *E. histolytica infection* with the season. higher prevalence rate of 18.1% *E. histolytica* infection was reported during the wet season in this study but slightly lower than 26.9% reported by Dawah *et al*,2010 in Kano[22], but consistent with some of the earlier studies of Park (2002) and Mawashi (2003),who reported similar prevalence rates in their studies[36,37]. Higher rate of fecal-oral contamination may be implicated during wet season and coincides with intensive farming activities. Low prevalence rate of parasite during the dry season is attributed high temperature and low humidity which is not favorable for parasite growth.

Conclusions To curb the relatively high levels of amoebiasis in Calabar there is need for surveillance systems and health education targeting parents and guardians of children under five years aimed at early and proper treatment of the disease. The Ministry of Health should intensify health campaign especially in children less than five years of age and their parents / guardians, particularly females, on ways to improve hygiene practices at home to avoid infection. There is need for residents in Calabar, Cross River State to emphasize use of safe water for all domestic chores if the benefit of personal hygiene is to be realized.

REFERENCES

1). Van Hal SJ, Stark DJ, Fotedar R, Marriott D, Ellis JT, Harkness JL. Amoebiasis: current status in Australia. *Med J Aust*. 2007;186 (8):412-416.

- 2). Choudhuri G, Rangan M. (2012)Amebic infection in humans. *Indian J Gastroenterol*; 31(4):153-162.
- 3). Ngui R, Ishak S, Chuen CS, Mahmud R, Lim Y.A(2011). Prevalence and risk factors of intestinal parasitism inrural and remote West Malaysia. *PLOS Negl. Trop. Dis.*; 5(3):e974. https://doi.org/10.1371/journal.pntd.0000974 PMID: 21390157
- 4). Lozano R, Naghavi M, Foreman K, Lim S, Shibuya K, Aboyans V,(2012). Global and regional mortalityfrom 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the Global Burden of Disease Study 2010. *The lancet*; 380(9859):2095–2128. https://doi.org/10.1016/S0140-6736(12)61728-0 PMID: 23245604
- 5) Haque, D., Duggal, P., Kabir M., Roy, S., Fair, B. M. (2006). *Entamoebahistolytica* infection in children and prevention from subsequent Amoebiasis. *Infect Immun.* 74: 904-909.
- 6)Oliveira FM, Neumann E, Gomes MA, Caliari MV(2015). *Entamoebadispar*: could it be pathogenic. *Trop Parasitol*.; 5(1):9–14. https://doi.org/10.4103/2229-5070.149887 PMID: 25709947
- 7). Chacin-Bonilla L(2013). [An update on amebiasis. Rev Med Chil.;141(5):609-615
- 8) Cui Z, Li J, Chen Y, Zhang L(2019). Molecular epidemiology, evolution, and phylogeny of *Entamoeba*spp. *InfectionGenetic and Evolution* 75:104018.https://doi.org/10.1016/j.meegid.2019.104018 PMID: 31465857
- 9)Ozumba, U. C. (1997). Antimicrobial susceptibility pattern and sero-group distribution of *Shigella*species at Enugu, Nigeria. *Post-Graduate Medicine Journal*. 4: 1-3.
- 10)Ogunlesi, T. A., Okeniyi, J. A. O., Oyedeji, O. A., Oseni, S. B. A., Oyelami, O. A., Njokonma, O. F. (2005)Childhood Dysentery in Ilesa, Nigeria: the unusual role of *E. histolytica*. *The Internet Journal of Tropical Medicine*.2. (2).
- 11). Norhayati M, Fatmah MS, Yusof S, Edariah AB: (2003)Intestinal parasitic infections in man: a review. *Med J Malaysia* 58:296-305.
- 12). Benetton ML, Goncalves AV, Meneghini ME, Silva EF, Carneiro M(2005): Risk factors for infection by the *Entamoeba histolytica/E. dispar* complex: an epidemiological study conducted in outpatient clinics in the city of Manaus, Amazon Region, Brazil. *Trans R Soc Trop Med Hyg*, 99:532-540.
- 13). Nair GV, Variyam EP.(2014) Noninvasive intestinal amebiasis: *Entamoeba histolytica* colonization without invasion. CurrOpin Infect Dis.;27(5):465-469.

- 14). Slack A.(2012). Parasitic causes of prolonged diarrhoea in travellers- diagnosis and management. AustFam Physician.41(10):782-786.
- 15). Petri WA, Haque R.(2013) Entamoeba histolytica brain abscess .HandbClin Neurol. 2013;114:147-152.
- 16). Singh A, Banerjee T, Kumar R, Shukla SK. (2019) Prevalence of cases of amebic liver abscess in a tertiarycare centre in India: A study on risk factors, associated microflora and strain variation of *Entamoebahistolytica.PloS one*.; 14(4):e0214880. https://doi.org/10.1371/journal.pone.0214880 PMID:30943253
- 17). Karanis P, Kourenti C, Smith H,(2007) Waterborne transmission of protozoan parasites: a worldwide review of outbreaks and lessons learnt. *J Water Health*.;5(1):1-38.
- 18)Ali IKM, Roy S.(2020) A Real-Time PCR Assay for simultaneous detection and differentiation of four common *Entamoeba* species that infect humans. *J. Clin. Microbiol.* 59(1), e01986–20. https://doi.org/10.1128/JCM.01986-20 PMID: 33115843
- 19) Sub-national HDI-Area Database (2021), HDI global database. Retreived December 12,2021
- 20), Naing, LWinn, T and Rusli, B. (2006) Practical issues in calculating the sample size for prevalence studies. *Journal of orofacial science*, 1:9-14
- 21 Stanley, S.L. Jr. (2003). Amoebiasis. The Lancet 361: 1025-1034
- 22)Dawah, I S,InaboH and Jatua E.D.(2010). Comparative study of microscopy with ELISA antibody based amoebiasis diagnosis in patient presenting with dysentery at government hospital in kaduna metropolis. *Continental Journal of Biomedical sciences*. 4:43-49
- 23)WHO (2015) Estimates of the global burden of foodborne diseases: foodborne disease burden epidemiology reference group 2007-2015.retrieved December 21
- 24)Zahida, T., Shabana, K. and Lashari, M.H. (2010). Prevalence of *Entamoebahistolytica*in humans, Pakistan. *Pakistan Journal of Pharmaceutical Sciences* **23**: 344-348.
- (25) Rinne, S., Rodas, E. J., Galer-until, R., Glickman, L.T (2005). prevalence and Risk factors for protozoan and nematodes infections among children in an Ecuadorian highland community. *Transaction of the Royal Society of Tropical Medicine*. 99: 585-92

- **26**) Al-Harthi, S.A. and Jamjoom, M.B. (2007). Diagnosis and Differentiation of Entamoeba infection in Makkah Al Mukarramah using microscopy and stool Antigen Detection Kits. *World Journal of Medical Sciences* **2**: 15-20.
- 27) Jamaiah I, Shekhar KC. (1999) Amoebiasis: A 10 year retrospectivestudy at the University Hospital, Kuala Lumpur. *Med J Malaysia* 1999;54:296-302.
- 28) Pham Duc, Hung Nguyen-Viet, Jan Hattendorf, JakobZinsstag, PhungDac Cam3 andPeter Odermatt (2011) Risk factors for *E histolytica* infection inan agricultural community in Hanam province, *Vietnam Parasites & Vectors*, 4:102http://www.parasitesandvectors.com/content/4/1/102
- 29)Ross, J.T., Blair, H.S., Edwin, R. and Van, T. (2003). Health and illness in the community Oxford University Press Inc. New York, USA.; p. 3-76.
- 30)Espinosa-Cantellano, M. and Espinosa-Cantellano, A. (2000). Pathogenesis of intestinal amoebiasis: from molecules to disease. *Journal of Clinical Microbiology* **13**: 318-331 31) Karaman, U., Atambay, M., Aycan, O., Yologlu, S. and Daldal, N.(2006). Incidence of intestinal parasites in municipal sanitary workers in Malatya. *Turkish Society for parasitology* **30**: 181-183
- 32)Ajonina C, Buzie C, Mo" ller J, OtterpohlR(2018). The detection of *Entamoebahistolytica* and *Toxoplasma gondii*in wastewater. *J. Toxicol. Environ.* 81(1–3):1–5. https://doi.org/10.1080/15287394.2017. 1392399 PMID: 29173133
- 33)Olsen, A., Samuelson, H., Onyango-Ouma (2001). Study of risk factors for intestinal helminths and protozoan infection using epidemiological approaches.?? *Journal Biosoc-Society*. 33: (4) 56 9-584.
- 34)Rinne, S., Rodas, E. J., Galer-until, R., Glickman, L.T (2005). prevalence and Risk factors for protozoan and nematodes infections among children in an Ecuadorian highland community. *Transaction of the Royal Society of Tropical Medicine*. 99: 585-92
- 35)Kosar S, Afshan K, Salman M, Rizvi S, Naseem AA, Firasat S,(2017). Prevalence and risk factors associated with intestinal parasitic infections among schoolchildren in Punjab, Pakistan. Trop Biomed. 770–780. PMID: 33592946

36)Mawashi, K. Y. (2003). The prevalence of intestinal parasites in some parts of Katsina State Unpublished *Masters'Thesis.Department of Microbiology, Ahmadu Bello University, Zaria.* 5-8529-9.

37)Park, K. (2002). Amoebiasis. In Park K. (ed). Park's Textbook of preventive and Soc. Med. 17th Edition. M/S BarnasidasBhanor Publishers, Jabalpur, India. 184-185.

