

Original Research Article
**Impact of soil biological parameters on soil
health in the intensively cultivated deltaic
Inceptisol of Tamil Nadu, Coimbatore.**

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ABSTRACT

An experiment was conducted during 2021-22 at Agricultural Research Station, Kattuthottam, Thanjavur, Tamil Nadu to identify the impact of biological parameters on different cropping sequences. Soil samples were ~~are~~ taken from different cropping sequences from the experimental plot during active vegetative stage. Samples were analysed for soil biological parameters viz., Urease, Acid phosphatase, Dehydrogenase, Soil microbial count (Bacteria, fungi and actinomycetes) and Microbial Biomass Carbon. Different cropping sequences showed their effect as variations in soil biological properties. The cropping sequence T₄, sunhemp-rice+dhaincha(10:1)-green gram showed more biological activity with urease activity of 40.6 NH₄ µg/g/h, acid phosphatase activity of 43.1 P-NP µg/g/h, dehydrogenase activity (137.9 TPF µg/g/day), microbial biomass carbon value (307 mg kg⁻¹), bacterial count (55.6 cfu g⁻¹ soil), fungal count (23.5 cfu g⁻¹ soil) and actinomycetes count (41.2 cfu g⁻¹ soil). Rice-rice-sesame sequence was observed to have less biological activity than other cropping sequences.

Keywords: *Biological properties, cropping sequence, ~~enzyme activity, microbial biomass carbon, microbial population,~~ soil health*

1. INTRODUCTION

'Soil health' is described as a condition of dynamic balance between flora and fauna and their surrounding soil environment, in which all the former's metabolic functions run smoothly without any interference, impedance from the latter [1]. Soil health is the ability of soil to continue and operate as a vibrant living ecosystem that supports plants, animals and humans. Soil health is a word used frequently in conversations about sustainable agriculture to indicate the overall state and quality of the soil resource. We have clean air and water, abundant crops and forests, productive grazing grounds, diversified fauna and attractive vistas because of healthy soil. All agricultural systems require management to function properly. Despite this, there is evidence of extensive damage of the environment due to erosion and degradation of organic matter in agricultural soils. Pollution, compaction and salinity increasing as well as other negative consequences.

Comment [up1]: Introduce the link between paragraph 1 and 2. For example, plant cover affects soil biological properties via quality of root exudates and decomposition of litter.

Rice (*Oryza sativa*) is a component of widely varying cropping sequences. Rice-based cropping sequence is an integral part of agriculture in Tamil Nadu. Several intense rice-based cropping sequences have been noticed and are being practiced by the farmers. While intensive agriculture, those involving exhaustive high yielding varieties of rice and other crops, has led to heavy withdrawal of nutrients from the soil by crops, imbalanced and indiscriminate use of chemical fertilizers has resulted in deterioration and degradation of soil health [2]. According to the FAO, global rice demand in 2020 would be 800 MT, whereas current output is 600 MT. To satisfy future requirements, an extra 200 million tonnes must be generated by boosting productivity per hectare [3]. Even when rice is planted with appropriate N, P and K, the productivity of the crop is dropping. Previously conducted experiment in the same established field ~~which~~ comprising of seven different crop rotations viz, rice-black gram, rice-sesame, rice+dhaincha-maize+green gram, rice+dhaincha-bhendi, rice+dhaincha-ragi, rice+dhaincha-varagu, rice+dhaincha-fodder cowpea with first crop of green manure in all rotations. The results revealed that crop rotations had a significant impact on soil quality by altering their properties [4]. Suitable rice-based cropping sequence has to be evaluated, to assess the stability in production. Hence, ~~in with~~ this context, a research work was carried out to evaluate the soil biological properties in different cropping sequences in order to ascertain the health of cultivated soil.

2. MATERIALS AND METHODS

2.1. Study site

The present investigation was carried out at Department of Soil Science and Agricultural Chemistry at Tamil Nadu Agricultural University, Coimbatore. For this study, the soil samples were collected from Agricultural Research Station, Thanjavur which is 272 km away from the Coimbatore in the eastern direction. The soil initial analysis showed it was sandy loam soil with 10 cropping sequences replicated thrice in Randomized Block Design. The area of each plot was is about 40 m². Samples were collected at active vegetative stage from the root zone at 0-15 cm which were considered as surface samples, during *kharif*, *rabi* and summer seasons of 2021-22. The cropping sequences were established with recommended dosage of fertilizer. The details of the cropping sequences are as follows (Table 1)

Table 1. List of treatments in cropping sequence

Treatments	Kharif	Rabi	Summer
T ₁	Rice (Co 51)	Rice (ADT 46)	Black gram (ADT 5)
T ₂	Rice (Co 51)	Rice (ADT 46)	Sesame (VRI 2)
T ₃	Dhaincha	Rice+dhaincha (5:1) (ADT 46)	Black gram (ADT 5)
T ₄	Sun hemp	Rice+dhaincha (10:1) (ADT 46)	Green gram (Co 6)
T ₅	Black gram (ADT 5)	Rice (ADT 46)	Groundnut (TMV 13)
T ₆	Green gram (Co 6)	Rice (Bio fortified-CR Dhan45)	Sesame (VRI 2)
T ₇	Fodder Cowpea Co(FC) 9	Rice (ADT 46)	Fodder Maize (African tall)
T ₈	Fodder Sorghum Co(FS) 29	Rice (Co 52)	Fodder Cowpea Co(FC) 9
T ₉	Maize hybrid (Co 6)	Rice (Seeraga samba)	Bhendi (Co 4)+black gram (ADT 5) (5:1)
T ₁₀	Ragi (Co 15)	Rice (Navara)	Cluster bean (MDU1)+black gram (ADT 5) (5:1)

Comment [up2]: What is the GPS coordinate of the location?

Comment [up3]: Do you mean Randomized Block Design ie RBD? RBD is the design on the field but the Design in the laboratory should be Completely Randomised Design(CRD)

2.2. Soil analysis

The soil analysis was carried out at the laboratory of Department of Soil Science and Agricultural Chemistry in Tamil Nadu Agricultural University, Coimbatore. Urease was determined by calculating µg of TPF /g/day [5]. Acid Phosphatase enzyme activity was determined using *p*-nitro phenyl phosphate as substrate colorimetrically at 410 nm [6]. The Dehydrogenase of the soil was determined as µg of NH⁴⁺ released /g/hr [7]. Microbial population (bacteria, fungi and actinomycetes) of soil were determined by [8] [9] [10] and [11] in cfu g⁻¹ soil. Microbial biomass carbon of the soil sample was analysed by using fumigation extraction technique [12].

Comment [up4]: For Table 1, remove all vertical lines. Secondly, remove all horizontal lines, except the base and 2 top horizontal lines

Comment [up5]: Foot notes should be here to describe the abbreviations on the treatments

Comment [up6]: should be NH₄⁺

3. RESULTS AND DISCUSSION

3.1. Urease and Acid Phosphatase

From Table 2, the values of urease activity in the soil ranged from 18.6 to 39.8 NH₄ µg/g/h with a mean value of 29.8 NH₄ µg/g/h during the *kharif* season, the higher value was recorded under T₄ cropping sequence and the lower value was recorded under T₉ cropping sequence. In *rabi* season, the values of urease activity of the soil ranged from 22.9 to 40.6 NH₄ µg/g/h with a mean value of 30.4 NH₄ µg/g/h. The higher value of urease activity present under T₄ cropping sequence and the lower present under T₉ cropping sequence. During summer season values of urease activity of the soil ranged from 16.0 to 31.1 NH₄ µg/g/h with a mean value of 24.5 NH₄ µg/g/h. The highest urease value was recorded in T₄ cropping sequence whereas the lowest value was found in T₂ cropping sequence. The inclusion of green manure crop and legume black gram in the T₄ cropping sequence may have caused the greatest urease activity to be detected. This crop may have fixed more N than in the T₉ crop. This is in line with [13]. The inclusion of readily decomposable organic materials did not hinder abiotic urease's capacity to hydrolyze urea, and thus considerably enhanced the microbiological production of urease by soil microorganisms [14].

Acid phosphatase activity of the soil samples ranged from 21.4 to 27.5 P-NP µg/g/h with a mean value of 25.1 P-NP µg/g/h during the *kharif* season, the higher value was recorded under T₄ cropping sequence and the lower value was recorded under T₂ cropping sequence. In *rabi* season, the values of acid phosphatase activity of the soil ranged from 19.2 to 33.6 NH₄ P-NP µg/g/h with a mean value of 26.3 NH₄ P-NP µg/g/h. The higher value of acid phosphatase activity present under T₄ cropping sequence and the lower present under T₂ cropping sequence. During summer season, values

Comment [up7]: what is the implication of having high or higher urease activity? Is it a sign of nitrogen gain or loss, nitrogen availability or unavailability? High urease activity in an acidic soil means soil N gain but in a basic or calcareous soil, it means soil N loss in the form of ammonia. The soil used for the experiment should have been analysed for Organic carbon, Total Nitrogen, Available P and pH. The pH of the soil would have been used to know whether increase in urease activity was a gain or loss in Nitrogen.

of acid phosphatase activity of the soil ranged from 25.5 NH₄ to 43.6 NH₄ P-NP µg/g/h with a mean value of 33.5 NH₄ P-NP µg/g/h. The highest acid phosphatase activity was recorded in T₄ cropping sequence whereas the lowest value was found in T₉ cropping sequence. Legumes in agricultural rotations may have generated more acid phosphatase enzymes than non-legume crops. Furthermore, compared to legumes and green manure crops, cereals did not require as much P for the symbiotic N fixation process [15] [16].

Table 2. Effect of various cropping sequence on Urease and Phosphatase

Cropping sequence	Urease (NH ₄ µg/g/h)			Acid Phosphatase (P-NP µg/g/h)		
	Kharif	Rabi	Summer	Kharif	Rabi	Summer
T ₁	39.8	40.6	31.1	24.2	30.1	39.3
T ₂	19.9	24.0	16.0	21.4	19.2	27.8
T ₃	35.3	35.1	29.5	25.6	32.4	41.9
T ₄	38.4	38.7	30.2	27.5	33.6	43.6
T ₅	32.1	27.5	28.3	26.9	22.5	37.4
T ₆	31.7	29.2	26.9	24.7	21.7	36.1
T ₇	29.2	30.4	19.7	24.5	29.8	35.5
T ₈	28.0	30.8	23.6	26.7	27.3	29.0
T ₉	18.6	22.9	18.1	23.9	20.8	25.5
T ₁₀	25.6	25.6	21.8	25.1	25.9	27.4
Mean	29.8	30.4	24.5	25.1	26.3	33.5
SEd	0.72	0.61	0.59	0.62	0.63	0.62
CD (0.05)	1.52	1.29	1.24	1.32	1.33	1.31

Comment [up8]: With the exception of swampy rice in association with Anabaena, nostoc and Beijerinckia Algae, cereals hardly have symbiotic associations for the purpose of supplying Nitrogen symbiotically. Cereals have symbiotic associations with root fungi known as arbuscular mycorrhizal fungi for the purpose of supplying phosphorus

Comment [up9]: Weak Argument. If legumes are generating higher acid phosphatase activity, it shows that the soil looses much phosphorus to legumes than cereals.

Comment [up10]: For Table 2, except for the vertical line that separated Urease from Acid Phosphatase, remove all vertical lines. Secondly, remove all horizontal lines, except the bottom and first 3 top horizontal lines

3.2. Dehydrogenase and Microbial biomass carbon

From Table 3, the values of dehydrogenase enzyme of the soil ranged from 21.1 to 33.6 TPF µg/g/day with a mean value of 27.9 NH₄ TPF µg/g/day during the *kharif* season, the higher value was recorded under T₄ cropping sequence and the lower value was recorded under T₈ cropping sequence. In *rabi* season, the values of dehydrogenase in the soil ranged from 42.7 to 137.9 TPF µg/g/day with a mean value of 81.3 TPF µg/g/day. The higher value of dehydrogenase activity present under T₄ cropping sequence and the lower value is present under T₂ cropping sequence. During summer season, values of dehydrogenase value in the soil ranged from 23.2 to 36.4 TPF µg/g/day with a mean value of 29.3 TPF µg/g/day. The highest dehydrogenase activity was recorded in T₄ cropping sequence whereas the lowest value was found in T₁₀ cropping sequence. The activity was brought on about by the presence of legume and green manure crop in the cropping sequence, which improved organic matter addition and root exudation [17]. *Rabi* season dehydrogenase levels were greater, which may be related to the increased organic matter storage caused by the cooler temperatures and favourable dehydrogenase activity-promoting conditions. Additionally, [18] noted that autumn (wet) had greater dehydrogenase activity than dry season.

In *kharif* season, microbial biomass carbon of the soil samples ranged from 223 to 279 mg kg⁻¹ with a mean value of 250 mg kg⁻¹, the higher value was recorded under T₄ cropping sequence and the lower value was recorded under T₂ cropping sequence. In *rabi* season, the values of microbial biomass carbon of the soil ranged from 245 to 307 mg kg⁻¹ with a mean value of 273 mg kg⁻¹. The higher value of microbial biomass carbon present under T₄ cropping sequence and the lowest present under T₂ cropping sequence. During summer season, values of microbial biomass carbon of the soil ranged from 152 to 269 mg kg⁻¹ with a mean value of 213 mg kg⁻¹. The highest microbial biomass carbon was recorded in T₄ cropping sequence whereas the lowest value was found in T₉ cropping sequence. Compared to other fractions of organic matter the turnover rate of soil microbial biomass carbon is rapid and higher due to microbial activity [19]. Cropping sequence had the significant impact on the soil microbial carbon.

Comment [up11]: There is a direct relationship between soil moisture and dehydrogenase activity. Soil organic matter that increases soil moisture retention will increase soil dehydrogenase activity.

Comment [up12]: Yes, but you have not explained your result adequately. why are you having more microbial biomass carbon under legume sequence than cereal sequence? This is due to higher microbial activity that will eventually lower C : N ratio and increase the availability of Nitrogen to plants

Table 3. Effect of various cropping sequences on soil dehydrogenase activity and soil microbial biomass carbon

Cropping sequence	Dehydrogenase (TPF µg/g/day)			Microbial biomass carbon (mg kg ⁻¹)		
	Kharif	Rabi	Summer	Kharif	Rabi	Summer
T ₁	23.5	50.8	24.6	237	251	189
T ₂	30.4	42.7	34.6	223	245	169
T ₃	32.8	112.0	33.1	241	295	254
T ₄	33.6	137.0	36.4	279	307	269

T ₅	28.3	74.2	32.0	265	287	237
T ₆	26.2	92.4	30.1	267	278	234
T ₇	29.6	89.3	27.7	259	274	222
T ₈	21.1	63.0	26.9	234	256	217
T ₉	24.8	44.6	25.3	243	263	152
T ₁₀	28.9	106.0	23.2	253	280	195
Mean	27.9	81.3	29.3	250.1	273.6	213.8
SEd	0.59	1.34	0.45	5.15	6.09	5.44
CD (0.05)	1.24	2.83	0.96	10.81	12.79	11.42

Comment [up13]: For Table 3, except for the vertical line that separated dehydrogenase from Microbial biomass carbon, remove all vertical lines. Secondly, remove all horizontal lines. except the base and 3 top horizontal lines

3.4. Soil microbial population (Bacteria, fungi and actinomycetes)

From Fig. 1, the bacterial population in the soil ranged from 41.1 to 50.1 cfu g⁻¹ soil with a mean value of 45 cfu g⁻¹ soil during the *kharif* season, the higher value was recorded under T₄ and the lower value was recorded under T₂. In *rabi* season, the values of bacterial population of the soil ranged from 43.4 to 55.6 cfu g⁻¹ soil with a mean value of 49 cfu g⁻¹. The higher value of bacterial population was is present under T₄ cropping sequence and the lower value was present under T₂ cropping sequence. During summer season, values of bacterial population of the soil ranged from 39.2 to 48.7 cfu g⁻¹ with a mean value of 44.8 cfu g⁻¹. The highest bacterial population was recorded in T₄ cropping sequence whereas the lowest value was found in T₈ cropping sequence. Fungal population of the soil samples ranged from 10.8 to 19.6 cfu g⁻¹ with a mean value of 15.1 cfu g⁻¹ during the *kharif* season, the higher value was recorded under T₄ cropping sequence and the lower value was recorded under T₂ cropping sequence. In *rabi* season, the values of fungal population of the soil ranged from 12.3 to 23.5 cfu g⁻¹ with a mean value of 17.7 cfu g⁻¹. The higher value of fungal population was is present under T₄ cropping sequence and the lower value was present under T₂ cropping sequence. During summer season, values of fungal population of the soil ranged from 9.2 to 16.1 cfu g⁻¹ with a mean value of 12.6 cfu g⁻¹. The highest fungal population was recorded in T₄ cropping sequence whereas the lowest value was found in T₉ cropping sequence. Actinomycetes population of the soil samples ranged from 33.7 to 38.5 cfu g⁻¹ with a mean value of 36 cfu g⁻¹ during the *kharif* season, the higher value was recorded under T₄ cropping sequence and the lower value was recorded under T₂ cropping sequence. In *rabi* season, the values of actinomycetes population of the soil ranged from 36.9 to 41.2 cfu g⁻¹ with a mean value of 39 cfu g⁻¹. The higher value of actinomycetes population was present under T₄ cropping sequence and the lower value was present under T₂ cropping sequence. During summer season, values of actinomycetes population of the soil ranged from 31.1 to 35.4 cfu g⁻¹ with a mean value of 33.7 cfu g⁻¹. The highest actinomycetes value was recorded in T₄ cropping sequence whereas the lowest value was found in T₉ cropping sequence. In this study, the legume-dominated sequence with green manure crop had a higher microbial population than the cereal-dominated system because the former system was better for the growth and development of the soil microorganisms. This could be because bacteria and the nodules found in the roots of the leguminous crops have a symbiotic interaction. Legumes may also enhance SOC, which is good for the growth of microorganisms, and their roots produce sugar-like compounds that aid in the growth of soil bacteria in the rhizosphere [20] [21]. Legume-based cropping methods boost microbiological and enzymatic activity because they supply high-quality biomass (low C:N ratio) to the soil through active root development and exudation, nodule degeneration and leaf shedding. Different environmental conditions in the root zone are brought about by the legume systems, which have an impact on nutrient uptake and carbon exudation [22].

Comment [up14]: Too lengthy result. why not discuss as you present each result than waiting to give a general discussion at the end of the whole result? note that bacteria can use 2 sources of carbon: organic and inorganic carbon. on the other hand, fungi and actinomycetes can use only organic carbon source. Bacteria are also more saprophytically more competent than fungi and actinomycetes in that sequence. Any cropping sequence that has the highest population of fungi is an indication that it has more organic than inorganic carbon and that it is a source of not easily decomposable organic material. Conversely, cropping sequence with higher bacteria population shows that it is a rich source of easily decomposable organic material. Due to their structure and C:N ratio between 7:1 and 25:1, fungi need a greater amount of carbon to grow and reproduce and will therefore collect the required from the soil organic matter. Bacteria, which have a smaller C:N ratio than fungi (between 5:1 and 7:1), need food rich in nitrogen like green manure, legume residues etc because they have higher nitrogen requirement and take more nitrogen from the soil for their own requirements.

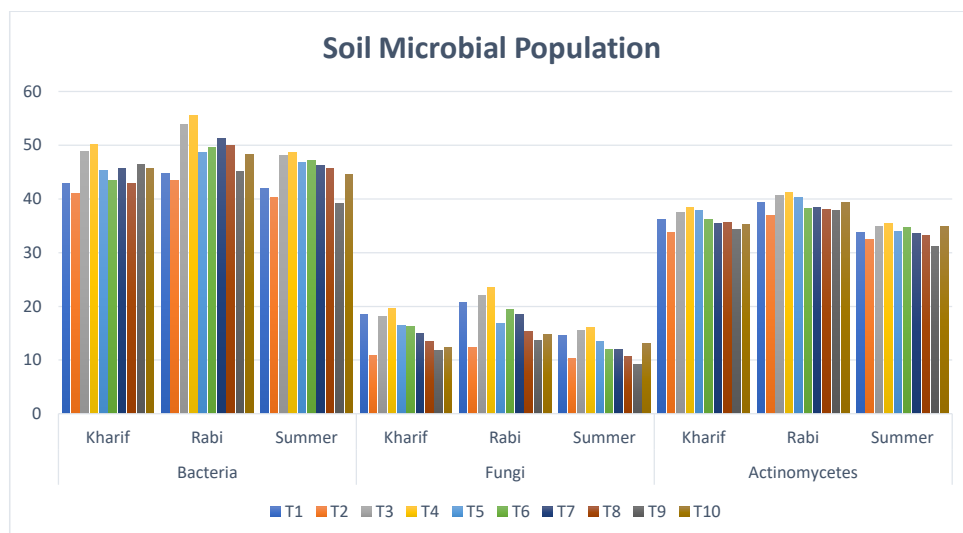


Fig. 1. Effect of various cropping sequences on soil microbial populations (bacteria, fungi and actinomycetes)

Comment [up15]: This Figure 1 cannot be described well without error bars. Please retake the bar chart.

4. CONCLUSION

From the investigation, it was observed that the soil biological parameters differ significantly due to the effect of various cropping sequences. It was observed that, most of the parameters show higher values in *rabi* season than *kharif* and summer. Under various cropping sequences taken for research work, sunhemp-rice+dhaincha(10:1)-green gram cropping sequence shows higher fertility status than other cropping sequences as it contains higher biological activity and better soil properties than others due to inclusion of green manure and legume crop. The cropping system rice-rice-sesame sequence shows minimum value in most of the soil biological properties than other cropping sequence and should therefore be avoided if the soil must be healthy.

REFERENCES

- Goswami, Rattan RK. Soil Health- Key to sustained agriculture productivity. Fertiliser News. 1992;37(123), 53-60.
- John PS, George M, Jacob R. Nutrient mining in agro – climatic zones of Kerala. Fertilizer News. 2001;46: 45 – 52, 55 – 57.
- Swaminathan MS. An Evergreen Revolution, Crop Sci.2006;46: 2293-2303.
- Gunasekaran Yazhini, Kaliappan Sathiya, Porpavai Shanmugasundaram. Developing Soil Quality Indices for different crop rotations of Deltaic Inceptisol regions of India. Communications in Soil Science and Plant Analysis. 2021;52. 10.1080/00103624.2021.1885683.
- Tabatabai MA, Bremner JM, Assay of Urease Activity in Soils. American Journal of Soil Science Society. 1972;41, 350-352.
- Tabatabai MA, Bremner, JM. Use of p-nitrophenol phosphate for the assay of soil phosphatase activity. Soil Biology Biochemistry. 1969;1, 301-307.
- Casida L, Klein D, Santoro T. Soil Dehydrogenase Activity. Soil Science, 1964; 98, 371-376.
- Wright HD. "The Importance of Adequate Reduction of Peptone in the Preparation of Media for the Pneumococcus and Other Organisms". The Journal of Pathology and Bacteriology. 1933;37 (2): 257–282.
- Mossel DA, AM Kleynen-Semmeling, HM Vincentie, H Beerens, M Catsaras. "Oxytetracycline-Glucose-Yeast Extract Agar for Selective Enumeration of Moulds and Yeast in Foods and Clinical Material". The Journal of Applied Bacteriology. 1970;33 (3): 454–457.
- Kenknight G, JH Muncie. "Isolation of Phytopathogenic Actinomycetes". Phytopathology. 1939;29: 1000–1002.
- Aryal, S. Spread plate technique-principle, procedure and uses. 2017.

Comment [up16]: upgrade reference

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12. Jenkinson DS, Powlson DS. The Effects of Biocidal Treatments On Soil. V: A Method For Measuring Soil Biomass. *Soil Biol, Biochem.* 1976;8:209-213.
13. Roldan A, Caravaca F, Hernandez MT, Garcia C, Sanchez-Brito C, Velasquez M, Tiscareno M. No-tillage, crop residue additions, and legume cover cropping effects on soil quality characteristics under maize in Patzcuaro watershed (Mexico). *Soil and Tillage Research.* 2003 Jul 1;72(1):65-73.
14. Surucu A, Ozyazici MA, Bayrakli B, Kizilkaya R. Effects of green manuring on soil enzyme activity. *Fresenius Environmental Bulletin.* 2014;23(9):2126-32.
15. Makoi JH, Ndakidemi PA. Selected soil enzymes: examples of their potential roles in the ecosystem. *African Journal of Biotechnology.* 2008;7(3).
16. Maseko ST, Dakora FD. Rhizosphere acid and alkaline phosphatase activity as a marker of P nutrition in nodulated *Cyclopia* and *Aspalathus* species in the Cape fynbos of South Africa. *South African journal of botany.* 2013 Nov 1; 89:289-95.
17. Babu YJ, Li C, Frolking S, Nayak DR, Datta A, Adhya TK. Modelling of methane emissions from rice-based production systems in India with the denitrification and decomposition model: field validation and sensitivity analysis. *Current Science.* 2005 Dec 10;1904-12.
18. Zhang WJ, Wang XJ, Xu MG, Huang SM, Liu H, Peng C. Soil organic carbon dynamics under long-term fertilizations in arable land of northern China. *Biogeosciences.* 2010 Feb 2;7(2):409-25.
19. Smyrna. Impact of different cropping systems on soil carbon pools and carbon sequestration. M.Sc., Agri (soil science) thesis, Tamil Nadu Agricultural University, Coimbatore 2016.
20. Narula, Neeru, Erika Kothe, Rishi Kumar Behl. "Role of root exudates in plant-microbe interactions". *Journal of Applied Botany and Food Quality.* 2012;82.2: 122-130.
21. Porpavai S, Devasenapathy P, Siddeswaran K, Jayaraj T. Impact of various rice-based cropping systems on soil fertility. *Journal of Cereals and Oilseeds.* 2011;2(3):43-46.
22. Alvey S, Bagayoko M, Neumann G, Buerkert A. Cereal/ legume rotation in two West African soils under controlled conditions. *Plant Soil.* 2000;231:45–54.

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