

## Original Research Article

Changes in activity of phenylalanine ammonia lyase (PAL), phenols and  $\beta$  1, 3 Glucanase in crossandra plants affected by *Fusarium incarnatum* (Desm.)Sacc and treated with bioagents, organic amendments, silver nanoparticles and fungicide in pot culture

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### **ABSTRACT**

Laboratory and pot culture experiments were conducted at Agricultural college and Research institute, Madurai, to study the effect of different bioagents, organic amendments, silver nanoparticles and fungicide in management of crossandra wilt caused by *Fusarium incarnatum* and nematode *Pratylenchus. delattrei*. During the study observations are made on changes in activity of phenylalanine ammonia lyase (PAL), phenols and  $\beta$  1, 3 Glucanase in crossandra plants. All the treatments applied recorded increased levels of phenylalanine ammonia lyase (PAL), phenols and  $\beta$  1, 3 Glucanase in treated plants, but among all soil application (SA) of *T.viride* @ 2.5 kg/ha at 20 DAP plus soil drenching (SD) of carbendazim @ 0.1% at 30 DAP plus SA of *T.viride* @ 2.5 kg/ha at 50 DAP plus Foliar application (FA ) of *P. fluorescens*@ 1.0 kg/ha at 70 DAP plus Foliar application (FA ) of *B. subtilis* @ 1.0 kg /ha at 90 DAP was found to be significantly higher in induction of phenylalanine ammonia lyase (PAL), phenols and  $\beta$  1, 3 Glucanase in crossandra plants. The same results are also noticed in later stage in reducing disease incidence compare to control and other treatments.

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Key words : Crossandra, *Fusarium spp*, phenylalanine ammonia lyase, phenols,  $\beta$  1, 3 Glucanase

## INTRODUCTION

Crossandra (Fire cracker) is an important commercial flower, mainly grown in India, Tropical Africa and Madagascar (Bailey, 1963). Crossandra belongs to the family of Acanthaceae and Native of Southern India and Sri Lanka. It is an erect, evergreen sub shrub growing to 1 meter with glossy, wavy-margined leaves and fan-shaped flowers, which may appear at any time throughout the year and easy to grow. It can be cultivated very conveniently by small farmers.

The common name "firecracker flower" refers to the seed pods, which are found after the flower has dried up, and tend to "explode" when near high humidity or rainfall. The "explosion" releases the seeds onto the ground, thereby creating new seedlings. It is popularly known as Kanakambaram in Tamil, Malayalam and Telugu and Kanakambara in Kannada. In Maharashtra it is known as Aboli. There are 50 species but only a few like *Crossandra infundibuliformis*, *C. guineensis*, *C. nilotica*, *C. muronata* and *C. subacaulis* are cultivated.

The flowers are commonly used for hair adornment. Though not fragrant, these flowers are very popular because of their attractive bright colour, light weight and good keeping quality. The flowers are used for making garlands, either alone or in combination with jasmine flowers. Steady market demand as well as assured and regular income have made crossandra a profitable venture for south Indian farmers.

Crossandra (*Crossandra infundibuliformis*) is affected by various fungal, bacterial, nematode and viral diseases. Among the various fungal diseases wilt disease caused by *Fusarium spp.* is one of the major problem in Crossandra production and limits the crop cultivation and it is also always associated with nematode such as *Pratylenchus delettrei* (Srinivasan and Muthukrishnan, 1975.) Management of this disease has become very difficult due to its soil borne and complex nature.

Many scientists reported that different methods such as use of biological agents/chemicals/bio technological/Physical and cultural methods are available to control/manage the plant diseases. Integrated approach for the management of disease involves the use of two or more methods in a compatible manner to reduce disease incidence with minimal hazards on environment and maximization of economic profit. Biocontrol agents, organic amendments, chemicals and more recently nanoparticles etc, can be combined to develop

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an efficient, eco-friendly, compatible and profitable disease management strategy that conserves natural resources and beneficial microbes. In all the methods of disease control, activation of defense mechanisms are very important for effective disease management. . Kamalakannan (2004) reported that soil application of bio control agents such as *Trichoderma* species and bacterial isolates like *P. fluorescens* induced coleus plants to synthesize more amount of Peroxidase, Polyphenol oxidase, Phenylalanine ammonia - lyase and total phenols. PAL is the key enzyme in inducing synthesis of salicylic acid (SA) which induces systemic resistance in many plants. Seed treatment and seedling root dipping with PGPR induced early and enhanced levels of PAL in rice plants (Nayar, 1996). Cahill and McComb (1992) observed that the inoculation of resistant variety of eucalyptus with *Phytophthora cinnamomi* resulted in rapid accumulation of phenolics within 24h. M'Piga *et al.* (1997) reported that application of *P. fluorescens* strain-28 brought about cell wall thickening, deposition of phenolic compounds and formation of callose resulting in restricted growth of *F. oxysporum* f. sp. *Lycopersici*. The enzyme  $\beta$ -1, 3-glucanase in disease resistance was first reported by Kauffmann *et al.* (1987). Daugrois *et al.* (1992) reported a rapid induction of two  $\beta$ -1, 3-glucanases in the incompatible interaction between bean and *C. lindemuthianum*.

So keeping in view, the role of defense enzymes in disease management, pot culture studies were carried to observe the changes in levels of defense enzymes in treated crossandra plants and disease incidence at Department of Plant Pathology, Agricultural College and Research Institute, Madurai, TNAU.

## **MATERIALS AND METHODS**

The local crossandra cultivar growing in the Madurai area was used in all pot culture experiments. The pathogen (*F. incarnatum*) mass multiplied on sand maize medium was incorporated in the pots at 3 per cent (w/w) and nematode was inoculated @ 1 nematode per gram of soil. All the treatments like biagents, nanoparticles, organic amendments and chemicals were applied as per schedule. The observations on enzyme studies were carried out after last application as mentioned in the treatment schedule. The leaves were collected from the pots 0, 3, 5, 7 and 9 days after last application in each treatment separately and washed several times with sterile distilled water before enzyme

extraction. Each treatment was replicated six times with each replication containing three pots with two plants in each pot. The treatment details are as follow

List 1: The treatment

T <sub>1</sub>	SA of <i>P.fluorescens</i> (Pf-18) @ 2.5 kg/ha at 20 DAP + Module A
T <sub>2</sub>	SA of <i>T.v</i> (Tv-9) @ 2.5 kg/ha at 20 DAP + Module A
T <sub>3</sub>	SA of <i>B. subtilis</i> (Bs-10) @2.5 kg/ha at 20 DAP + Module A
T <sub>4</sub>	SA of Neem cake @ 250 kg/ha at 20 DAP + Module A
T <sub>5</sub>	SD of carbendazim @ 0.1% at 20 DAP + Module A
T <sub>6</sub>	SA of Phorate10G @10 kg/ha at 20DAP + Module A
T <sub>7</sub>	FA of nano particles @ 800 ppm at 20 DAP + Module A
T <sub>8</sub>	SA of <i>P.f</i> (Pf-18) @ 2.5 kg/ha at 20 DAP + Module B
T <sub>9</sub>	SA @ <i>T.v</i> (Tv-9) @ 2.5 kg/ha at 20 DAP + Module B
T <sub>10</sub>	SA of <i>B.s</i> (Bs-10) @ 2.5 kg/ha at 20 DAP + Module B
T <sub>11</sub>	SA of Neem cake @ 250 kg/ha at 20 DAP + Module B
T <sub>12</sub>	SD of carbendazim @ 0.1% at 20 DAP + Module B
T <sub>13</sub>	SA of Phorate10G @10 kg/ha at 20DAP + Module B
T <sub>14</sub>	FA of nano particles @ 800 ppm at 20 DAP + Module B
T <sub>15</sub>	SA of <i>P.f</i> (Pf-18) @ 2.5 kg/ha at 20 DAP + Module C
T <sub>16</sub>	SA <i>T.v</i> (Tv-9) @ 2.5 kg/ha at 20 DAP + Module C
T <sub>17</sub>	SA of <i>B.s</i> (Bs-10) @ 2.5 kg/ha at 20 DAP + Module C
T <sub>18</sub>	SA of Neem cake @ 250 kg/ha at 20 DAP + Module C
T <sub>19</sub>	SD of carbendazim @ 0.1% at 20 DAP + Module C
T <sub>20</sub>	SA of Phorate10G @10 kg/ha at 20DAP + Module C
T <sub>21</sub>	FA of nanoparticles @ 800 ppm at 20 DAP + Module C
T <sub>22</sub>	Inoculated control (Pathogens and nematode)

Module A.= Foliar application (FA) of nano particles @ 800ppm at 30 DAP, SA of *T. viride* (Tv-9) @ 2.5 kg/ha at 50 DAP + FA of *P.f* (Pf-18) @ 1.0 kg/ha at 70 DAP + FA of *B.s* (Bs-10) @ 1.0 kg/ha at 90 DAP

Module B = FA of carbendazim @ 0.1% at 30 DAP + SA of *T.v* @ 2.5 kg/ha at 50 DAP and FA of *P.f* (Pf-18) @ 1.0 kg/ha at 70 DAP + FA of *B.s* (Bs-10) @ 1.0 kg/ha at 90 DAP

Module C= SD of carbendazim @ 0.1% at 30 DAP + SA of *T.v* @ 2.5 kg/ha at 50 DAP + FA of *P.f* (Pf-18) @ 1.0 kg/ha at 70 DAP + FA of *B.s* (Bs-10) @ 1.0 kg/ha at 90 DAP

#### Assay of Phenyl Alanine Ammonia Lyase (PAL)

Five hundred mg of leaf collected from each treatment was homogenized in five ml of cold 25 mM borate HCl buffer (pH 8.8) containing 5 mM mercaptoethanol, 0.4 ml per liter separately. The homogenate was centrifuged at 15,000 rpm for 15 minutes and the supernatant obtained from the samples of each treatment was used as enzyme source separately. The assay mixture consists of 0.2 ml of enzyme extract, 1.3 ml water and 0.5 ml borate buffer. The reaction was initiated by the addition of one ml of 12 mM L-Phenylalanine. The reaction mixture was incubated for one h at 32°C. The reaction was stopped by the addition of 0.5 ml of 2N HCl. A blank was run in which phenylalanine was added after adding 2N HCl. The absorbance was measured at 290 nm. The enzyme activity was expressed as  $\mu\text{mol}$  of cinnamic acid/ minute/ g of leaf (Dickerson *et al.*, 1984).

#### Assay of Phenols

One g of the leaf sample was ground in a pestle and mortar in 10 ml of 80 per cent methanol. The homogenate was centrifuged at 10,000 rpm for 20 minutes. The supernatant was evaporated to dryness and the residue was dissolved in five ml of distilled water. From this, 0.2 ml was taken and the volume was made up to three ml with distilled water. To that 0.25 ml of (1N) Folin-Ciocalteu reagent was added. After three minutes, one ml of 20 per cent sodium carbonate was added and mixed thoroughly. Then the tubes were placed in boiling water for one minute and cooled. The absorbance was measured at 725 nm against a reagent blank. The phenol activity was expressed as  $\mu\text{g}$  of catechol  $\text{g}^{-1}$  of leaf tissue (Zieslin and Ben Zaken, 1993).

#### Assay of $\beta$ -1, 3- glucanase

$\beta$ -1, 3-glucanase activity was assayed by the laminarin dinitrosalicylic acid method (Pan *et al.*, 1991b). The reaction mixture was consisted of 62.5 $\mu\text{l}$  of four per cent laminarin and 62.5  $\mu\text{l}$  of enzyme extract. The reaction was carried out at 40°C for 10 minutes. The reaction was then stopped by adding 375  $\mu\text{l}$  of dinitrosalicylic acid and heated for five

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minutes on boiling water, vortexed and its absorbance was measured at 500 nm. The enzyme activity was expressed as  $\mu\text{g}$  glucose released  $\text{min}^{-1} \text{mg}^{-1}$  of protein.

## RESULTS AND DISCUSSION

Changes in phenylalanine ammonia lyase (PAL) activity in crossandra plants :

In crossandra plants after treating with different combinations of bio agents, organic amendments, nanoparticles and chemicals the activity of PAL was induced in all the treated plants which were challenge inoculated with the pathogens and nematode. The enzyme activity reached the maximum at five days after last application and maintained at higher level up to seven days after application and slowly declined thereafter in all the treatments. Whereas in healthy and pathogen and nematode inoculated control plants the activity was less than that in all the other treatments. The treatment T<sub>16</sub> was found to record higher activity of PAL (0.895  $\mu\text{mol}$  of transcinnamic acid / min /g of leaf tissue) (Table 1).

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Phenyl propanoid metabolism starts with the conversion of L-phenylalanine into trans-cinnamic acid by the enzyme phenylalanine ammonia-lyase (PAL) and supplies the precursors for flavanoid pigments, lignins and phytoalexins (Hahlbrock and Scheel, 1989; Massala *et al.*, 1980). Increase in PAL activity subsequently increased the phenolic contents and leads to disease resistance. In the present study, treatment with integrated module T<sub>16</sub> induced the plants to synthesize PAL when they were challenge inoculated with *F.incarnatum* and *P. delattrei*. When turmeric plants were sprayed with *P. fluorescens* isolates, PAL activity increased (Kavitha, 2004, Meena *et al.*, 1999). Phenylalanine ammonia lyase played an important role in the biosynthesis of phenolic phytoalexins (Daayf *et al.*, 1997). The product of PAL activity was transcinnamic acid which was an immediate precursor for the biosynthesis of salicylic acid, a signal molecule in systemic acquired resistance (SAR) (Klessig and Malamy, 1994). Cell wall strengthening, through the deposition of lignin, proceeded by the induction of the synthesizing enzymes were played an important role in the defense response of *Lycopersicon esculentum* in reaction to a variety of elicitors (Mandal, 2009).

### Changes in phenols and $\beta$ 1, 3 Glucanase activities in crossandra plants :

The accumulation of phenol was found to increase from the third day after treatment and attained a peak on fifth days after last application and thereafter slowly declined. The treatment T<sub>16</sub> recorded maximum total phenol content (668.02  $\mu$ g of catchol/ g leaf tissue) at 5 days after last application whereas no significant change in the phenol content was observed in the uninoculated control (Table 2).

$\beta$  1, 3 glucanase activity was at a lower level in the uninoculated control. The treatment T<sub>16</sub> was found to record high enzyme activity (57.43  $\mu$ g of glucose / min /g of leaf tissue) at 5 days after last application and it was followed by T<sub>2</sub> which recording (56.43  $\mu$ g of glucose / min /g of leaf tissue) at 5 days after last application. In the inoculated control the enzyme activity was found to increase slightly (Table 3).

Many phenols and their oxidative products such as quinones were highly toxic to the invading pathogens (Sequeira, 1991) and enhanced the mechanical strength of host cell wall. The results of the present study demonstrated that higher levels of phenol accumulation were observed in the T<sub>16</sub> treated crossandra plants. Similar findings were reported in rice against *R. solani* (Meena *et al.*, 1999, Radja Commare, 2004), sugarcane against *Colletotrichum falcatum* (Viswanathan and Samiyappan, 1999), groundnut against *Cercospora personata* (Meena *et al.*, 2000), and turmeric against *Pythium aphanidermatum* (Kavitha, 2004). Raj kumar (2006) reported that the increase in phenol content in banana plants upto five days when treated with combined biocontrol agent application of BBs<sub>1</sub>+BBs<sub>2</sub>+BPf<sub>1</sub>.

Different types of pathogens and abiotic stress have been found to induce pathogen related (PR) and host-coded proteins. Synthesis and accumulation of PR proteins play an important role in plant defense system (Van Loon, 1997). The enzyme act upon the fungal cell wall, resulting in degradation and loss of inner content of cells (Maurhofer *et al.*, 1994).

Maurhofer *et al.* (1994) also reported the induction of  $\beta$ -1, 3-glucanase and induction of systemic resistance by *P. fluorescens* strain CHAO in tobacco. Bargabus *et al.* (2002) reported that, *B. pumilus* isolates 203-6 and 203-7 induced  $\beta$ -1, 3-glucanase in sugar beet. The present investigation confirms the earlier work of Xue *et al.* (1998) who reported that the inoculation of root rot pathogen *Rhizoctonia* spp. or anthracnose pathogen *Colletotrichum*

*lindimutheanum* induced systemic resistance by induction of peroxidases,  $\beta$ , 1-3-glucanases in the bean plants.

## CONCLUSION

It was found that soil application (SA) of *T.viride* @ 2.5 kg/ha at 20 DAP plus soil drenching (SD) of carbendazim @ 0.1% at 30 DAP plus SA of *T.viride* @ 2.5 kg/ha at 50 DAP plus Foliar application (FA ) of *P. fluorescens*@ 1.0 kg/ha at 70 DAP plus Foliar application (FA ) of *B. subtilis* @ 1.0 kg /ha at 90 DAP was found to be higher in induction of phenylalanine ammonia lyase (PAL), phenols and  $\beta$  1, 3 Glucanase in crossandra plants so that offering resistance in later stage in preventing disease incidence.

**Comment [DS29]:** Needs 1- recommendations  
2- Acknowledgments.

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**Table. 1** Changes of phenylalanine ammonia lyase (PAL) activity in crossandra plants treated with integration of different combinations of bio agents, organic amendments, nanoparticles and chemicals on the management of wilt in pot culture experiment

T. No.	Name of the treatment	$\mu$ mol of transcinamic acid / min /g of leaf tissue*					%Disease incidence
		Days after last application					
		0	3	5	7	9	
T <sub>1</sub>	SA of <i>P.fluorescens</i> (Pf-18) @ 2.5kg/ha at 20 DAP + Module A	0.472	0.684	0.878	710	685	58.3(49.79)*
T <sub>2</sub>	SA of <i>T.viride</i> (Tv-9) @ 2.5kg/ha at 20 DAP + Module A	0.485	0.702	0.884	0.750	0.695	50.0(44.98)
T <sub>3</sub>	SA of <i>B.subtilis</i> (Bs-10) @2.5kg/ha at 20 DAP + Module A	0.479	0.682	0.870	0.706	0.680	61.1(51.41)
T <sub>4</sub>	SA of Neem cake @ 250 kg/ha at 20 DAP + Module A	0.486	0.658	0.846	0.691	0.652	63.9(53.06)
T <sub>5</sub>	SD of carbendazim @ 0.1% at 20 DAP + Module A	0.478	0.690	0.878	0.734	0.682	52.8(46.58)
T <sub>6</sub>	SA of Phorate 10G @10 kg/ha at 20DAP + Module A	0.483	0.694	0.880	0.742	0.688	63.9(53.06)
T <sub>7</sub>	FA of nano particles @ 800 ppm at 20 DAP + Module A	0.492	0.692	0.884	0.745	0.680	55.6(48.18)
T <sub>8</sub>	SA of <i>P.f</i> (Pf-18) @ 2.5 kg/ha at 20 DAP + Module B	0.472	0.680	0.874	0.706	0.674	36.1(36.92)
T <sub>9</sub>	SA of <i>T.v</i> (Tv-9) @ 2.5 kg/ha at 20 DAP + Module B	0.479	0.695	0.875	0.700	0.678	30.6((33.54)
T <sub>10</sub>	SA of <i>B.s</i> (Bs-10) @ 2.5 kg/ha at 20 DAP + Module B	0.473	0.682	0.864	0.698	0.673	36.1(36.92)
T <sub>11</sub>	SA of Neem cake @ 250 kg/ha at 20 DAP + Module B	0.474	0.655	0.840	0.685	0.642	44.4(41.79)
T <sub>12</sub>	SD of carbendazim @ 0.1% at 20 DAP + Module B	0.471	0.688	0.874	0.730	0.680	33.3(35.24)
T <sub>13</sub>	SA of Phorate 10G @ 10kg/ha at 20DAP + Module B	0.472	0.692	0.870	0.740	0.675	41.7(40.18)
T <sub>14</sub>	FA of nano particles @ 800 ppm at 20 DAP + Module B	0.479	0.690	0.881	0.743	0.671	36.1(36.92)
T <sub>15</sub>	SA of <i>P.f</i> (Pf-18) @ 2.5 kg/ha at 20 DAP + Module C	0.468	0.688	0.883	0.712	0.689	27.8(31.79)
T <sub>16</sub>	SA of <i>T.v</i> (Tv-9) @ 2.5 kg/ha at 20 DAP + Module C	0.470	0.712	0.895	0.790	0.712	16.7(24.08)
T <sub>17</sub>	SA of <i>B.s</i> (Bs-10) @ 2.5 kg/ha at 20 DAP + Module C	0.472	0.685	0.875	0.693	0.678	19.4(26.15)
T <sub>18</sub>	SA of Neem cake @ 250 kg/ha at 20 DAP + Module C	0.481	0.660	0.852	0.695	0.654	22.2(28.11)
T <sub>19</sub>	SD of carbendazim @ 0.1% at 20 DAP + Module C	0.473	0.692	0.885	0.742	0.690	16.7(24.08)
T <sub>20</sub>	SA of Phorate 10G @ 10kg/ha at 20DAP + Module C.	0.468	0.702	0.888	0.754	0.695	19.4(26.15)
T <sub>21</sub>	FA of nanoparticles @ 800 ppm at 20 DAP + Module C	0.475	0.694	0.882	0.748	0.687	22.2(28.11)
T <sub>22</sub>	Inoculation of pathogen and nematode only	0.480	0.524	0.533	0.520	0.521	75.0(60.04)
T <sub>23</sub>	Un inoculated control	0.484	0.498	0.499	0.482	0.480	2.73(0.627)

\* Mean of three replications

DAP – Days after planting

CD (P=0.05)

Treatments 0.03

Days 0.01

Treatments × Days 0.06

Module A.= Foliar application (FA) of nano particles @ 800ppm at 30 DAP + SA of *T. viride* (Tv-9) @ 2.5 kg/ha at 50 DAP + FA of *P.f* (Pf-18)@ 1.0 kg/ha at 70 DAP + FA of *B.s* (Bs-10) @ 1.0 kg/ha at 90 DAP

Module B = FA of carbendazim @ 0.1% at 30 DAP + SA of *T.v* (Tv-9) @ 2.5 kg/ha at 50 DAP + FA of *P.f* (Pf-18) @ 1.0 kg/ha at 70 DAP + FA of *B.s* @ 1.0 kg/ha at 90 DAP

Module C= SD of carbendazim @ 0.1% at 30 DAP + SA of *T.v* (Tv-9) @ 2.5 kg/ha at 50 DAP + FA of *P.f* (Pf-18)@ 1.0 kg/ha at 70 DAP + FA of *B.s* (Bs-10) @ 1.0 /ha at 90 DAP

**Table 2. Changes of phenol activity in crossandra plants treated with integration of different combinations of bio agents, organic amendments, nanoparticles and chemicals on the management of wilt in pot culture experiment**

T. No.	Name of the treatment	µg of catachol /g of leaf tissue*					%Disease incidence
		Days after last application					
		0	3	5	7	9	
T <sub>1</sub>	SA of <i>P.fluorescens</i> (Pf-18) @ 2.5 kg/ha at 20 DAP + Module A	524.45	595.54	611.12	605.44	600.45	58.3(49.79)**
T <sub>2</sub>	SA of <i>T.viride</i> (Tv-9) @ 2.5 kg/ha at 20 DAP + Module A	521.31	630.82	664.97	662.87	657.46	50.0(44.98)
T <sub>3</sub>	SA of <i>B.subtilis</i> (Bs-10) @ 2.5kg/ha at 20 DAP + Module A	532.03	593.50	608.06	601.77	598.42	61.1(51.41)
T <sub>4</sub>	SA of Neem cake @ 250 kg/ha at 20 DAP + Module A	524.45	572.68	594.40	590.55	586.3	63.9(53.06)
T <sub>5</sub>	SD of carbendazim @ 0.1% at 20 DAP + Module A	525.32	610.20	648.52	608.33	605.46	52.8(46.58)
T <sub>6</sub>	SA of Phorate 10G @10kg/ha at 20DAP +Module A	522.78	620.45	655.60	650.30	645.22	63.9(53.06)
T <sub>7</sub>	FA of nano particles @800 ppm at 20 DAP + Module A	525.21	612.80	650.77	645.22	641.58	55.6(48.18)
T <sub>8</sub>	SA of <i>P.f</i> (Pf-18) @ 2.5 kg/ha at 20 DAP + Module B	525.73	590.23	605.32	601.88	595.64	36.1(36.92)
T <sub>9</sub>	SA of <i>T.v</i> (Tv-9) @ 2.5 kg/ha at 20 DAP + Module B	520.95	592.34	660.35	658.75	598.54	30.6((33.54)
T <sub>10</sub>	SA of <i>B.s</i> (Bs-10) @ 2.5 kg/ha at 20 DAP + Module B	521.31	592.34	604.96	600.22	596.24	36.1(36.92)
T <sub>11</sub>	SA of Neem cake @ 250 kg/ha at 20 DAP + Module B	522.54	570.86	590.06	588.43	584.08	44.4(41.79)
T <sub>12</sub>	SD of carbendazim @ 0.1% at 20 DAP + Module B	524.28	608.70	644.32	602.66	602.46	33.3(35.24)
T <sub>13</sub>	SA of Phorate 10G @10kg/ha at 20DAP +Module B	525.31	615.34	652.85	648.55	643.44	41.7(40.18)
T <sub>14</sub>	FA of nano particles @800 ppm at 20 DAP + Module B	523.53	610.83	645.87	640.32	634.52	36.1(36.92)
T <sub>15</sub>	SA of <i>P.f</i> (Pf-18) @ 2.5 kg/ha at 20 DAP + Module C	524.93	530.55	598.45	620.18	610.74	27.8(31.79)
T <sub>16</sub>	SA of <i>T.v</i> (Tv-9) @ 2.5 kg/ha at 20 DAP + Module C	522.01	635.94	668.02	660.26	6658.94	16.7(24.08)
T <sub>17</sub>	SA of <i>B.s</i> (Bs-10) @ 2.5 kg/ha at 20 DAP + Module C	524.45	598.42	612.85	611.75	605.24	19.4(26.15)
T <sub>18</sub>	SA of Neem cake @ 250 kg/ha at 20 DAP + Module C	525.32	576.74	598.44	596.53	588.6	22.2(28.11)
T <sub>19</sub>	SD of carbendazim @ 0.1% at 20 DAP + Module C	522.78	612.40	652.25	613.37	608.64	16.7(24.08)
T <sub>20</sub>	SA of Phorate 10G @ 10kg/ha at 20DAP +Module C.	525.21	621.38	658.60	653.66	647.33	19.4(26.15)
T <sub>21</sub>	FA of nanoparticles @800 ppm at 20 DAP + Module C	525.73	615.08	653.88	649.22	645.83	22.2(28.11)
T <sub>22</sub>	Inoculation of pathogen and nematode only	520.95	524.44	546.81	533.48	526.45	75.0(60.04)
T <sub>23</sub>	Un inoculated control	521.31	521.15	526.84	521.38	519.10	2.73(0.627)

\* Mean of three replications

DAP – Days after planting

CD (P=0.05)

Treatments 0.03

Days 0.01

Treatments × Days 0.06

**Table 3. Changes of  $\beta$  1, 3 Glucanase activity in crossandra plants treated with integration of different combinations of bio agents, organic amendments, nanoparticles and chemicals on the management of wilt in pot culture experiment**

T. No.	Name of the treatment	µg of glucose / min /g of leaf tissue*					%Disease incidence
		Days after last application					
		0	3	5	7	9	
T <sub>1</sub>	SA of <i>P.fluorescens</i> (Pf-18) @ 2.5 kg/ha at 20 DAP + Module A	38.09	43.12	53.37	52.74	50.81	58.3(49.79)
T <sub>2</sub>	SA of <i>T.viride</i> (Tv-9) @ 2.5kg/ha at 20 DAP + Module A	37.23	45.67	56.43	54.34	52.72	50.0(44.98)
T <sub>3</sub>	SA of <i>B.subtilis</i> (Bs-10) @ 2.5kg/ha at 20 DAP + Module A	36.67	42.21	52.48	51.87	50.34	61.1(51.41)
T <sub>4</sub>	SA of Neem cake @ 250 kg/ha at 20 DAP + Module A	37.36	41.96	51.12	50.28	49.13	63.9(53.06)
T <sub>5</sub>	SD of carbendazim @ 0.1% at 20 DAP + Module A	36.53	44.67	55.55	53.43	51.92	52.8(46.58)
T <sub>6</sub>	SA of Phorate 10G @ 10kg/ha at 20DAP + Module A	37.02	45.09	53.45	53.23	52.34	63.9(53.06)
T <sub>7</sub>	FA of nano particles @ 800 ppm at 20 DAP + Module A	38.17	43.59	53.42	52.85	50.93	55.6(48.18)
T <sub>8</sub>	SA of <i>P.f</i> (Pf-18) @ 2.5 kg/ha at 20 DAP + Module B	38.12	42.21	52.77	51.75	50.12	36.1(36.92)
T <sub>9</sub>	SA of <i>T.v</i> (Tv-9) @ 2.5 kg/ha at 20 DAP + Module B	38.02	44.63	55.60	53.21	51.60	30.6((33.54)
T <sub>10</sub>	SA of <i>B.s</i> (Bs-10) @ 2.5 kg/ha at 20 DAP + Module B	37.94	41.35	51.80	50.94	50.15	36.1(36.92)
T <sub>11</sub>	SA of Neem cake @ 250 kg/ha at 20 DAP + Module B	37.13	41.24	50.74	49.88	48.05	44.4(41.79)
T <sub>12</sub>	SD of carbendazim @ 0.1% at 20 DAP + Module B	36.98	43.61	52.88	52.21	51.30	33.3(35.24)
T <sub>13</sub>	SA of Phorate 10G @ 10kg/ha at 20DAP + Module B	37.64	44.54	53.08	53.62	51.96	41.7(40.18)
T <sub>14</sub>	FA of nano particles @ 800 ppm at 20 DAP + Module B	37.26	43.14	51.67	51.98	50.23	36.1(36.92)
T <sub>15</sub>	SA of <i>P.f</i> (Pf-18) @ 2.5 kg/ha at 20 DAP + Module C	37.36	44.45	54.82	53.74	51.93	27.8(31.79)
T <sub>16</sub>	SA of <i>T.v</i> (Tv-9)@ 2.5 kg/ha at 20 DAP + Module C	36.53	46.60	57.43	55.38	52.72	16.7(24.08)
T <sub>17</sub>	SA of <i>B.s</i> (Bs-10) @ 2.5 kg/ha at 20 DAP + Module C	37.02	43.46	53.84	52.87	51.70	19.4(26.15)
T <sub>18</sub>	SA of Neem cake @ 250 kg/ha at 20 DAP + Module C	37.94	42.64	52.14	51.30	50.13	22.2(28.11)
T <sub>19</sub>	SD of carbendazim @ 0.1% at 20 DAP + Module C	38.17	45.54	52.14	51.33	52.90	16.7(24.08)
T <sub>20</sub>	SA of Phorate 10G @ 10kg/ha at 20DAP + Module C.	36.67	46.23	53.84	52.41	51.44	19.4(26.15)

T <sub>21</sub>	FA of nanoparticles @ 800 ppm at 20 DAP + Module C	37.36	44.55	53.41	52.76	49.91	22.2(28.11)
T <sub>22</sub>	Inoculation of pathogen and nematode only	36.53	39.36	44.96	43.92	41.72	75.0(60.04)
T <sub>23</sub>	Un inoculated control	37.02	38.9	38.72	38.53	38.21	<b>2.73(0.627)</b>

\* Mean of three replications

DAP – Days after planting

**CD (P=0.05)**

**Treatments            0.03**

**Days                    0.01**

**Treatments × Days   0.06**