Original Research Article

In vitro efficacy of compatible microbial consortia on seed germination and seedling vigour in tomato (Solanum lycopersicum L.)

ABSTRACT

In vitro study was conducted to check out the efficacy of indigenous compatible microbial consortia [(T-1; P. fluorescens P-7 + P. fluorescens P-12 + Trichoderma sp. T-5 + Trichoderma sp T-11), (T-2; P. fluorescens P-12 + Trichoderma sp. T-5 + Trichoderma sp. T-11) and (T-3; P. fluorescens P-7 + Trichoderma sp. T-5 + Trichoderma sp. T-11)] on plant growth promoting activities like seed germination, seedling vigour index, shoot length, root length, dry and fresh weight of shoot, dry and fresh weight of root by standard filter paper method. The result shows that, T-3 significantly increased vigour index of tomato seedlings (93.24 %), including germination per cent (20.84 %), shoot length (61%) and root length (59.36 %) over control at 10 DAS. On the basis of our findings , all the tested microbial consortia had significantly promoted the plant growth over control where T-3 was the best among the tested microbial consortia exhibiting plant growth promoting potential for commercial exploitation.

Keywords: Trichoderma, Pseudomonas fluorescens, indigenous, commercial

1. INTRODUCTION

Tomato is a member of the Solanaceae family, representing one of the most valuable plant families for vegetable and fruit crops. Tomatoes contain many health-promoting compounds and are easily integrated as a nutritious part of a balanced diet [1]. In addition to consuming the fresh fruits, consumers use tomatoes in processed products such as soups, juices, and sauces [2,3]. The yield of tomato is restricted to a great extent due to various diseases and insect pests associated with tomato cultivation [4]. There is a great need to produce quality and healthy seedlings, which are capable of withstanding adverse abiotic and biotic stresses after transplanting with improved mineral nutrient uptake [5].

Although the use of chemicals to reduce or prevent losses caused by this agent seems simple and successful, the damage inflicted by the residual effects of chemicals on humans and environment should certainly be taken into account. Also unplanned and wide use of fungicides often leads to serious environmental problems besides affecting the health of users and consumers. Moreover, innovative and safe methods like use of biocontrol agents need to be identified and evaluated for continuous search to develop ecofriendly strategies to reduce the dependence on harmful chemicals [6].

Comment [D1]: Write full name first time
Comment [D2]: .

Comment [D3]: Delete space
Comment [D4]: Delete space
Comment [D5]: Delete space
Comment [D6]: 61.00%
Comment [D7]: Delete space
Comment [D8]: Write full name first time
Comment [D9]: Experimental
Comment [D10]: of the present study

Comment [D11]: , tomato

Comment [D12]: ,

Comment [D13]: Specify name

Comment [D14]: Bio-control

Comment [D15]: Eco-friendly

A pre-sowing inoculation of planting material as well as the planting medium with the consortia of beneficial microorganisms is an innovative approach for production of quality and healthy seedlings in horticultural production. A microbial consortium is a carrier based product containing nitrogen fixing, phosphorus and potassium solubilising and plant growth promoting microorganisms in a single formulation. The synergistic effect of the formulated microbes can help in providing healthy and vigorous seedlings and considerably reducing the cost of cultivation by reducing fertilizer requirement of vegetables [7].

Plant-growth-promoting microorganism (PGPM) is a term that applies to all microorganisms (e.g., bacteria, actinomycetes, fungi, and algae) that have a beneficial effect on plant growth through the action of either direct or indirect mechanisms (e.g., mineral nutrition, ethylene reduction, disease suppression) [8]. It has been suggested that, due to the plethora of interactions that can occur when single species are inoculated in the field, positive and consistent results in terms of facilitating plant growth are not always achieved [9]. Most studies of PGPMs are based on interactions of single microorganisms with plants, evaluating different parameters of growth and plant health, such as length or weight of the plant or its individual tissues, chlorophyll content, or the nutritional content of its tissues or fruits [8,10,11,12]. However, more consistent positive results may be obtained by inoculating plants with microbial consortia containing two or more beneficial microorganisms [13,14].

2. MATERIAL AND METHODS

2.1 Preparation of liquid compatible microbial consortia (CMC)

A 250 ml suspension of each selected native isolates of *Trichoderma* sp. (T-5 and T-11) were prepared from 9 days old cultured PDA medium plates. The plates were rinsed with sterile distilled water and the mycelia were carefully scraped off the agar with a bent glass rod. This suspension was filtered through filter paper (Whatman No. 1) to separate the spores from the mycelia. The concentration was adjusted to 3.7×10^8 spores/ml with the help of haemocytometer [15]. A 250 ml of each selected native isolates of *Pseudomonas fluorescens* (P-7 and P-12) cell suspension was prepared by inoculating the strain into King's B broth followed by shaking for 48 hrs (150 rpm) at room temperature. The bacterial suspension was roughly adjusted optically at 1 x 10^9 cfu/ml (O.D. 600=1) [16].

Liquid consortia were prepared by mixing equal volume of each selected isolate just before use [17].

2.2 In vitro efficacy of liquid CMC on tomato seedlings

The healthy seeds of tomato cv. Pusa Ruby were selected for experimental purpose. Tomato seeds were surface sterilized with 1.0 % sodium hypochlorite for 2 min for all treatments followed by three rinsed with sterile distilled water. The *in vitro* experiment was conducted in a complete randomised design (CRD) and five replications were maintained for each treatment. The total five numbers of treatments *viz.*, T1 (*P. fluorescens* P-7 + *Trichoderma* sp. T-5 + *Trichoderma* sp. T-11),

Comment [D16]: materials

Comment [D17]: crop

Comment [D18]: system

Comment [D19]: crops

Comment [D20]: italicized

Comment [D21]: Merge this sentence with immediate above paragraph after reference 16.

Comment [D22]: solution

Comment [D23]: minute

Comment [D24]: times

T2 (*P. fluorescens* P-12 + *Trichoderma* sp. T-5 + *Trichoderma* sp. T-11), T3 (*P. fluorescens* P-7 + *P. fluorescens* P-12 + *Trichoderma* sp. T-5 + *Trichoderma* sp. T-11), T4- chemical control and T5-control were used. This experiment was carried out by standard filter paper method (three layered moistened filter papers in Petri plates, 10 seeds/plate and 20 seeds/replication) [18].

2.3 Wet seed treatment

The surface sterilized seeds were soaked with liquid formulations of consortia [@ 1.0 % or 10 µl/ 1 g seeds; 10 µl formulation of CMC added in 990 µl of sterile distilled water/1 g seed (400 tomato seeds)] and shade dried in laminar air flow for 5 hours. For chemical control treatment The surface sterilized seeds were treated with captan 50 % WP (seed dressing @ 0.3 % or 3mg/1g seed). For control treatment, surface sterilized seeds were soaked in sterile distilled water (@ 1ml/1g seed) and shade dried in laminar air flow for 5 hours [17].

Per cent germination at 10 DAS: Per cent germination was calculated using the following formula – Per cent germination = (No. of seeds germinated / Total no. of seeds sown) X 100

Seedlings shoot length and root length (cm): The root length and shoot length of individual seedlings (10 seedlings/ replication) were measured. The shoot length was measured from collar region to the tip of the seedling with the help of a scale and the mean shoot length was expressed in cm. The root length measured from collar region to the tip of primary root with the help of a scale and the mean root length was expressed in cm.

Seedling vigour index (SVI): The vigour index of seedlings was calculated by adopting the method suggested by Abdul Baki and Anderson (1973) [19] and expressed in number by using the below formula.

SVI = Germination (%) x [Mean shoot length (cm) + Mean root length (cm)].

Fresh weight (mg) of seedling shoot and root: The fresh weight (mg) of root and shoot of individual seedlings (10 seedlings /replication) were measured.

Dry weight (mg) of seedling shoot and root: The dry weight of root and shoot of individual seedlings (10 seedlings/ replication) were measured after oven drying at 60° C (when constant weight obtained) for 24 hours.

3. RESULTS AND DISCUSSION

3.1 In vitro efficacy of liquid CMC on tomato seedlings

Standard filter paper method was carried out *in vitro* to check the efficacy of liquid consortia on tomato seed germination and seedling vigour index. Per cent germination was recorded at 10 DAS. Tomato seed germination per cent was significantly highest in T-3 (87.00%) followed by T-2 (85.00%) and T-1 (82.00%). The lowest seed germination per cent was observed in control treatment (72.00%) (Table 1). These results revealed that the T-3 significantly increased seed

Comment [D25]: italicized

Comment [D26]: italicized

Comment [D27]: reduce space

Comment [D28]: italicized

Comment [D29]: italicized

Comment [D30]: p

Comment [D31]: , the

Comment [D32]: Delete space

Comment [D33]: 3 mg/1 g

Comment [D34]: 1 ml/1 g

Comment [D35]: Delete space

Comment [D36]: -

Comment [D37]: 60 °C

Comment [D38]: Reduce space
Comment [D49]: Reduce space
Comment [D40]: Delete space
Comment [D41]: Delete space
Comment [D42]: Delete space
Comment [D43]: Delete space

germination per cent (20.84 %) over control treatment (Table 1). Murthy et al. (2013) reported that the application of liquid consortia of *Trichoderma* spp., significantly increased the tomato seed germination per cent at 10 DAS.

3.2 Seedlings shoot length and root length (cm) at 10 DAS

Among the tested treatments, significantly maximum shoot length and root length was observed in seed treated with T-3 (4.83 cm), (5.13 cm) respectively (Table 1) than the other treatment. Minimum shoot length and root length was observed in control treatment (3.00 cm) and (3.22 cm) respectively (Table 1). The T-3 significantly increased shoot length (61.00 %) and root length (59.36 %) over control treatment (Table 2). Murthy et al. (2013). reported significant increased the shoot length and root length of tomato seedling at 10 DAS with the application of liquid consortia of *Trichoderma* spp. [14].

3.3 Seedling vigour index (SVI) at 10 DAS

Among the tested treatments, significantly maximum seedling vigour index was recorded in seed treated with T-3 (865.67) than the other treatment . Minimum seedling vigour index was observed in control treatment (447.84) (Table 1). These results revealed that the T-3 significantly increased vigour index of tomato seedlings (93.29%) over control treatment (Table 2). The present findings are in harmony with the findings of Manikandan et al. (2010) where liquid formulation of *P. fluorescens* Pf1 significantly promoted tomato plant growth compared to untreated control [45].

3.4 Fresh weight (mg) of seedling shoot and root at 10 DAS

Significantly maximum fresh weight of shoot and root was recorded in T-3 (10.79 mg), (0.33 mg) and minimum in control (5.11 mg) and (0.16 mg) respectively (Table 1). These results revealed that the T-3 significantly increased shoot fresh weight (111.16 %) and root fresh weight (106.25 %) over control treatment (Table 2). The present findings are in agreement with Murthy et al. (2013)[14] where liquid consortia of *Trichoderma* spp., significantly increased the fresh weight of shoot at 10 DAS.

3.5 Dry weight (mg) of seedling shoot and root at 10 DAS

Among the tested treatments, significantly maximum dry weight of shoot and root was recorded in T-3 (0.67 mg), (0.40 mg) and minimum in control (0.04mg) and (0.018 mg) respectively (Table 1). The T-3 significantly increased shoot dry weight (67.50 %) and root dry weight (122.23 %) over control treatment (Table 2). Murthy et al. (2013) [14] reported that the application of liquid consortia of *Trichoderma* spp. significantly increased the dry weight of shoot at 10 DAS.

Table.1 *In vitro* efficacy CMC on tomato seed germination (%), seedling shoot length, root length, shoot fresh and dry weight, root fresh and dry weight and vigour index at 10 DAS

Comment [D44]: Delete space Comment [D45]: Delete space Comment [D46]: Follow journal pattern Comment [D47]: Italicized Comment [D48]:, Comment [D49]: Comment [D50]: Treatment Comment [D51]: Delete space Comment [D52]: Delete space Comment [D53]: Follow journal style Comment [D54]: Delete comma and reduce Comment [D55]: delete Comment [D56]: When Murthy et al reported this, how you can cite reference of others, justify? Comment [D57]: s Comment [D58]: [15] Comment [D59]: reduce space Comment [D60]: s Comment [D61]: delete space Comment [D62]: delete space Comment [D63]: [20] Comment [D64]: italicized Comment [D65]: 0.04 mg

Comment [D66]: ,
Comment [D67]: delete space
Comment [D68]: delete space
Comment [D69]: [20]
Comment [D70]: italicized

Comment [D71]: of

Treatment	Seed germination	Seedling shoot at 10 DAS	Seedling root at 10 DAS	Seedling
ı	%			vigour index at

		Shoot	Fresh wt.	Dry wt.	Root	Fresh wt.	Dry wt.	10 DAS	Comment [
		length (cm)	(mg)	(mg)	length (cm)	(mg)	(mg)		Comment [
T-1	82.00 (65.12)	4.09	8.59	0.57	4.90	0.28	0.03	738.74	Comment [
T-2	85.00(67.51)	4.45	9.14	0.59	4.97	0.30	0.03	800.34	Comment [
T-3	87.00(68.95)	4.83	10.79	0.67	5.13	0.33	0.04	865.67	
T-4	75.00(60.41)	3.26	5.45	0.42	3.58	0.18	0.02	513.14	
T-5	72.00 (58.14)	3.00	5.11	0.40	3.22	0.16	0.01	447.84	
Sem+-	2.81(2.07)	0.05	0.31	0.02	0.07	0.02	1.32	23.16	
C.V. (%)	7.83(7.22)	3.23	9.04	11.06	3.68	19.10	31.69	7.67	
CD(p=0.01)	11.30 (8.31)	0.23	1.27	0.10	0.28	0.09	0.01	93.16	
CD(P=0.05)	8.29(6.09)	0.16	0.93	0.57	0.21	0.069	0.01	68.31	

^{*}Values in parenthesis are angular transformed values.

Table.2 In vitro efficacy of CMC on per cent increase of tomato seed germination (%), shoot length, root length, shoot fresh and dry weight, root fresh and dry weight and seedling vigour index at 10 DAS

Treatme	Seed	Per cent increase of plant growth promotion over control					Seedlin	
nt	ger. At 10	Seedling shoot			Seedling shoot Seedling root			g vigour
	DAS	Length	Fresh wt.	Dry wt.	Length	Fresh wt.	Dry wt.	index
T1	13.89	36.34	68.10	42.50	52.17	75.00	66.67	64.95
T2	18.06	48.34	78.86	47.50	54.34	87.50	66.67	<mark>78.72</mark>
T3	20.84	61.00	111.16	67.50	59.36	106.25	122.23	93.24
T4	04.17	08.67	06.65	05.00	11.18	12.50	11.12	14.58
T5	-					•	•	Comment [D76]: Properly align word and write full name of words

4. CONCLUSION

With the high rate of increasing in world population, our basic requirements of food should be made while enhancing sustainability in crop production. Biological means of plant disease management as well as plant growth promotion can be employed as an important alternative to enhance quality and quantity of crop production. The current investigation showed that all the tested native compatible microbial consortia had promoted the growth of tomato seedlings over control at different ranges. The highest seed germination (87%) and seedling vigour (865.67) were recorded in T-3 followed by T-2 and T-1 respectively. These results show their potential to promote seedlings growth as compared to chemical check and control. However, they can be further evaluated to study their potential in field. Moreover, measures to enhance shelf life, doses of applications and awareness among the farmers can be taken to enhance plant growth which in turn promote crop productivity.

Comment [D77]: World population is increasing at high rate, therefore,

Comment [D78]: To provide

Comment [D79]: Of growing population

Comment [D80]: 87.00%

Comment [D81]: ,

Comment [D82]: the

Comment [D83]: reduce space

[D72]: weight of shoot
[D73]: weight of shoot
[D74]: weight of root
[D75]: weight of root

REFERENCES

- Martí R, Roselló S, Cebolla-Cornejo J. 2016. Tomato as a source of carotenoids and polyphenols targeted to cancer prevention. Cancers (Basel). 2016;E58. doi: 10.3390/cancers8060058
- Krauss S, Schnitzler WH, Grassmann J, Woitke M. The influence of different electrical conductivity values in a simplified recirculating soilless system on inner and outer fruit quality characteristics of tomato. Journal of Agriculture Food and Chemistry. 2006;54:441–448.
- 3. Li Y, Wang H, Y. Zhang Y, Martin C. 2018. Can the world's favorite fruit, tomato, provide an effective biosynthetic chassis for high-value metabolites?. Plant Cell Reports. 2018;37:1443–1450. doi: 10.1007/s00299-018-2283-8
- 4. Panthee DR, Chen F. Genomics of fungal resistance in tomato. Current Genomics. 2010;11:30-39.
- Nzanza B, Diana M, Puffy, S. Tomato (Solanum lycopersicum L.) seedling growth and development as influenced by *Trichoderma harzianum* and arbuscular mycorrhizal fungi. African Journal of Microbiology Research. 2011; 5(4):425-431.
- Deepti S, Nidhi, D. Bioefficacy of fungicides and plant extract against Alternaria solani causing early blight of tomato. International Conference on Plant, Marine and Environmental Sciences. 2015.
- Jayashree C, Jagadesh, KS. 2017. Testing the effect of the microbial consortium on growth of vegetable seedlings in a farmer's nursery. International Journal of Current Microbiology and Applied Sciences. 2017; 6(2):1636-1639.
- Abhilash PC, Dubey RK, Tripathi V, Gupta VK, Singh, HB. 2016. Plant growth-promoting microorganisms for environmental sustainability. Trends in Biotechnology. 2016;34:847-850
- 9. Babalola OO. Beneficial bacteria of agricultural importance. Biotechnology Letters. 2010;32:1559-1570.
- Khan A, Singh J, Upadhayay VK, Singh AV, Shah S. Microbial biofortification: A green technology through plant growth promoting microorganisms. p. 255-269. In sustainable green technologies for environmental management. Springer: Singapore. 2019 ISBN 9789811327728.
- Ney LD, Franklin K, Mahmud, Cabrera M, Hancock D, Habteselassie M. Impact of inoculation with local effective microorganisms on soil nitrogen cycling and legume productivity using composted broiler litter. Applied Soil Ecology. 2020; 154:103567.
- 12. Mahmud K, Franklin D, Ney L, Cabrera M, Habteselassie M, Hancock, D. Improving inorganic nitrogen in soil and nutrient density of edamame bean in three consecutive summers by utilizing a locally sourced bio-inocula. Organic Agriculture. 2021;1-11.
- 13. Sharma S, Compant S, Ballhausen MB, Ruppel, S, Franken P. The interaction between *Rhizoglomus irregulare* and hyphae attached phosphate solubilizing bacteria increases

Comment [D84]:

Comment [D85]: favourite

Comment [D86]:

Comment [D87]: Reduce space

- plant biomass of *Solanum lycopersicum*. Microbiological Research. 2020;240:126556 doi: 10.1016/j.micres.2020.126556. Epub 2020 Jul 9.
- Zhang S, Merino N, Okamoto A, Gedalanga P. 2018. Interkingdom microbial consortia mechanisms to guide biotechnological applications. Applied Microbiology and Biotechnology. 2018;11:833-847.
- Dubos B. Fungal antagonism in aerial agrobiocenoses. p. 107-135. In. Innovative approaches to plant disease control. 1st ed. Chet, W. John, Sons, New York. 1987.
- Mulya K, Wataneabe M, Goto M, Takikawa Y, Tsuyumu S. Suppression of bacterial wilt disease of tomato by root dipping with *P. fluorescens*. Annual Phyto Pathological Society of Japan. 1996;62:134-140.
- 17. Srinivasan K, Mathivanan N. Biological control of sunflower necrosis virus disease with powder and liquid formulations of plant growth promoting microbial consortia under field conditions. Biological Control. 2009; 51:395-402.
- 18. International Seed Testing Association (ISTA). International rules for seed testing. Seed Science and Technology. 1993;21:1-288.
- Abdul-Baki A, Anderson, JD. Vigour determination in soybean seed by multiple criteria.
 Crop Science. 1973;13:630-633.
- Murthy NK, Devi NK, Srinivas C. Efficacy of *Trichoderma asperellum* against *Ralstonia solanacearum* under greenhouse conditions. Annals of Plant Sciences. 2013;2(9):342-350.
- 21. Manikandan R, Saravanakumar, D, Rajendran L, Raguchander T, Samiyappan R. Standardization of liquid formulation of *Pseudomonas fluorescens* Pf 1 for its efficacy against *Fusarium* wilt of tomato. Biological control. 2010;54:83-89.