Review Article

ANTIBIOGRAM PATTERN OF BACTERIA ISOLATES FROM GROUND WATER (BOREHOLE WATER) RESOURCES FROM ISOLATED MICROORGANISMS

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Abstract

The study considered antibiogramme of bacteria isolates from selected ground water (borehole water) resources susceptibility pattern of the isolated microorganisms in Port Harcourt during the dry and wet season of 2020. Seven (7) sampling locations, designated as Abuloma, Borikiri, Eagle Island, Fimie, Macoba-Isaka, Rumuekini and Rumuokoro were established. Water samples were collected from boreholes in 10 ml sterile sampler and placed in an ice-parked container and send to the laboratory for analysis (Vibrio, *Stapylococcus*, Total Heterotrophic bacteria, Salmonella, Shigella, Total coliform and Fecal coliforms). Microbial enumeration was undertaken and count presented in percentages. Bacteria isolates for susceptibility and resistance where performed using Ofloxacin, Augumentin, Cetazidine, Gentamycin, Cefiximine, Ciproflaxin, Cefturoxine and Nitrofurantoin. There was an observed increased rate of resistance antibiotics in this study which could be attributed to the widespread misuse of various antibiotics without control. Thus, it is recommended that groundwater source (boreholes) be sited away from septic tanks source(s) as possible interference under the influence of proximity could become inevitable.

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Keyword: Groundwater, Antibiogram, Susceptibility, Resistance, Borehole sources

Introduction

Water is generally known to occupy over 70% of the earth crust. It is a fundamental requirement for life and this becomes inevitable for humans (Gideon *et al.,* 2017). Water is by far the most abundant substance in animal and plant tissues as well as the world around us. It accounts for about 70% of the human body and it is higher in many growing plants (Odesina, 2003).Water is not only used all over the world in large quantities for drinking purposes, but it is used even in greater quantities for cooking, a solvent in industrial processes, bleaching, raising steam to drive engines or turbines to generate electricity as well as washing (Odesina, 2003). However, due to increases in human population, industrial and recreational activities, and agricultural activities among others the access to quality water consumption has become a teething problem to many mostly in developing countries. However, having access to good and potable water has been an age-long challenge super imposed by mostly anthropogenic activities and less of nature mostly in Sub-Sara Africa.

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Water bacteria from different sources (human, animal and environmental) are able to interact and resistance evolves as a consequence of promiscuous exchange and shuffling of genes, genetic platforms and genetic vectors (Baquero *et al.,* 2008). The most important felt needs in public health is water and the availability of safe water dictates the quality of life since water is a basic requirement About 1.2 billion people worldwide lack access to safe drinking water (Wilkes *et*

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al., 2009). The sub-Sahara Africa accounts for over 1/3 of the number, and is lagging behind in progress towards the MDG target, with only about 60%of the population using improved sources of drinking water (WHO, 2010).Water intended for human consumption must be free from organism and from concentration of chemical substances that may be a hazard to health. Infectious diseases posses' major public health challenge to man when water sources are mostly contaminated by E. coli (Feachem,2001).

In public and environmental health, antimicrobial resistance is a threat. It is a characteristic of pathogen causing different diseases but generally not a problem of disease pathology rather one of the limited therapy options. These antibiotic resistance bacteria have been found in a surprising diverse range of environments including clinics, animal pens, orchards, food, sewage as well as chlorinated and un chlorinated water supplies (Sammie *et al*., 2012; Chopra *et al*., 2001). Bacteria are common contaminants worldwide and the release of human and animal wastes in to the environment exacerbates bacterial contamination especially in aquatic setting. Increase resistance to antibiotics may pose great challenge for the effective treatment of bacterial infection. However, Biograms are used by clinicians to access local susceptibility rates, as an aid in selecting empiric antibiotic therapy. An antibiogram shows the aggregate number of bacteria tested against antimicrobial and incorporates the extent of bacterial isolates vulnerable to every antimicrobial operator tested (Fridkin *et al*., 2001). Plasmid analysis helps in the differentiating isolated microorganisms and is thus a useful in the epidemiologic investigation of enteric disease outbreak (Soumik *et al*., 2010). Whereas antibiotics are metabolites having preferential antimicrobial activity; hence they are widely used for curing of human ailments caused by microorganisms. These antibiotics compounds are used either in their natural form or as semi-synthetic derivatives. The latter are usually produced by isolating the antibiotic nucleus and subjecting it to chemical modification (Wright, 2007; Kemper, 2007).The overuse of antibiotic in human and animals for treatment leads to the release of antibiotics and antibiotic resistant strains into the environment as stated by (Silbergeld *et al*.,2008; Ghafur *et al*.,2010).Antibiotic resistance is a natural phenomenon and represents an evolutionary response to the strong selective pressure resulting from these compounds (Chaturvedi *et al*.,2008; Ogbonna &Azuonwu., 2019). In this scenario, an antibiotic may kill virtually all the bacteria causing a disease in a patient but a few bacteria that are genetically may survive. These go on to reproduce or to transfer their resistance to others of their species through the processes of gene exchang of casese.

According to (Prescott *et al*.,2001) the wide spread of water borne diseases as a result of contamination of water sources, concerns have been raised that the disease failed to be cured due to resistance to commonly prescribed antibiotics by the contaminating microorganisms originating from livestock excreta and human sewage. (Matthew *et al*.,2007) reported that for the past years antibiotic resistant bacteria species are very ubiquitous in the environment and their negative impact has greatly increased drastically. The improper use of antibiotic and lack of awareness are considered as the most important factor for the emergence, selection, and dissemination of antibiotic resistant bacteria species in the environment (Neu, 1992; Abera *et al*., 2013).

Several literatures have shown that inadequately treated sewage and wastes are mainly the sources of antibiotic resistant bacteria in the environment (Hanwood *et al*., 2001; Inversen *et al*., 2002). This is said to be largely due to the failure or neglecting regulatory agencies in controlling drug use and as such these drugs are usually obtained over the counter without the supervision of the prescription by a licensed medical officer of health (Sanders,2005).

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MATERIALS AND METHODS

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Study Area

The study areas are fast growing sub-urban in Port Harcourt with geographical coordinates of Latitude 4° 49' 27.0012" N and Longitude 7° 2' 0.9996" E. It has 9 meters above sea level with a tropical climate. It has a significant rainfall pattern in most months of the year and a short dry season with little effect (Okafor, 1973; Demographia, 2016; **www.wikipedia.com,** 2010). Temperature varies by 24C throughout the year.

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S/N	Sample Location	GPS	Sample Water Source	Sample Codes
1	Abuloma	4.7432°N 7.0821°E	Ground water (Borehole)	AB
2	Borikiri	4.7463°N 7.0364°E	Ground water (Borehole)	BO
3	Eagle Island	4.8667°N 6.9833°E	Ground water (Borehole)	EI
4	Fimie	4.7829°N 6.9773°E	Ground water (Borehole)	FI
5	Macoba-Isaka	4.8873°N 6.9415°E	Ground water (Borehole)	MI
6	Rumuekini	4.7878°N 7.0415°E	Ground water (Borehole)	RK
7	Rumuokoro	4.4424°N 6.5915°E	Ground water (Borehole)	RO

Sample Location

Table 1: Sample Collection Areas

Sample Collection

Samples were collected in seven (7) different boreholes with the aid of a sterilized 10mls plastic containers haven rinsed these containers with the sample. The containers were placed under the running tap and were Stoppard while the tap was still running. Samples were thus placed in an ice-packed cooler and were transported to the microbiology laboratory of Rivers State University for microbial analysis.

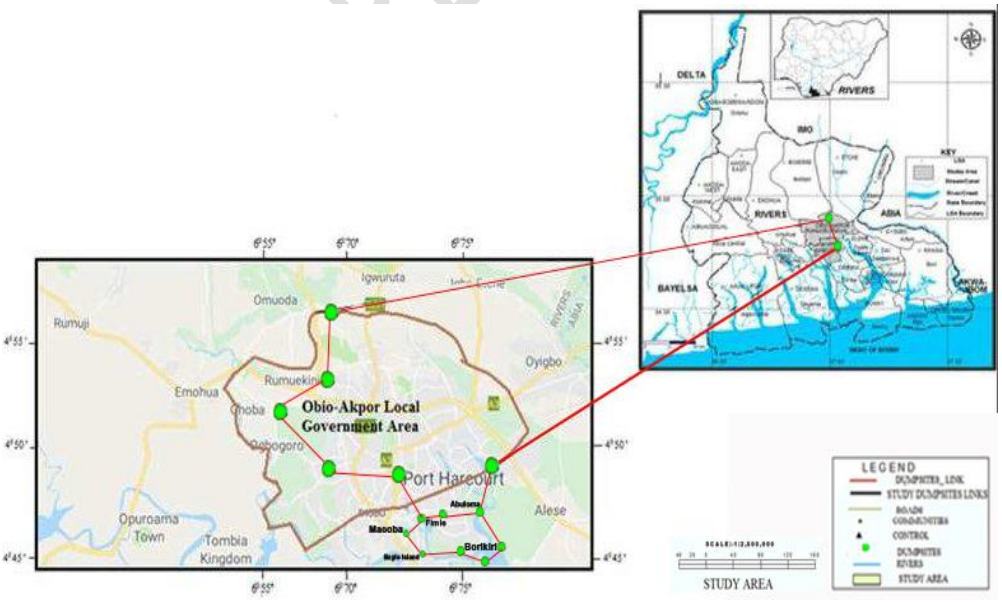

Fig 1. Map Rivers State showing Sample Collection Area

Methodology

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Microbiological/Bacteriological Analysis/Antibiotic Sensitivity Test

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All glass wires were sterilized in a hot box oven at 160^{o}C for 1hr. Samples were serially diluted with normal saline. Media for selector of microorganisms' isolates were SS agar, N agar, EM agar, MacConkey agar, and Mannitol salt agar and TCBS agar. 0.1ml aliquot was used, Samples were inoculated. The inocular was incubated at 37^{o}C for 2hrs and the special at 45^{o}C for 48hrs.Pour plate technique for culture medium and nutrient agar were used. The nutrient's agar was prepared according to the manufacturer's instruction and allowed to cool to 45°C.

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TBC, TCC and other microorganisms were enumerated, serial dilution and specific dilution into clean and sterilized Nutrient agar plate method was carried out using 0.1ml dilution of 10^{-1} to 10^{-2} of bacteria suspension, and inoculated plate were incubated at 37oc for 28-28hrs at home temperature. Bacteria colonies of the plate were counted randomly and purified by sub-culturing into fresh agar plate using the Streak technique.

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Colonies were sub-cultured to obtain pure isolates which were then characterized by gram's staining for the identification and characterization of bacteria. Biochemical tests (Voges Proskauer, Methyl red, Citrate, Catalase, Coagulase, Oxidase, Motility and Sugar fermentation tests) were respectively carried out. Identity of the isolates was matched with Bergery's Manual of Determinative Bacteriology (BMD) for Confirmation (Holt *et al*.,2002).

The antibiotic sensitivity test was performed by the Kirby-Bauer technique. Discs were commercially prepared for Gram-positive and Gram-negative organisms. The isolate from a fresh culture was cultured for about 18-24hours in peptone water was prepared according to the manufacturers formular. Mueller-Hinton Agar was prepared in line with manufacturer's formular.25 ml of the medium was inoculated in each plate and was allowed to set and solidify. A turbid suspension of the isolates was made in distilled using 0.5 McFarland Standard and prepared as a comparator. A sterile swab was dipped into the bacteria suspension, pressed on the side the sides of the bottle to allow excess drip off, then used to evenly Streak the entire surface of the Mueller-Hinton agar plates. The plates were incubated at 37^{o}C for 24 hours. Zone of inhibition was measured in millimeters, recorded after incubation period and interpreted was grouped using the criteria of the National Committee on Clinical Laboratory Standard (NCCLS) of the World Health Organizations.

Result and Discussion

Microbial Count of Groundwater Sources

In Abuloma, Heterotrophic Bacteria Count, Total Coliform Count and Feacal Coliform Count during the dry season had mean 0.65x10^{6} cfu/ml, 5.53x10^{4} cfu/ml and 1.55x10^{4} cfu/ml while their wet season values were 1.8x10^{6} cfu/ml, 7.25x10^{4} cfu/ml and 2.55x10^{4} cfu/ml respectively. At Borikiri, Heterotrophic Bacteria Count, Total Coliform Count and Feacal Coliform Count during the dry season had mean values of 0.65x10^{4} cfu/ml, 0 cfu/ml and 0 cfu/ml with a corresponding mean value of 1.55x10^{4} cfu/ml, 0.34x10^{4} cfu/ml and 0 cfu/ml during the wet

season. Also at Eagle Island, Total Heterotrophic Count, Total Coliform Count and Feacal Colifirm Count during the dry season had mean values of $10.25x10^6$ cfu/ml, $8.25x10^4$ cfu/ml and $3.4x10^4$ cfu/ml and a wet season value of $11.75x10^4$ cfu/ml, $0.9x10^4$ cfu/ml and $5.2x10^4$ cfu/ml respectively. The mean value for Fimie and Macoba-Isaka for Total Heterotrophic Count, Total Coliform Count and Feacal Coliform Count during the dry season was $0.2x10^6$ cfu/ml, 0 cfu/ml, and 0cfu/ml; $0.6x10^4$ cfu/ml, $0.5x10^4$ cfu/ml and 0 cfu/ml with their corresponding wet season mean values as $0.8x10^4$ cfu/ml, $0.3x10^4$ cfu/ml and 0 cfu/ml; $1.35 x10^4$ cfu/ml, $0.6 x10^4$ cfu/ml and $0.2x10^4$ cfu/ml respectively. Rumuekini and Rumuokoro had a separately dry season mean value of $4.8x10^4$ cfu/ml and $1.25x10^4$ cfu/ml for Total Heterotrophic Bacteria Count dry, $4.25x10^6$ cfu/ml and 0 cfu/ml for Total Coliform Count, $2.4x10^4$ cfu/ml and 0 cfu/ml for Feacal Coliform Count their respective wet season mean values were $9.2x10^4$ cfu/ml and $2.75x10^4$ cfu/ml for Total Heterotrophic Bacteria Count, $4.9x 10^4$ cfu/ml and $0.2x10^4$ cfu/ml for Total Coliform Count and $3.7x10^4$ cfu/ml and 0 cfu/ml for Feacal Coliform Count (Table 2.1, Fig. 1).Total Coliform Count dry season and Feacal Coliform Count dry and wet seasons were significant at $P<0.05$ for Borikiri, Fimie and Rumuokoro while at Macoba-Isaka, there was significance of FCC during the dry season at $P<0.05$ (Table 2.1).

Table 2.1: Variation of Bacteriological Parameters of Borehole Water across samplings Locations

Sample Location	THB Dry Season	Wet Season	TCC Dry Season	Wet Season	FCC Dry Season	Wet Season	t test (p-value)
ABULOMA	$0.65 x10^6$	$1.8 x10^6$	$5.35 x10^4$	$7.25 x10^4$	$1.55 x10^4$	$2.55 x10^4$	0.05
BORIKIRI	$0.6 x10^6$	$1.55 x10^6$	0	$0.3 x10^4$	0	0	0.05
EAGLE ISLAND	$10.25 x10^6$	$11.75 x10^6$	$8.25 x10^4$	$0.9 x10^4$	$3.4 x10^4$	$5.2 x10^4$	0.05
FIMIE	$0.2 x10^6$	$0.8 x10^6$	0	$0.3 x10^4$	0	0	-
MACOBA-ISAKA	$0.6 x10^6$	$1.35 x10^6$	$0.5 x10^4$	$0.6 x10^4$	0	$0.2 x10^4$	-
RUMUEKINI	$4.8 x10^6$	$9.2 x10^6$	$4.25 x10^4$	$4.9 x10^4$	$2.4 x10^4$	$3.7 x10^4$	-
RUMUOKORO	$1.25 x10^6$	$2.75 x10^6$	0	$0.2 x10^4$	0	0	-

TBH=Total heterotrophic bacteria, TCC=Total coliform bacteria, FCC=Fecal coliform count.

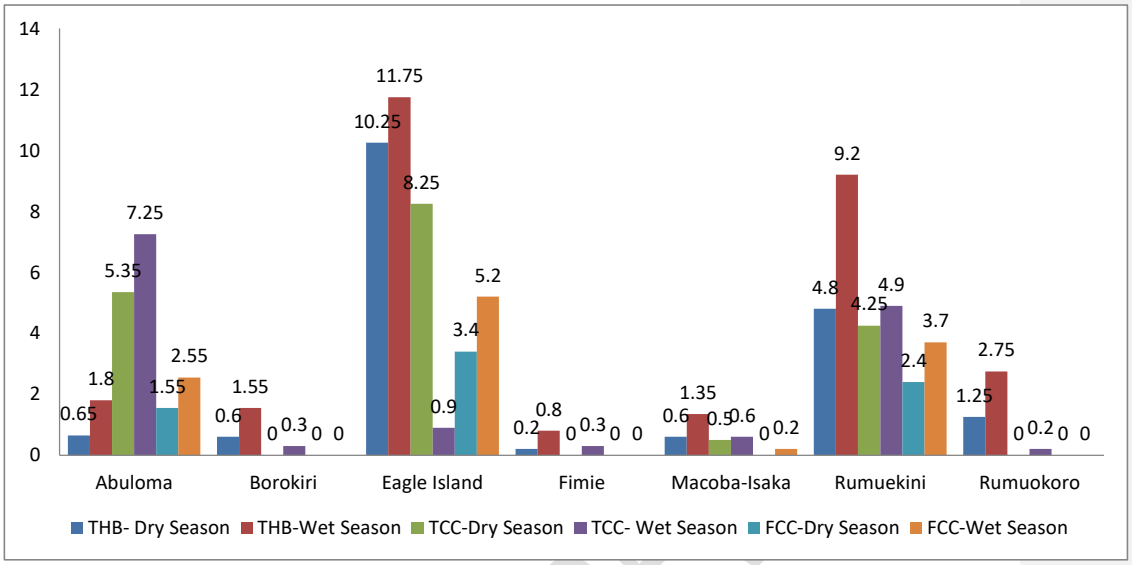

Figure 2: Variation Plot of Bacteriological Parameters of Borehole Water across Sampling Locations

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In Abuloma, Borikiri, Fimie and Eagle Island the mean Vibrio count during the dry season were ($0.8x10^4$) cfu/ml, (0)cfu/ml, ($6x10^4$) cfu/ml, and (0) cfu/ml. Their respective wet season count were 1.25 $x10^4$ cfu/ml ,0 cfu/ml, 0 cfu/ml and 0cfu/ml respectively. The dry season mean value of Macoba-Isaka, Rumuekini and Rumuokoro were 0cfu/ml, $4x10^4$ cfu/ml and 0cfu/ml with a corresponding wet season values of 0cfu/ml,0cfu/ml,$7.15x10^4$ cfu/ml and 0cfu/ml respectively (Table 2.2). Abuloma, Borikiri and Eagle Island showed mean count during the dry season for *Staphylococcus* was $5.5x10^4$ cfu/ml, 0cfu/ml and $6.25x10^4$ respectively. Fimie, Macoba -Isaka, Rumuekini and Rumuokoro had mean values for *Staphylococcus* count varies as 0cfu/ml, $5.5x10^4$ cfu/ml, $5.2x10^4$ and $2.1x10^4$ during the dry season. For the wet season, mean value for *Salmonella* Count in Abuloma, Borikiri and Eagle Island were $7x10^4$ cfu/ml, $0.5x10^4$ cfu/ml and $5.7x10^4$ cfu/ml while Fimie, Macoba-Isaka, Rumuekini and Rumuokoro mean values for the wet season varies from $0.7x10^4$ cfu/ml, $7x10^4$, $4.2x10^4$ and $3.5x10^4$ respectively (Table, 2.2).

The mean count for *Salmonella* during the dry season for Abuloma, Borikiri and Eagle Island were $0.7x10^4$ cfu/ml, 0cfu/ml and 0cfu/ml while the mean *Salmonella* counts for Fimie, Macoba-Isaka, Rumuekini and Rumuokoro mean during dry season were 0cfu/ml, 0cfu/ml, $0.55x10^4$ and 0cfu/ml respectively. More so, for the wet season, Abuloma, Borikiri and Eagle Island had their mean values of $0.8x10^4$cfu/ml,0cfu/ml and 0cfu/ml while for Fimie, Macoba-Isaka, Rumuekini and Rumuokoro, the mean values were from 0cfu/ml,0cfu/ml, 0cfu/ml, $0.55x10^4$ cfu/ml and $0.2x10^4$ cfu/ml respectively. Shigella Count for Abuloma, Borikiri and Eagle Island mean value for dry season ranged from $1.02x10^4$ cfu/ml,$1.02x10^4$ cfu/ml and 0cfu/ml respectively for Fimie, Macoba-Isaka, Rumuekini and Rumuokoro mean value for these locations during the dry season varied from $0.6x10^4$ cfu/ml, 0cfu/ml, $1.15x10^4$ and 0cfu/ml respectively. For mean values for *Shigella* count during the wet season or Abuloma, Borikiri, Eagle Island were $1.02x10^4$ cfu/ml, $1.02x10^4$ cfu/ml, and 0cfu/ml, However, Fimie, Macoba-Isaka, Rumuekini and Rumuokoro location ranged from $0.65x10^4$ cfu/ml, $0.1x10^4$ cfu/ml,$1.15x10^4$ and 0cfu/ml respectively (Table 2.2; Fig. 2).

Table 2.2 Variation of Microbial Parametersof Borehole Water across Sampling Locations.

S L	Vibrio Count		Staphylococcus		Salmonella Count		Shigella Count		t test
	Dry Season	Wet Season	Dry Season	Wet Season	Dry Season	Wet Season	Dry Season	Wet Season	(p value)
AB	0.8×10^4	1.25×10^4	5.5×10^4	7×10^4	0.7×10^4	0.8×10^4	1.02×10^4	1.02×10^4	-
BO	0	0	0	0.5×10^4	0	0	1.02×10^4	1.02×10^4	-
E I	6×10^4	0	6.25×10^4	5.7×10^4	0	0	0	0	0.05
FI	0	0	0	0.7×10^4	0	0	0.6×10^4	0.65×10^4	0.05
MI	0	0	5.5×10^4	7×10^4	0	0	0	0.1×10^4	-
RU	4×10^4	7.15×10^4	5.2×10^4	4.2×10^4	0.55×10^4	0.55×10^4	1.15×10^4	1.15×10^4	0.05
RK	0	0	2.1×10^4	3.5×10^4	0	0.2×10^4	0	0	-

AB=Abuloma, BO=Borikiri, EI=Eagle Island, FI=Fimie, MI=Macoba-Isaka, RU=Rumuekini, RK=Rumuokoro, SL=Sample Location.

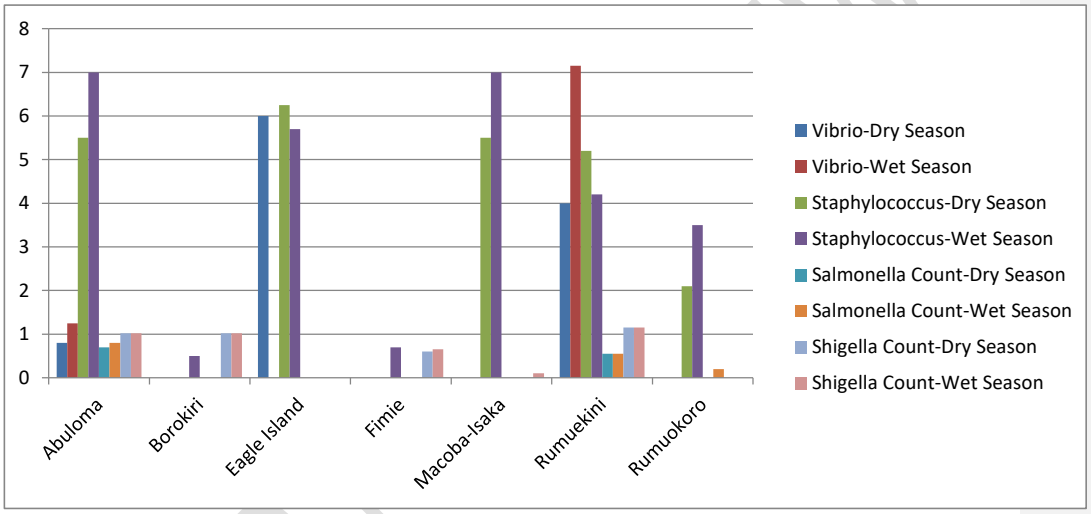

Figure 3: Variation Plot of Microbial Parameters of Borehole Water across Sampling Locations

Distribution of Bacteria Isolates in Groundwater across Sampling Locations

The distribution and the percentage occurrence of bacterial isolates in groundwater showed wide variations. The present of ***staphylococcus sp.*** was observed with 65% in Eagle Island, 56% (Rumuokoro), 25% (Abuloma), 50% (Makoba-Isaka) and 30% (Rumuekini). It was absent in Fimie and Borikiri locations. ***Micrococcus sp.***was present in Eagle Island with 15% occurrence, 15% (Rumuokoro) and 5% (Abuloma). No occurrence of this species in Fimie, Borikiri, Isaka and Rumuekini. ***Shigella sp.*** occurred in Fimie with 30%, 3% in Abuloma and 10% in Rumuekini while there was no occurrence in Eagle Island, Rumoukoro, Borikiri and Isaka. ***Proteus sp,***was present in Eagle Island 20% occurrence, 30% (Rumuokoro), 10% (Fimie), 25% (Borikiri), 5% in Abuloma and 10% (Isaka). No occurrence in Rumuekini. ***E. Coli***occurred only in Abuloma and Rumuekini with 30% and 16% respectively. It was absent in Eagle Island, Rumuokoro, Fimie, Borikiri and Isaka (Table 2.2).The presence of ***Enterobacter sp.***, occurred only in Abuloma with 7% occurrence. All other locations had no occurrences. ***Citrobacter sp.***was present in Abuloma and Rumuekini locations having 5% and 4% separately. It was absent in other locations. ***Pseudomonas sp.*** was present in Abuloma only with 7% occurrence and was

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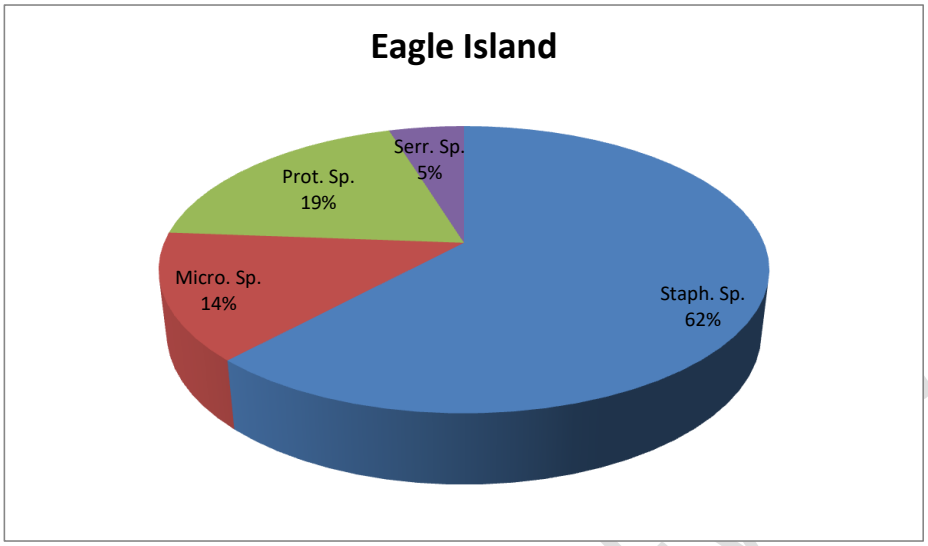
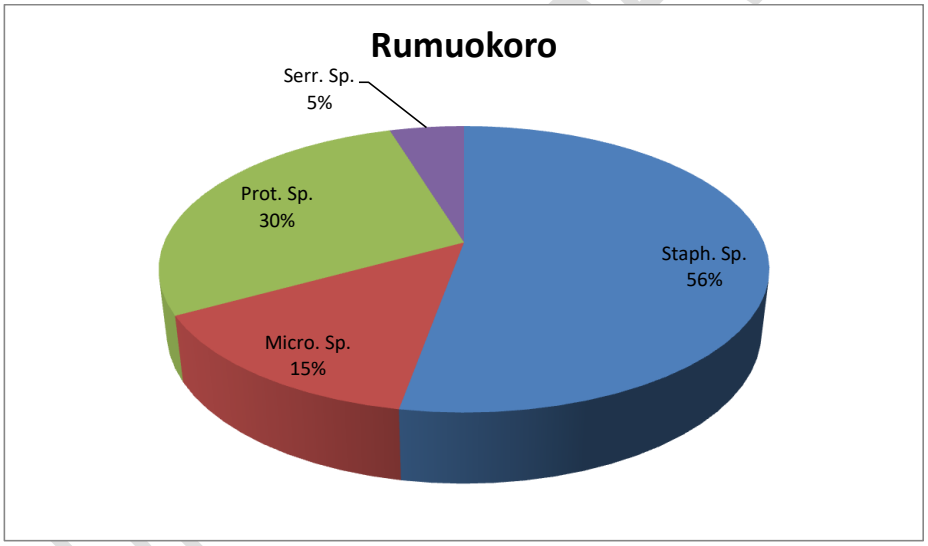
absent in other locations. However, ***Klebsiella sp., Salmonella sp., Bacillus sp., and Vibrio sp.*** were absent in Eaagle Island and Rumuokoro locations. ***Klebsiella sp.***was still absent in Fimie, Borikiri, Abuloma and Rumuekini but was present in Isaka with 30% occurrence. ***Salmonella sp.*** was further absent in Borikiri and Isaka but was present in Fimie (10%), Abuloma (5%) and Rumuekini (10%). ***Bacillus sp.***was further absent in Isaka and was present in other locations with 50% (Fimie), 75% (Borikiri), 3% (Abuloma) and 10% Rumekini. Furthermore, ***Vibrio sp***. was also absent in Fimie and Borikiri but was present in other locations with percentage occurrence as 5% (Abuloma), 10% (Macoba-Isaka) and 20% (Rumuekini) while ***Serratia sp.***was present in Eagle Island and Rumuokoro 5% each but was absent in other locations. The column-wise summation of the percentage occurrence of the isolates for the seven sampling locations defined one hundred percent (100%) signifying a perfect enumeration (Table 2.2).

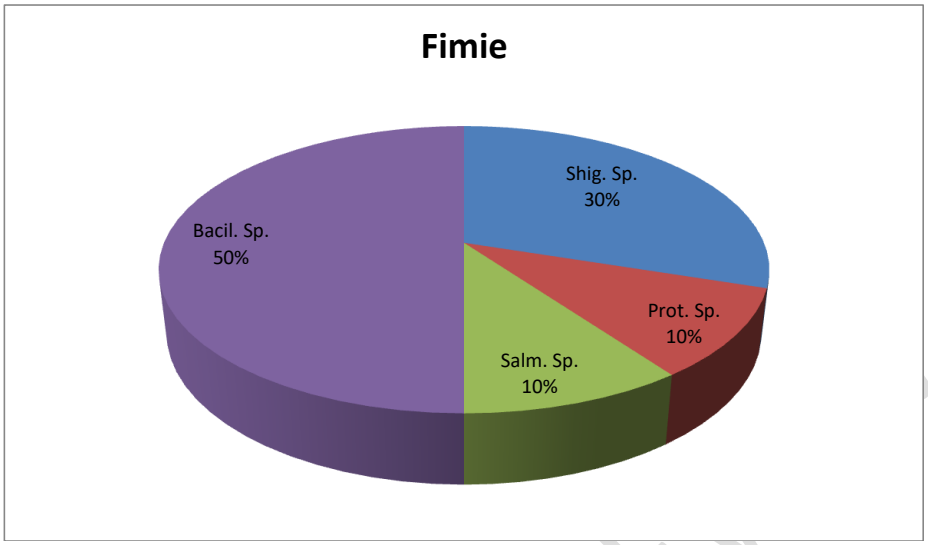
Table 3 Distribution and Percentage Occurrence of Bacterial Isolates in Groundwater across the Seven Sampling Locations

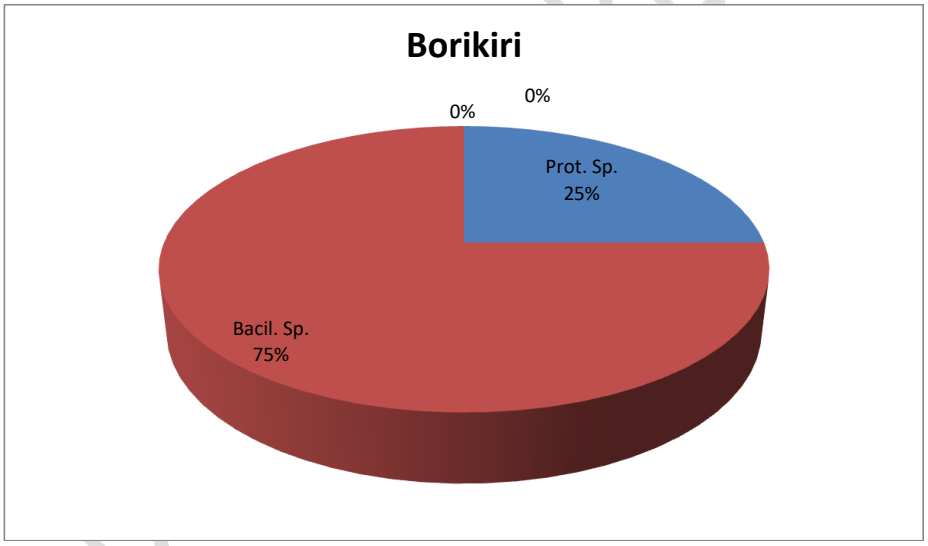
Isolates	**Eagle Island**		**Rumuokoro**		**Fimie**		**Borikiri**		**Abuloma**		**Isaka**		**Rumuekini**	
	O	%	O	%	O	%	O	%	O	%	O	%	O	%
Staph. sp.	+	65	+	56	-	0	-	0	+	25	+	50	+	30
Micro. sp.	+	15	+	15	-	0	-	0	+	5	-	0	-	0
Shig. sp.	-	0	-	0	+	30	-	0	+	3	-	0	+	10
Prot. sp.	+	20	+	30	+	10	+	25	+	5	+	10	-	0
E. coli	-	0	-	0	-	0	-	0	+	30	-	0	+	16
Enter. sp.	-	0	-	0	-	0	-	0	+	7	-	0	-	0
Citro. sp.	-	0	-	0	-	0	-	0	+	5	-	0	+	4
Pseu. sp.	-	0	-	0	-	0	-	0	+	7	-	0	-	0
Klebs. sp	-	0	-	0	-	0	-	0	-	0	+	30	-	0
Salm. sp.	-	0	-	0	+	10	-	0	+	5	-	0	+	10
Bacil. sp	-	0	-	0	+	50	+	75	+	3	-	0	+	10
Vibrio sp	-	0	-	0	-	0	-	0	+	5	+	10	+	20
Serr. sp.	+	5	+	5	-	0	-	0	-	0	-	0	-	0
Total		**100**		**100**		**100**		**100**		**100**		**100**		**100**

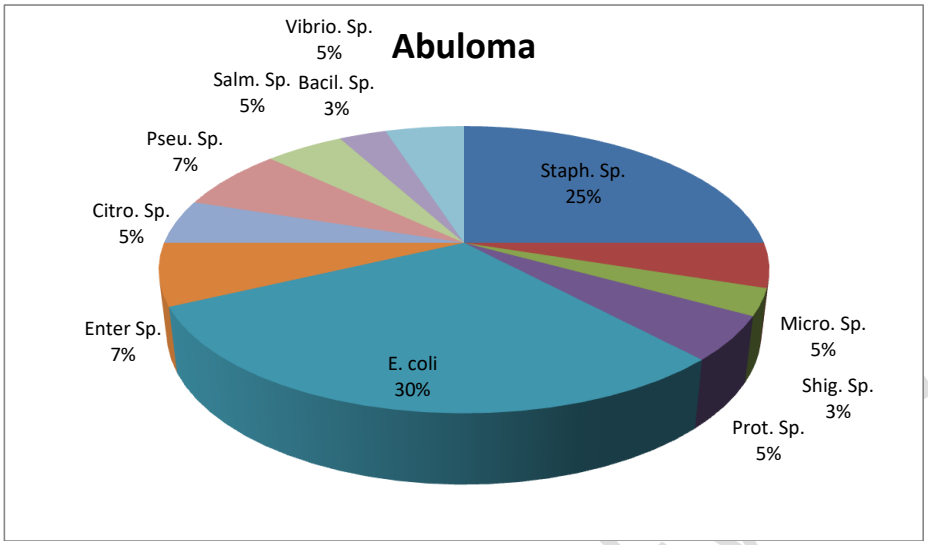
O = Occurrence, % = Percentage of Occurrence, - = Absent, + = Present, *Staph. sp.=Staphylococcus sp., Micro. sp.= Micrococcus sp, Shig. sp.=Shigella sp., Prot. sp.= Proteussp, E. Coli= E.coli, Enter. sp.=Enterobacter sp., Citro. sp.=Citrobacter sp.=Pseu. sp.= Pseudomonas sp.,Klebs. Sp=Klebsiellasp., Salm. sp.=Salmonella sp.,Bacil. Sp.=Bacillus sp.,Vibrio sp., =Vibrio sp., Serr. Sp.=Serratia sp.*

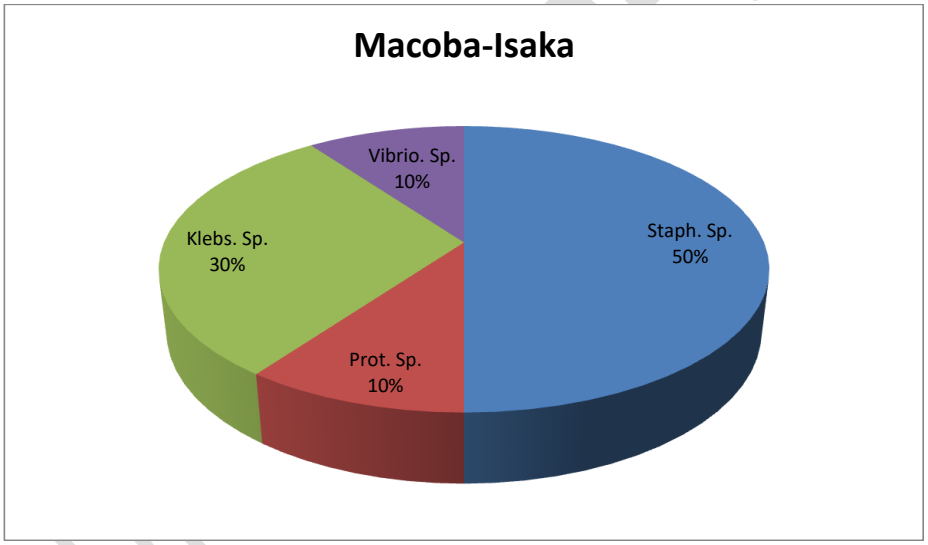

Eagle Island
Serr. Sp.
5%
Prot. Sp.
19%
Micro. Sp.
14%
Staph. Sp.
62%


Rumuokoro
Serr. Sp.
5%
Prot. Sp.
30%
Staph. Sp.
56%
Micro. Sp.
15%


Fimie
Shig. Sp.
30%
Prot. Sp.
10%
Salm. Sp.
10%
Bacil. Sp.
50%


Borikiri
0%
0%
Prot. Sp.
25%
Bacil. Sp.
75%


Abuloma
Vibrio. Sp. 5%
Salm. Sp. 5%
Bacil. Sp. 3%
Pseu. Sp. 7%
Staph. Sp. 25%
Citro. Sp. 5%
Micro. Sp. 5%
Enter Sp. 7%
E. coli 30%
Shig. Sp. 3%
Prot. Sp. 5%


Macoba-Isaka
Vibrio. Sp. 10%
Staph. Sp. 50%
Klebs. Sp. 30%
Prot. Sp. 10%

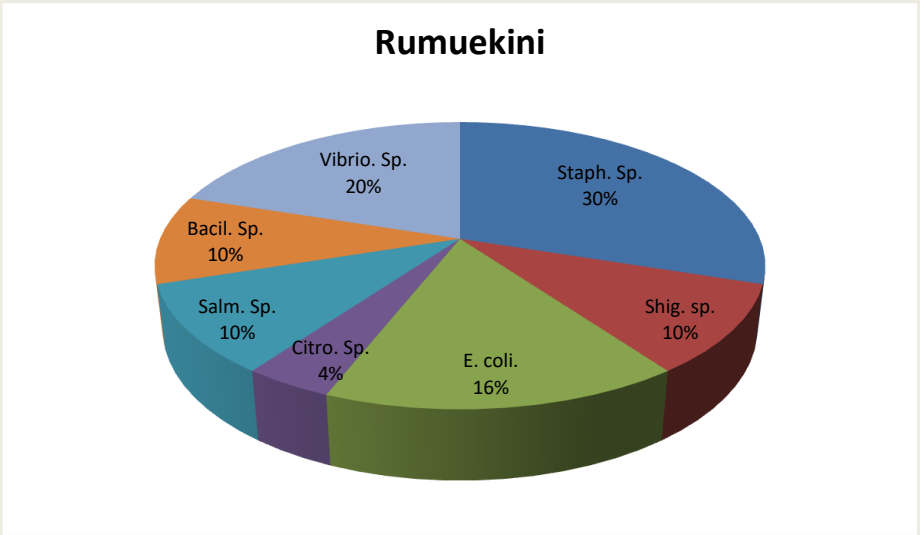


Fig 4. ***Percentage (%) Resistance of E.coli isolated from water sources***

Comment [Adebola M24]: Put figure below each pie chart or if figure 4 covered all, label the chart ABCD

The *E. coli* percentage resistance isolates from the water sources discovered that **Ofloxacin (OFL), Ciproflaxin (CPR), Nitrofurantoin (NIT)** had no value in percentage resistance of the *E coli* isolated. **Augmentin (AUG)** had 50% (Abuloma), 100% (Rumuokoro, Rumuekini, Makoba-Island and Eagle Island). **Ceftazidime** (CAZ) had 100% (Borikiri, Fimie, Abuloma, Rumuokoro, Rumuekini, Makoba-Island and Eagle Island) correspondingly. **Gentamicin (GEN)** had 50% (Abuloma), 60% (Rumuekini), 100% (Rumuokoro, Makoba-Island and Eagle Island). **Cefixime (CXM)** had 100% (Borikiri, Fimie, Abuloma, Rumuokoro, Rumuekini, Makoba-Island and Eagle Island) while **Cefuroxime (CRM)** had 100% for (Borikiri, Fimie, Abuloma, Rumuokoro, Rumuekini, Makoba-Island and Eagle Island) respectively (Fig. 4).

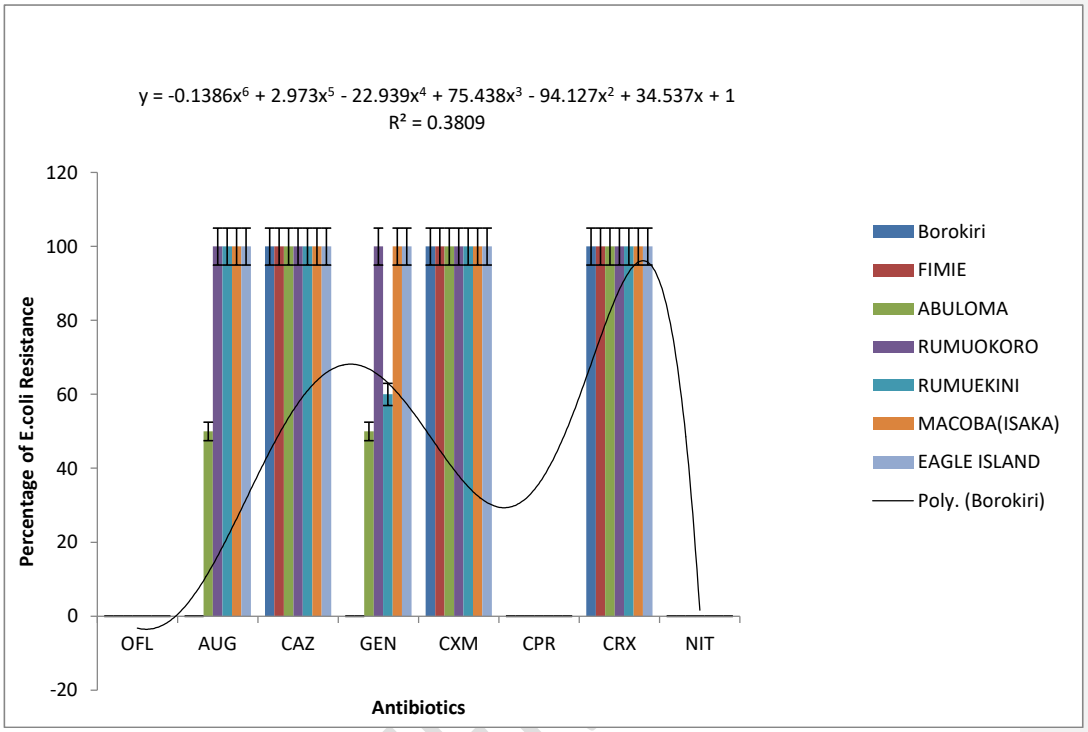
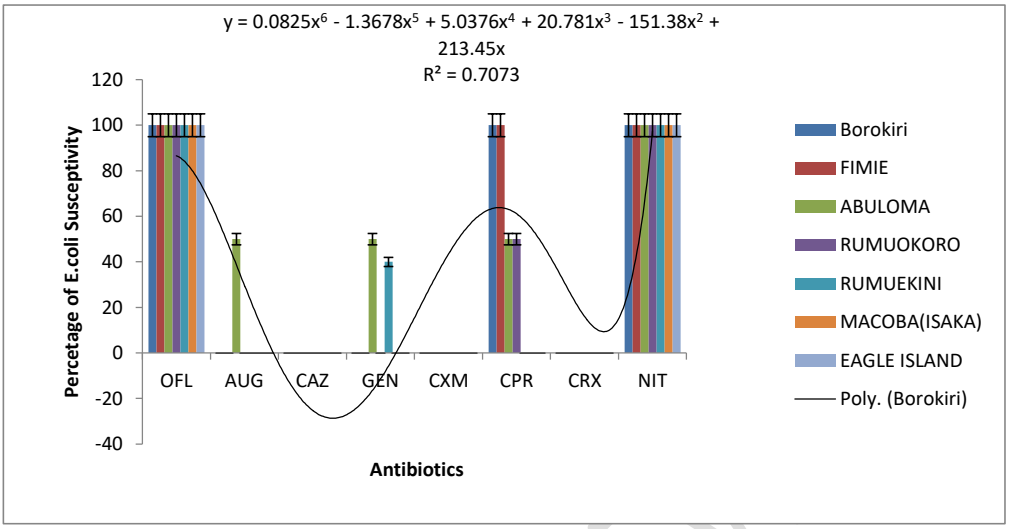


Fig. 5. Percentage (%) of Resistance of *E.coli* isolated from seven location to various commonly use Antibiotics

Percentage (%) of Susceptibility of E.coli isolated from water sources

The susceptibility in percentage of the *E coli* isolated from the seven water sources under investigation revealed that **Ofloxacin** had percentage value as 100% (Fimie, Rumuokoro, Rumuoekini, Abuloma, Makoba-Isaka, Eagle Island and Borikiri respectively). **Augmentin** had 50%in Abuloma. There was no percentage value for **Cetazidime, Cefiximeand Cefuroxime**. Gentamicin appeared 50% (Abuloma) and 40% (Rumuekini) while **Ciproflaxin** appeared as 100% (Fimie and Borikiri) and 50% (Rumuokoro and Abuloma) respectively. **Nitrofurantoin** had percentage values as 100% for Abuloma, Borikiri, Eagle Island, Fimie, Makoba-Isaka, Rumuekini and Rumuokoro correspondingly (Fig. 5).



(OFL)=Ofloxacin,(AUG)=Augmentin,(CAZ)=Cetazidime,(GEN)Gentamicin(CXM)=Cefiximine (CPR)=Ciproflaxin, (CRX)=Cefuroxime, (NIT)=Nitrofurantoin.

Figure 6 Susceptibility (%) of *E.coli* isolated from seven location to various commonly use Antibiotics

Conclusion

Comment [Adebola M25]: Take note of all these corrections
1. You have not discuss your results. To do this, identify your finding step wisely, make your deduction based on your findings and relate it to previous published research work.
2. Make the conclusion by following the objectives in a chronological order.
3. Do not cite any reference in conclusion
4. Give a few recommendation of your work.

An antibiogram shows the aggregate number of bacteria tested against antimicrobial and incorporates the extent of bacterial isolates vulnerable to every antimicrobial operator tested (Fridkin et al., 2001). In the current study, Ofloxacin, Nitrofurantoin and Cefixmine were most apparently susceptible antibiotics among others while Augumentin, Ceftazidine, Gentamycin, Ciproflaxin were observed to be most resistance antibiotic for the ***Escherichia* coli** from the groundwater sampled. Gentamycin appeared as intermediates in Abuloma and Rumuekinki which could also mean non susceptibility by the bacteria possibly at forthcoming time to this antibacterial agent. The increased rate of resistance antibiotics in this study could be attributed to widespread misuse of various antibiotics without control. This is in line with Fanuncio and Nuneza (2015). Consequently, it could be recommended that groundwater source (boreholes) be treated before it should be acceptable for domestic use. It should be sited away from septic tanks source as possible interference under the influence of proximity become inevitable.

COMPETING INTERESTS DISCLAIMER:

Authors have declared that no competing interests exist. The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of

knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

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