Evaluation of Phytochemical contents and antioxidant activity of *Mentha spicata* (spearmint) Grown in Kano, Nigeria

#### **ABSTRACT**

Aim: *Mentha spicata* (spearmint) is a plant that spreads mainly in the temperate and sub-temperate zones of the world including Africa and is used in pharmaceutical and food industries. It has numerous medicinal importances such as antibiotic, anti-inflammatory, anti-diabetic, anti-hyperlipidaemic and many more. This research examines the phytochemical and antioxidant property of *Mentha spicata*. Methodology: Sample of *Mentha spicata* leaves were collected fresh from the cultivated farm and was extracted using aqueous solvent prior to analysis. The phytochemicals determination was carried out in accordance with standard methods while the antioxidant property was evaluated using Diphenylpicrylhydrazyl (DPPH) radical scavenging assay. Results: The result showed that the aqueous extract contain alkaloids, phenols, flavonoids, tannins, saponins glycosides, Terpenoids and anthraquinones at varying concentrations. The antioxidant activity was measured in percentage at different concentration of the extract as follows; 25mg/dl (20.9%), 50mg/dl (10.78%), 75mg/dl (50.7%) and 100mg/dl (17.80%) respectively. The highest antioxidant activity was observed in 75mg/dl concentration. Conclusion: The antioxidant activity observed could be attributed to the presence of flavonoids and phenols. Therefore, Mentha spicata can be used as a source of antioxidant and could improve the health status of individuals.

Key words: Mentha spicata, Evaluation, phytochemicals, antioxidant,

#### 1. Introduction

The use of traditional herbs is getting more attention owing to general believe that the traditional herbs posses medicinal attributes among which are antioxidant, anti-diabetic, anticancer, antimicrobial, hypolidemic and so on. Extracts of traditional herbs are often providing opportunities for drug discovery [1]. Therefore, various parts of different types of plants are been used in the treatment of diseases [2]. Spearmint (Mentha spicata) is a member of Lamiaceae that spreads mainly in the temperate and sub-temperate zones of the world including Asia, Africa, Australia, North America, and Europe [3]. It is regarded as an important source of essential oils that is widely used as a raw material in pharmaceutical industries as well as food production. The genus *Mentha* includes 25 to 30 species such as spearmint, peppermint, wild mint, corn mint, curled mint, bergamot, American mint, Korean mint, etc. of which spearmint is the most common of them [4, 5]. Various parts of Mentha species are used in folk medicine over a long period of time in the treatment of many dysfunctions such as those of gastrointestinal tract [6]. Mentha spicata (spearmint) has been reported to possess many medicinal properties such as antibacterial and antioxidant [3, 7, 8], stomach pain-relieving agent, antispasmodic, digestive and carminative [9], hypolipidemic and anti-diabetic [10], and so on.

Research has shown that the mint main chemical compounds includes limonene, cineole, menthone, menthofuran, isomenthone, menthylacetate, isopulegol, menthol, pulegone and carvone [11]. Areias [12] has however reported mint to be rich in other constituents such as flavonoid glycoside (eg. Narirutin, Luteolin-7-o-rutinoside, Isorhoifolin and Hesperidin), polyphenols (e.g. Rosmaric acid, Eriocitrin, Cinamic acid, Caffeic acid and Narigenin-7-oglucoside); luteolin-diglucoronide and eriodictyol glucopyranosyl-rhamnopyranoside. The amount of mint compounds is different in various species [13]. Various factors including physiological variations, environmental conditions, geographic differences and genetic factors cause differences in chemical composition of these plants [13]. This

Comment [HC1]: importance

**Comment [HC2]:** kindly mention the name of the solvent

**Comment [HC3]:** kindly mention name of the source for method adopted

Comment [HC4]: gaining

Comment [HC5]: not required

Comment [HC6]: reference please

Comment [HC7]: not required

study is therefore aimed at determining the phytochemical and antioxidant property of Mentha spicata (Spearmint).

# 2. MATERIALS AND METHODS

#### 2.1 Sample collection and processing

Fresh leaves of spearmint were collected from the cultivated farms of Aminu Kano College of Islamic and Legal Studies and the T-junction of Kofar Famfo within Kano metropolis. The samples were identified and authenticated prior to analysis and then freeze dried. The dried leaves were ground in to fine powder and then used for the analysis.

## 2.2 Extraction of Spearmint Extract

Extraction of the powder Spearmint was conducted using water (Aqueous). The extracts were concentrated in vacuum using rotary evaporator. After concentration, the extract was allowed to dry and kept for further analysis as described by [14, 15].

# 2.3 Preliminary Phytochemical Screening

Qualitative phytochemical tests were carried out by using standard procedure of preliminary phytochemical screening in accordance with the standard methods described by [14, 15]. The screening was carried out for alkaloids, polyphenols, flavonoids, tannins, saponins, glycosides, Terpenoids, anthraquinones and anthocyanins.

# 2.4 Qualitative Analysis on Phytochemical Constituent of Mint Leaves

5g of the *Mentha spicata* extract was weighed and dissolved in 50ml of distilled water. The mixture was shaken gently and was allowed to dissolve for about five (5) minutes and then filtered. The filtrate was then subjected to the following qualitative tests.

### 2.4.1 Test for Alkaloids

Few drops of 1% HCl were added to the filtrate to which 5 drops of freshly prepared Dragen dorrf"s reagent (potassium bismuth iodide solution) was added. The solution (1 cm<sup>3</sup>) was treated with Wagners reagent (solution of iodine in potassium iodide). Formation of a precipitate indicated the presence of alkaloids [15].

## 2.4.2 Test for Anthraquinones

5ml of the filtrate was hydrolyzed with diluted Conc.  $H_2SO_4$ . 1 ml of dilute ammonia was added to it. Rose pink coloration suggested the positive response for anthraquinones [15].

## 2.4.3 Test for Phenols

Addition of 3 drops of 5% lead acetate to the filtrate leads to the formation of yellow precipitates indicating the presence of phenolic compounds [15].

# 2.4.4 Test for Tannins

0.5g of the dried powdered of Mentha spicata extract was boiled in 20 cm<sup>3</sup> of water in a test tube and then filtered. A few drops of 0.1% ferric chloride were added. Formation of brownish green coloration indicated the presence of tannins [15].

**Comment [HC8]:** more details required, like which equipment used, temperature for storage, labeling and transportation mechanism, training of staff

**Comment [HC9]:** more details like type of water, mention name of source of water if company purchased or in house prepared

Comment [HC10]: name of equipment

**Comment [HC11]:** in brief kindly mention the storage technique

**Comment [HC12]:** name the std method version year

**Comment [HC13]:** terpenoids all names in small letters

Comment [HC14]: weighing machine name

Comment [HC15]: name of equipment

Comment [HC16]: name of filter

**Comment [HC17]:** if possible add photos of each reaction

**Comment [HC18]:** exact volume in micro litres to be specified

**Comment [HC19]:** exact volume to be mentioned

Comment [HC20]: attach photos if possible

### 2.4.5 Test for Saponins

2 g of the powdered Mentha spicata extract was boiled in 20 ml of distilled water in a water bath and then filtered. 10 cm<sup>3</sup> of the filtrate was mixed with 5 ml of distilled water and shaken vigorously for a stable persistent froth. The frothing was mixed with 3 drop s of olive oil and shaken vigorously; formation of emulsion indicates the presence of saponin [15].

#### 2.4.6 Test for Flavonoids

5 ml of dilute ammonia solution were added to a portion of the aqueous filtrate of the powdered Mentha spicata extract followed by addition of concentrated H<sub>2</sub>SO<sub>4</sub>. A yellow coloration observed in each filtrate indicated the presence of flavonoids. The yellow colouration disappeared on standing [15].

# 2.4.7 Test for Terpenoids (Salkowski Test)

5ml of the filtrate of Mentha spicata extract was mixed in  $2 cm^3$  of chloroform;  $3 cm^3$  of concentrated  $H_2SO_4$  was carefully added to form a layer. A reddish brown coloration of the interface was formed to show positive results for the presence of terpenoids.

#### 2.4.8 Test for Cardiac Glycosides (Keller-Killani test)

5ml of the filtrate of the filtrate of Mentha spicata extract was treated with 2 cm<sup>3</sup> of glacial acetic acid containing one drop of ferric chloride solution. This was then treated with 1 cm<sup>3</sup> of concentrated sulphuric acid. A brown ring of the interface indicated a deoxysugar characteristic of cardenolides [15].

# 2.5 Quantitative Analysis of Phytochemical Constituent

Six of the phytochemicals namely; alkaloids, phenols, flavonoids, tannins, saponnins and cardiac glycosides were subjected quantitative analysis using standard methods.

## 2.5.1 Determination of Alkaloids

5 g of the extract of *Mentha spicata* was weighed into a 250 cm<sup>3</sup> beaker and 200 cm<sup>3</sup> of 10% acetic acid and ethanol was added and covered and was allowed to stand for 4 hr. This was filtered and the extract was concentrated on a water bath to one-quarter of the original volume. Concentrated ammonium hydroxide was added drop wise to the extract until the precipitation was complete. The whole solution was allowed to settle and the precipitate was collected and washed with dilute ammonium hydroxide and then filtered. The residue was the alkaloid, which was dried and weighed [15].

## 2.5.2 Determination of Total Phenols by spectrophotometric method

The fat free sample was boiled with 50ml of ether for the extraction of the phenolic component for 15 minutes. 5 cm<sup>3</sup> of the extract was pipetted into a 50 cm<sup>3</sup> flask, 10 cm<sup>3</sup> of distilled water was added. This was followed by the addition of 2 cm<sup>3</sup> of ammonium hydroxide solution and 5 cm<sup>3</sup> of concentrated amyl alcohol.. The samples were made up to mark and left to react for 30 minutes for color development. This was measured at 505nm [15].

#### 2.5.3 Determination of Tannins

Comment [HC21]: time taken for reaction

Comment [HC22]: equipment used

500 mg of the Mentha spicata extract was weighed into a 50 cm<sup>3</sup> plastic bottle followed by the addition of 50 cm<sup>3</sup> of distilled water and the mixture was shaken for 1hr in a mechanical shaker. This was filtered into a 50 cm<sup>3</sup> volumetric flask and made up to the mark. Then 5 cm<sup>3</sup> of the filtrate was pipetted out into a test tube and mixed with 2 cm<sup>3</sup> of 0.1 M FeCl<sub>3</sub> in 0.1N HCl and 0.008 M potassium ferrocyanide. The absorbance was measured at 720 nm within 10min [15].

#### 2.5.4 Determination of Saponin

20 g of Mentha spicata extract was placed into a conical flask and 100 cm3 of 20% aqueous ethanol was added. The mixture was heated over a hot water bath for 4hr with continuous stirring at about 55°C. The mixture was filtered and the residue re-extracted with another 200 cm³ of 20% ethanol. The combined extracts were reduced to 40ml over water bath at about 90°C. The concentrate was transferred into a 250 cm³ separating funnel and 20 cm³ of diethyl ether was added and shaken vigorously. The aqueous layer was recovered while the ether layer was discarded. The purification process was repeated. 60 cm³ of n-butanol was added. The combined n-butanol extracts were washed twice with 10 cm³ of 5% aqueous sodium chloride. The remaining solution was heated in a water bath. After evaporation the solution was dried in the oven to a constant weight [15].

# 2.5.5 Test for Flavonoids

10 g of Mentha spicata extract was extracted repeatedly with 100 cm³ of 80% aqueous methanol at room temperature. The whole solution was filtered through whatman filter paper No 42 (125 mm). The filtrate was later transferred into a crucible and evaporated into dryness over a water bath and weighed to a constant weight [15].

## 2.5.6 Determination of Cardiac Glycosides

A tincture of the sample was prepared by preparing 10% extract in 70% alcohol by shaking 1g of pulverized Mentha with 10 cm<sup>3</sup> 70% alcohol. The mixture was left overnight with occasional shaking for 2hr and then filtered. 10 cm<sup>3</sup> of the purified filtrate transferred in to a dry stopped Erlynmeyer flask was added to 10 cm<sup>3</sup> of Baltet's reagent. The blank was prepared at the same time using 10 cm<sup>3</sup> of distilled water instead of the purified filtrate and 10 cm<sup>3</sup> of Baltet's reagent. This was made to stand for 1hr, for maximum color development. The solution was diluted with 20 cm<sup>3</sup> of distilled water and mixed. The intensity of the color obtained was measured at 495nm using spectrophotometer. The color was stable for several hours. The difference between experiment and blank (E-B) is equal to the original reading. The percentage total glycoside was calculated using the absorptivity of digitoxin = 170, similarly treated at 495 nm as follows:

% Total cardiac glycoside = (A x 100 / 17) g% Calculated as digitoxin.

Where A = absorbance of the colour at 495nm.

# 2.6 Determination of Antioxidant Activity

For antioxidant activity determination, 20g of Mentha spicata extract was dissolved in 100 cm<sup>3</sup> absolute methanol and soaked overnight. The resulting solution was filtered and transferred to clean vessel and subsequently evaporated to dryness in order to obtain the extract [16].

# 2.6.1 DPPH Radical Scavenging Activity

Diphenylpicrylhydrazyl (DPPH) radical scavenging activity was measured as described by [17] with some modifications. The reaction mixture up to 3 cm³ containing 0.2 cm³ of DPPH and 2.8 cm³ of test solution at various concentrations i.e. (20, 40, 60, 80,100mg/ml) of the extract fractions was incubated at 37°C for 30minutes. The

absorbance of the resulting solution was measured at 517nm using spectrophotometer (Beckman model DU-40) The percentage inhibition of DPPH radical scavenging activity was calculated by comparing the results of the test with those of the control (not treated with extract) using the following equation: DPPH scavenging activity = (Ac–As)/  $Ac \times 100$  Where Ac=Absorbance of control, As=Absorbance of sample [18].

# 2.7 Statistical Analysis

Results were analyzed statistically using Analysis of variance (ANOVA). Difference of mean values were determined at P < 0.05 level of significance.

### 3. RESULTS

# 3.1 Phytochemical Contents

The results for qualitative and quantitative phytochemical constituents of aqueous extract of Mentha spicata are presented in tables 1 and 2.

Table 1: Qualitative Phytochemical Constituents of Mentha spicata grown in Kano, Nigeria

Phytochemicals	Status
Alkaloids	+
Polyphenols	+
Flavonoids	+
Tannins	+
Saponins	+
Glycosides	+
Terpenoids	+
Anthraguinones	+

Table 2: Quantitative Phytochemical Constituents of Mentha spicata grown in Kano, Nigeria

18 0.08 04
04
15
47
04

# 3.2 Antioxidant Activity

Comment [HC23]: sr no column kindly add

The antioxidant activity of Mentha spicata extract used in the study was measured using Diphenylpicrylhydrazyl radical (DPPH) scavenging assay. This was measured at various concentrations in mg/dl (i.e 25, 50, 75 and 100). The highest antioxidant activity was observed at 75 mg/dl concentration (table 3). The antioxidant activity is based on the reduction of DPPH free radical by an antioxidant [16].

Table 3: Antioxidant Activity of Mentha spicata Extract Grown in Kano, Nigeria

Concentration in mg/dl	% Inhibition
25	20.91
50	10.78
75	50.70
100	17.80

### 4. DISCUSSION

As shown in the table 1 above, the aqueous extract of Mentha *spicata* showed the presence of eight phytochemicals, namely alkaloids, phenols, flavonoids, tannins, cardiac glycosides, saponins, terpenoids and anthraquinones. Six of the eight phytochemicals determined were subjected to quantitative analysis and the results indicated that the extract of Mentha spicata contain varying amount of phytoconstituents such as flavonoids  $(8.29 \pm 0.18 \text{ mg/g})$ , alkaloids  $(32.09 \pm 0.08 \text{ mg/g})$ , phenols  $(7.96 \pm 0.04 \text{ mg/g})$ , tannins  $(2.15 \pm 0.15 \text{ mg/g})$ , saponins  $(1.51 \pm 0.47 \text{ mg/g})$  and glycosides  $(2.54 \pm 0.04 \text{ mg/g})$  (table 2). This phytochemicals have various medicinal properties. *Mentha spicata* possesses astringent, antiseptic, antispasmodic, anti-inflammatory, analgesic, and anti-carcinogenic properties [19]. The anti-inflammatory effect could be attributed to the presence of alkaloids, flavonoids, phenolic acids, and tannins [20, 21, 22, 23]. The anti-carcinogenic activity could be attributed to the presence of Terpenes and saponin [24, 25]. The analgesic properties can be attributed to the presence of some alkaloids such as morphine [26]. The antispasmodic effect may result from the presence of phenolic acids [20]. The astringent property of Mentha spicata is perhaps due to the presence of tannins [27], which may also be responsible for the antiseptic effect [22]. The present study is in agreement with various studies that plants contain a vast number of phytochemicals [28, 29].

The concentration of flavonoids  $(32.09 \pm 0.08 \text{ mg/g})$  and phenols  $(7.96 \pm 0.04 \text{ mg/g})$  determined in the current study are higher than those reported by [30]. Plant polyphenols such as flavonoids have vast array of biological activities, including anti-inflammatory, anti-oxidative and free radical-scavenging property [31]. Dietary flavonoids possess anticancer, anti-inflammatory and anti-infectious properties and can be used to prevents obesity and as potential anti Covid-19 [32].

The antioxidant activity of the Mentha spicata extract used in this study has indicated that, the extract possess an antioxidant property. However, the result showed that Mentha spicata exhibits the highest antioxidant activity at 75 mg/dl concentration. Antioxidants are indispensable for cellular response in order to deal with oxidative stress under physiological conditions and can be effective through several ways [33]. They act as preventive (preventing the formation of lipid free radicals), chain breaking antioxidants (interfering with the circulation of the autoxidation chain reaction), as singlet oxygen quenchers, through synergism with other antioxidants; as reducing agents which convert hydroperoxides into stable compounds; as metal chelators that convert metal pro-oxidants (iron and copper derivatives) into stable products; and finally as inhibitors of pro-oxidative enzymes (lipoxygenases) [34].

The higher antioxidant activity (50.70 mg/dl) observed in this study could be attributed to the presence of flavonoids and phenolic compounds in Mentha spicata extract. Flavonoids have multi beneficial activities including an antioxidant activity [35]. The antioxidant activities of flavonoids and phenolic acids and further suggested that extracts of plants containing flavonoids and phenolic acids can be considered as promising antiaging sources for use in cosmetic formulations [35].

#### 5. Conclusion

Extract of Mentha spicata is rich in phytochemicals that have numerous medicinal importance that could promote health and can be used as a source of antioxidants. The medicinal properties of Mentha spicata could therefore be attributed to the presence of phytochemicals determined.

**Comment [HC24]:** can include names of chemicals and their benefit

#### COMPETING INTERESTS DISCLAIMER:

Authors have declared that no competing interests exist. The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

#### REFERENCES

Rabi R. A., Abdulmumin Y. and Abdulmumin T. M. (2020). Characterization and Physico-Chemical Property of River Red Gum (Eucalyptus camaldulensis) Leave Oil. Sch Int J Tradit Complement Med 3(3): 39-45

Abdulmumin, Y., Abdulmumin1, T. M., Muhammad, I. A., Murtala, M., Dalhatu, M. M., Amina, L. A., Bichi, S. A. and Sarki, S. I. (2021). Bioassay Guided Fractionation, Phytochemicals and Toxicity Evaluation of Eucalyptus camaldulensis Leave Extracts. *SARJNP* 4(2): 36-43

Jianu, C., Stoin, D., Cocan, I., David, I., Pop, G., Lukinich-Gruia, A. T., Mioc, M., Mioc, A., S, oica, C., Muntean, D. (2021). *In Silico* and *In Vitro* Evaluation of the Antimicrobial and Antioxidant Potential of Mentha smithiana R. GRAHAM Essential Oil from Western Romania. *Foods* 10:815

Choudhury, R. P., Kumar, A. and Garg, A. N. (2006). Analysis of Indian mint (*Mentha spicata*) for essential, trace and toxic elements and its antioxidant behaviour. *J. Pharm. Biomed. Anal.* 41:825–32.

Arumugam, P., Priya, N. G., Subathra, M. and Ramesh, A. (2008). Anti-inflammatory activity of four solvent fractions of ethanol extract of *Mentha spicata* L investigated on acute and chronic inflammation induced rats. *Environ. Toxicol. Pharmacol.* 26:92–

Mogosan, C., Vostinaru, O., Oprean, R., Heghes, C.; Filip, L.; Balica, G.; Moldovan, R.I. A comparative analysis of the chemical composition, anti-inflammatory, and antinociceptive effects of the essential oils from three species of Mentha cultivated in Romania. Molecules 2017, 22, 263.

Hussain, A. I., Anwar, F., Shahid, M., Przybylski, R., Hussain, A. I., Anwar, F., Shahid, M., Ashraf, M., Hussain, A. I., Anwar, F., Shahid, M., Ashraf, M. and Przybylski, R. (2011). Chemical Composition, and Antioxidant and

Antimicrobial Activities of Essential Oil of Spearmint (Mentha spicata L.) From Pakistan Chemical Composition, and Antioxidant and Antimicrobial Activities of Essential Oil of Spearmint (Mentha spicata L.) From Pakistan. 2905. https://doi.org/10.1080/10412905.2010.9700269

Hajjaj, Y. P. M., Yousuf, N. N., Shohel, M., Bhattacherjee, R., Kumas, D. B. (2013). Analgesic, anti-inflammatory and antipyretic effect of *Mentha spicata* (spearmint) Br. J. Pharm. Res. 3:854–64

Adelpoor, M. J. and Golparvar, A. R. (2013). Chemical composition of essential oils of three ecotypes of *Mentha spicata* L from Kohgiluyeh va Boyer-Ahmad Province, Iran. Iran.J. Herb Drugs. 4:63–8

Mahsan, B., Mahmood, A. and Ashkan, J. J. (2017). Study of Hypoglycemic, Hypocholesterolemic and Antioxidant Activities of Iranian *Mentha Spicata* Leaves Aqueous Extract in Diabetic Rats. *Iran J Pharm Res.* 16(Suppl): 75–82.

Mascher, H., Kikuta, C. and Schiel, H. (2001). Pharmacokinetics of menthol and carvone after administration of an enteric coated formulation containing peppermint oil and caraway oil. *Arzneimittelforschung* 51: 465-469

Areias, F., Valentao, P., Andrade, P., Ferreres, F. and Seabra, R. (2001). Phenolic fingerprint of peppermint leaves. *Food chem*73: 307-311.

Park, Y. J., Baskar, T. B, Yeo, S. K., Arasu, M. V. and Al-Dhabi, N. A. (2016). Composition of volatile compounds and in vitro antimicrobial activity of nine *Mentha spp. Springer plus* 5: 1628.

Gambo, I., Rabi, R. A., Abdulmumin, Y., Abdulmumin, T. M. and Dalhatu, M. (2020). *Comparative Analysis on Phytochemicals , Proximate composition and Mineral Element of Aqueous Seed Extract of Three Varieties of Wheat Cultivated in Kazaure*. 5(3):292–296.

Velvan, S. (2015). Phytochemical Techniques- a review. world Journal of Science and Research 1(2):80-91.

Ibrahim, A. M., Anwar A.Y., Sani, M. A., Ya'u, S. A., Tasi'u, A. M., Sani M.Y3, Abdulmumin Y, Murtala, M., Musa, H., Sadiya, A. B., Abdullahi, N., Maimuna, D. M., Salisu, A. A. and Tasi'u, M. (2020). Assessment of Antioxidant Activity and Mineral Elements Composition of Fenugreek Seed Extract. *Dutse Journal of Pure and Applied Sciences (DUJOPAS)*, 6:2

Blois, M. S. (1958). Antooxidant determinations by the use of a stable free radical. Nature 181:1199 - 1200

Molyneux, P., (2004). "The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity". Songklanakarin J. sci. technol, 26(2):211-219.

Chawla, S. and Thakur, M. (2014). Effect of thermal processing on total phenolic content and antioxidant activity of *Mentha* leaves. *Asian J. Bio. Sci.*, 9 (2): 200-203.

Ghasemzadeh, A., Jaafar, H. Z. E. and Rahmat, A. (2010). Antioxidant activities, total Phenolics and flavonoids content in two varieties of Malaysia Young Ginger (*Zingiberofficinale Roscoe*). *Molecules*. 15: 4324-4333.

Tapas, A. R., Sakarkar, D. M. and Kakde, R. B. (2008). Flavonoids as Nutraceuticals: A Review. *Tropical Journal of Pharmaceutical Research*. 7: 1089-1099.

Dolara, P., Luceri, C., De Filippo, C., Femia, A.P., Giovannelli, L., Carderni, G., Cecchini, C., Silvi, S., Orpianesi, C. and Cresci, A. (2005). Red wine polyphenols influence carcinogenesis, intestinal microflora, oxidative damage and gene expression profiles of colonic mucosa in F344 rats. *Mutation Research*. 591: 237–46.

Mottaghipisheh, J. and Stuppner, H. A. (2021). Comprehensive Review on Chemotaxonomic and Phytochemical Aspects of Homoisoflavonoids, as Rare Flavonoid Derivatives. *Int. J. Mol. Sci.* 22:2735. https://doi.org/10.3390/ijms22052735

Langenheim, J. H. (1994). Higher plant terpenoids: A phytocentric overview of their ecological roles. *Journal of Chemical Ecology*. 20: 1223-1280.

George, F., Zohar, K., Harinder, P.S. and Klaus, B. (2002). The Biological Action of Saponins in Animal Systems: A review. *British Journal of Nutrition*. 88: 587–605.

Saxena, Y., Saxena, M., Nema, R., Singh, D. and Gupta, A. (2013). Phytochemistry of Medicinal Plants. *Journal of Pharmacognosy and Phytochemistry*. 1(6):168-182.

De Bruyne, T., Pieters, L., Deelstra, H. and Vlietinck, A. (1999). Condensed vegetables tannins: biodiversity in structure and biological activities. *Biochemical System Ecology*. 27: 445–59.

Ullah, N., Khurram, M., Amin, M. U., Afridi, H. H., Khan, F. A., Khayam, M. U., Ullah, S., Najeeb, U., Hussain, J., and Asif Khan, M. (2011). Comparison of Phytochemical constituents and antimicrobial activities of *Mentha spicata* from four northern districts of Khyber Pakhtunkhwa. *Journal of Applied Pharmaceutical Science*. 01 (07):72-76.

Edeoga, H. O., Okwu, D. E. and Mbaebie, B.O. (2005). Phytochemical Constituent of Some Nigerian Medicinal Plants. *African Journal of Biotechnology*. 4(7): 685-688.

Garg, D., Muley A., Khare, N. and Marar, T. (2012). Comparative Analysis of Phytochemical Profile and Antioxidant Activity of Some Indian Culinary Herbs. *Research Journal of Pharmaceutical, Biological, Chemical Science*. 3(3): 546-554.

Jantas, D., Malarz, J., Le, T.N. and Stojakowska, A. (2021). Neuroprotective Properties of Kempferol Derivatives from Maesa membranacea against Oxidative Stress-Induced Cell Damage: An Association with Cathepsin D Inhibition and PI3K/Akt Activation. *Int. J. Mol. Sci.* 22:10363.

Vasantha Rupasingh, H.P. (2020). Special Issue "Flavonoids and Their Disease Prevention and Treatment Potential": Recent Advances and Future Perspectives. Molecules 25:4746; doi:10.3390/molecules25204746 <a href="https://www.mdpi.com/journal/molecules">www.mdpi.com/journal/molecules</a>

Abdulmumin Y., Mika'il, T. A., 1, Sarki, S. I., Jibril, I. A., Muhammad, I. U., Wudil A. M. and Alhassan A. J. (2020). Biochemical and *In-vivo* Antioxidant Activities of Ethyl Acetate Fractions of Boswellia Papyrifera (Del.) Stem Bark on CCl4-Induced Liver Damage in Wister Rats. *Malaysian Journal of Chemistry* 22 (1):8-18

Kancheva, V. D. (2009). Phenolic antioxidants – radical-scavenging and chain-breaking activity: A comparative study. *Eur. J. Lipid. Sci. Technol.* 111:1072-1089.

Mottaghipisheh, J. and Stuppner, H. A. (2021). Comprehensive Review on Chemotaxonomic and Phytochemical Aspects of Homoisoflavonoids, as Rare Flavonoid Derivatives. *Int. J. Mol. Sci.* 22:2735. https://doi.org/10.3390/ijms22052735

