

Isolation of Cellulolytic Fungi from Rice Husk

ABSTRACT

Aim: To isolate cellulose degrading fungi from rice husk

Study Design: The experiment was carried under aseptic condition in 3 replicates.

Place and Duration of Study: Department of Biological Sciences, Microbiology Programme, Clifford University, Ihie Campus, Owerrinta, Abia State, Nigeria, between May 2021 to August 2021.

Methodology: Rice husk from a rice mill was collected and kept until visible microbial growth was noticed. The organisms were isolated, characterized and screened for enzyme activities.

Results: Cellulolytic organisms were isolated from rice husk, an agricultural by-product of rice milling. The isolates were identified as *Penicillium* and *Aspergillus* sp. The two fungal isolates were screened for enzyme activity using 0.5ml Carboxymethyl cellulose (CMC) as carbon source, the highest enzyme activity of 0.448µg/ml/min was recorded for *Aspergillus* sp. at 48 hours while *Penicillium* sp. had enzyme activity of 0.388µg/ml/min at day 1.

Keywords: *Aspergillus niger*, Cellulase, Enzyme production, *Penicillium*, Rice husk.

1. INTRODUCTION

Rice husk, an agricultural by-product of rice milling, is abundantly available in Nigeria and has no direct nutritional value so it is left to rot or used as fuel. An attempt at feeding it to poultry resulted in poor growth performance as a result of low nutritional quality, though high in fiber and lignin content [1]

Cellulose is the most abundant biological compound on terrestrial and aquatic ecosystem. It is the dominant waste material from agricultural industry in the form of stalks, stems and husk. there has been great interest in utilizing cellulose as an energy resource and feed, the cellulose is composed of D-glucose units linked together to form linear chain via β-1, 4-glycosidic linkages [2]. Cellulose is one of the important additives to manufacture of bioplastics [3], food packaging materials [4], pharmaceutical, food, cosmetic and other industries [5]. Cellulose is a natural polymer having a linear structure, crystalline form and not easily to dissolve.

Carbohydrate materials (sugars, starch and cellulose) are valuable and natural industrial raw materials used worldwide [6]. A lot of useful products can be produced from the monomeric units of these carbohydrate materials. However, in order to convert starch and cellulose to useful products, they need to be hydrolyzed into their monomeric units by either enzymes or chemicals (acids or bases). Although chemical hydrolysis is presently faster and cheaper than enzymatic method, it is not environmentally friendly and requires special (non-corroding) vessels for the reaction to take place. Bioconversion using enzymes are safer and more environmentally friendly than the use of chemicals [7].

Plant biomass contains cellulose as the major component of the cell walls. Cellulose accounts for 50% of the dry weight of plant biomass and approximately 50% of the dry weight of secondary sources of biomass such as agricultural wastes; cellulose is a strong fibrous, crystalline polysaccharide, resistant to hydrolysis and is water insoluble [8].

Comment [MME1]: These keywords cannot be found in the abstract, what we have in the abstract are *Aspergillus* species (sp.) and cellulose. Any word not found in the abstract is not good for keyword and cannot be keyword.

Also, the words highlighted in yellow in the abstract need correction. sp. should be written in full at the first instance and then abbreviation subsequently. The correct abbreviation is spp. Furthermore, there should be space between number and unit except %. Moreover, the sp of *Penicillium* should be correctly written as spp. There is need for conclusion in this your abstract.

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Cellulolytic enzymes play an important role in natural biodegradation processes in which plant lignocellulosic materials are efficiently degraded by cellulolytic fungi, bacteria, actinomycetes and protozoa. In industry, these enzymes have found novel applications in the production of fermentable sugars and ethanol, organic acids, detergents and other chemicals. Cellulases provide a key opportunity for achieving tremendous benefits of biomass utilization. As important as this enzyme is, it is not readily available and expensive, therefore this study was designed to isolate organism that can secrete the enzyme as this will be a major breakthrough for the industries where the enzyme has found usefulness.

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2. MATERIALS AND METHODS

2.1. Collection of Samples

The Rice Husk was collected from a Rice milling industry in Abakiliki, Ebonyi state, Nigeria.

2.2. Isolation of the Microorganisms from Rice Husk

10 g of rice husk was weighed into 90 ml distilled water; 10 folds serial dilution was done. 0.1 ml each of 10^{-7} , 10^{-8} , 10^{-9} and 10^{-10} were plated out on Sabouraud dextrose Agar using pour plate method and was kept at room temperature for 7 days. Subculture of the growths were done on Sabouraud dextrose agar (SDA) and kept at room temperature for 7 days.

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2.3. Identification of the Fungal Isolates

A slide culture was prepared to identify the organism on the plate and after 7 days it was observed under the microscope. A small portion of the mycelia growth was carefully picked with the aid of a sterile inoculating needle and placed in a drop of lactophenol cotton blue on a microscope slide and covered with a cover slip. The slide was examined under the microscope, first with (x10) and then with (x40) objective lens for morphological examination. The isolates were further identified macroscopically using their cultural characteristics according to Gilma [9] and Barnett and Hunter [10]

2.4. Degradation Ability of Fungal Isolates on Carboxymethyl Cellulose Media (CMC)

Carboxymethyl cellulose (CMC) medium was prepared and the fungal isolates were inoculated on the CMC media for 7 days to check for the growth of the isolates.

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2.5. Screening for Cellulase Activity

The isolates were grown in broth containing 1% CMC as carbon source. They were incubated at room temperature for 72 h, after which enzyme assay was carried out. The cellulase activity was measured.

2.6. Cellulase Assay

The method used involved estimating the amount of reducing sugar produced by the activity of the enzyme on buffered 1 % CMC. The amount of reducing sugar produced was estimated using the dinitrosalicylic acid (DNSA) method by Miller [11]. The reaction mixture containing 0.5 ml of supernatant and 0.5 ml of 1 % CMC was incubated at 50 °C in a water bath for 30 mins. The reaction was terminated by adding 3 ml DNSA and then boiled for 10 mins in a boiling water bath. The control tubes contained the reaction mixture but lacked the crude enzyme solution. Absorbance was taken at 540 nm using a spectrophotometer [11]. The amount of reducing sugar produced was derived from a glucose concentration curve.

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One unit of cellulase was defined as the amount of enzyme which released 1 µg of glucose from cellulose per ml per min under the assay conditions.

3. RESULTS AND DISCUSSION

3.1. Isolation and Screening of Cellulose Degrading Fungi

Few species of fungi were isolated on Sabouraud dextrose Agar medium and only two species were able to degrade cellulose. These fungal isolates were identified by cultural (Appendix) and microscopic characteristics and were identified as *Aspergillus* sp. and *Penicillium* sp. The microscopic examinations are as shown in Figures 1 and 2.

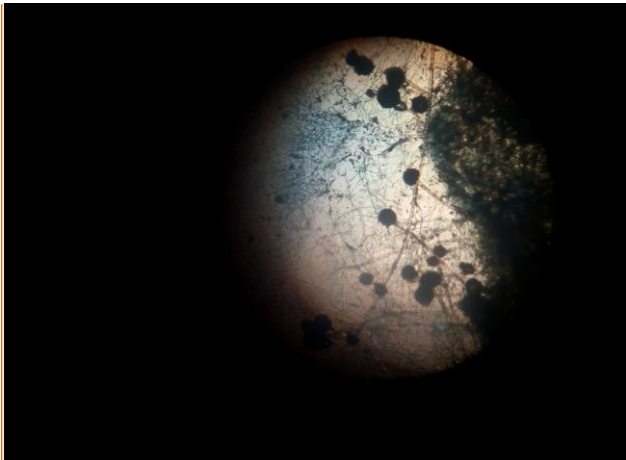


Figure 1: Microscopic View of *Aspergillus* Sp.

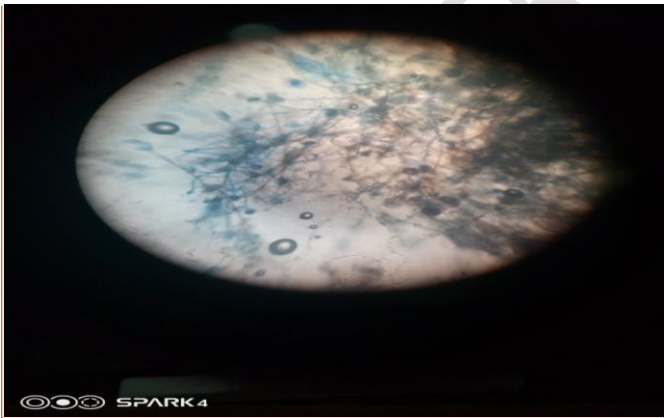


Figure 2: Microscopic view of *Penicillium* sp

Enzyme activities of *Aspergillus* and *Penicillium* are as shown in Figures 3 and 4.

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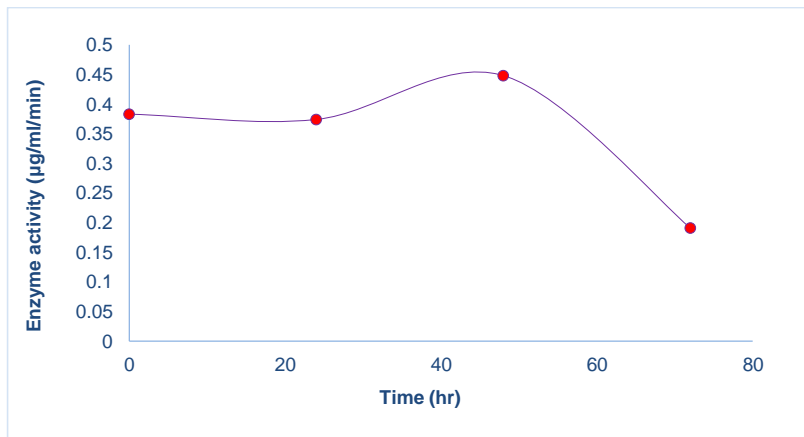


Figure 3: Enzyme Activity of *Aspergillus sp.* using 1 % CMC as carbon source

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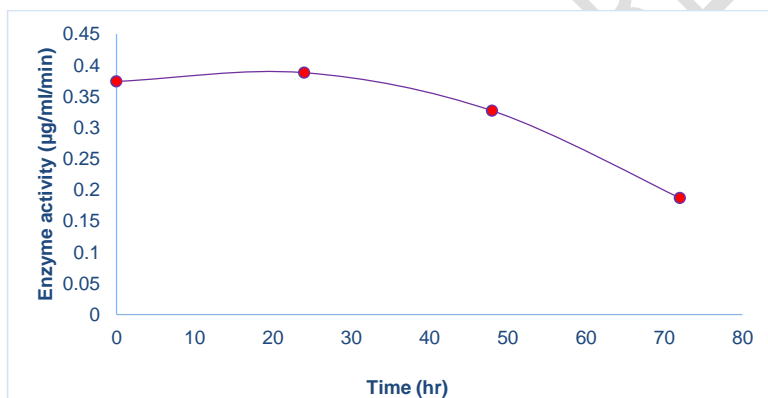


Figure 4: Enzyme Activity of *Penicillium Sp.* using 1 % CMC as carbon source

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Comment [MME23]: Please obey the rules guiding the writing of scientific/botanical names; both genus name and specific name cannot start with capital letter/uppercase.

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Comment [MME30]: This should be in the methodology aspect of this manuscript.

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The two species of fungi (*Aspergillus sp.* and *Penicillium sp.*) were able to produce cellulase to degrade the cellulose content of rice husk. The result of this study is in agreement with the result of Oyeleke *et al.*, [12], who isolated *Aspergillus niger* and other forms of bacteria from the gut of *Archachatina Marginata* (Giant African Snail), also Edor *et al.*, [1], conducted a study where *Aspergillus niger* synthesized cellulase which biodegrade the cellulose content of rice husk. The cellulase activity was measured by the release of reducing sugar over the period of biodegradation.

In this study, *Aspergillus sp.* has the highest enzyme activity at Day 2 (48 hrs), this agreed with work of Oyeleke *et al.*, [12] who reported the highest enzyme activity for cellulase from *Aspergillus niger* at Day 2.

The result of this study has demonstrated the isolation of cellulolytic fungi from rice husk, these organisms can be useful in so many industries thereby reducing the cost of production and also reducing organic waste in the environment.

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4. CONCLUSION

The result of this study shows that *Aspergillus* and *Penicillium* sp. can secrete the enzyme cellulase which is a very important enzyme in so many industries and very expensive. Thus isolating organism that can secrete this enzyme is an added advantage to industries like the Agricultural industries, food industries, textile industries, pulp and paper industries, fermentation industries.

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Comment [MME39]: I think the conclusion should start with statement that these fungi can be isolated from rice husk before stating that they secrete cellulase enzyme.

REFERENCE

1. Eder SP, Edogbanya OP, Kutshik JR. Cellulase activity of *Aspergillus niger* in the biodegradation of rice husk. *MOJ Biology and Medicine*. 2018;3(2): 49-51.
2. Abedin IJ. Isolation and identification of cellulose degrading Bacteria from the soil sample. Department of Mathematics and Natural Sciences. 2015.
3. Agustin MB, DeLeon ERP, Breunaobra JL, Alonzo SMM, Patriana MF, Hirose F, Ahmad B. Starch based bioplastics reinforced with cellulose nanocrystals from agricultural residues. *International Conference on Advance in Engineering and Technology*. 2014;593-597.
4. De Moura MR, Mattoso LHC, Zucolotto V. Development of cellulose- based bactericidal nanocomposites containing silver nanoparticles as their use as active food packaging. *Journal of food engineering*. 2012;109: 520-524.
5. Salehudia MH, Salleh E, Mamat SNH, Muhamad II. Starch based active packaging film reinforce with empty fruit bunch (EFB) Cellulose nanofibers. *Procedia Chemistry*. 2014: 9: 23-33.
6. Sudha PN, Aisverya S, Nithya R, Vijayalakshmi K. Industrial applications of Marine carbohydrates. *Advance Food Nutrition Resource*. 2014;73: 143-181
7. Ogbonna CN, Nnaji OB, Chioke OJ. Isolation of amylase and cellulase producing fungi from decaying tubers and optimization of their enzyme production in Solid and Submerged cultures. *International Journal of biotechnology and Food Science*. 2018;6 (1): 9-17.
8. Rawway M, Salah GA, Ahmed SB. Isolation and Identification of cellulose degrading bacteria from different sources at Assiut Governorate (Upper Egypt). *An International Journal of Ecology of Health and environment*. 2018;6(1): 15-24.
9. Gilman CJ. A manual of soil fungi. Lower State College Press: Univ Park 1982:216
10. Barnett HL, Hunter BB. Illustrated Genera of Imperfect Fungi. Burgess. Pub. Co., Minnesota, USA.1972:241
11. Miller GL. Use of Dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical Chemistry*. 1959;31: 426 - 428.
12. Oyeleke SB, Egwim EC, Oyewole OA, John EE. Production of Cellulase and protease from Microorganisms Isolated from Gut of *Archachatina Marginata* (Giant African Snail). *Science and Technology*. 2012;2(1): 15-20.

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APPENDIX



Plate 1 *Aspergillus* sp. On Sabourad Dextrose Agar media

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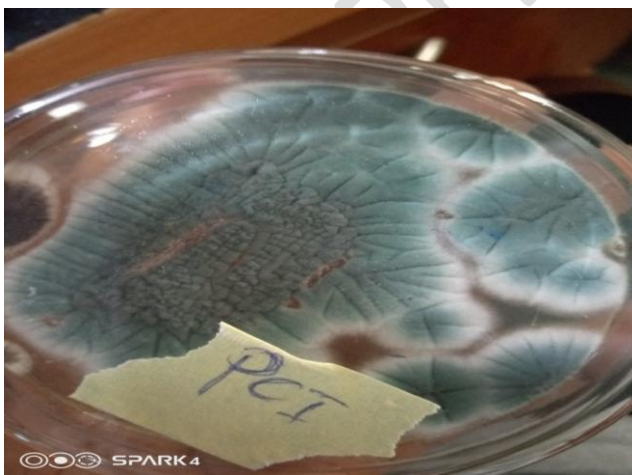


Plate 2 *Penicillium* sp. On Sabourad Dextrose Agar media

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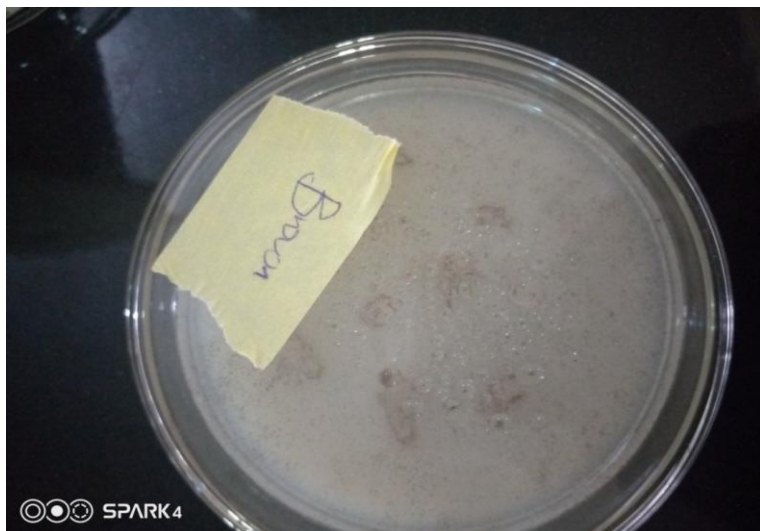


Plate 3 *Aspergillus* sp. On Carboxymethyl Cellulose Media

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Plate 4 *Penicillium* sp. On Carboxymethyl Cellulose media

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