# Nutritional Composition and Antioxidant Property of Methanol Extract of Corchorus olitorius Leaf

## **ABSTRACT**

**Aims**: This study investigates the phytochemicals, nutritional and antioxidant constituents of methanol extract of *C. olitorius* leaf using standard biochemical procedures.

**Methodology:** Corchorus olitorius (213.81 g) powdered leaves sample was soaked in 2.4 L of methanol respectively for 72 hr. Afterwards, the sample was filtered through a double-layered muslin cloth to obtain a filtrate which was placed in a rotary evaporator to dry off the solvent and stored. The proximate analysis, phytochemicals screening, mineral contents, antioxidant ability and phenolic compositions were determined for Corchorus olitorius.

Results:The proximate analysis revealed that the *Corchorus olitorius* extract contained 16.00 % ash, 9.60 % fat, 11.80 % moisture, 4.50 % crude fibre, 10.15 % crude protein and 48.80 % carbohydrate contents. Phytochemical screening indicated that flavonoids, tannins, cardiac glycosides, saponins, phenols and steroids were present in appreciable concentrations except for quinones and terpenoids. The mineral analysis of the extract showed considerable levels of potassium (1715.69 mg/100 g), calcium (33.43 mg/100 g), (sodium 49.62 mg/100 g), iron (16.78 mg/100 g) and manganese (9.44 mg/100 g) while magnesium (4.39 mg/100 g), copper (2.11 mg/100 g), zinc (2.94 mg/100 g) and lead (0.21 mg/100 g) were reduced. The extract showed high reducing power, diphenyl picrylhydrazine and  $H_2O_2$  radicals scavenging abilities. However, the total antioxidant capacity was low compared to the standard, ascorbic acid. High performance liquid chromatography result revealed that quercitrin, quercetin, chlorogenic acid, syringic acid, epicatechin and kaempferol were present in high amounts in the extract.

**Conclusion:** Altogether, findings from this study indicated that *C. olitorius* leaf extract is a rich source of phytonutrients and mineral elements with ample antioxidative property (*in vitro*) that may be of relevance in the management of some degenerative conditions.

**Keywords:** Corchorus olitorius, Phytochemicals, high performance liquid chromatography, in vitro antioxidant

## 1 INTRODUCTION

Regular consumption of vegetables has been linked to the management of several chronic conditions [1]. Currently, the emphasis is on foods that are high in dietary supplements or have positive health benefits [2]. The use of ingredients with increased doses of plant antioxidants, nourishing fibres, natural flavourings, mineral deposits, phytonutrients, and less man-made materials, amongst many other things, has caught the awareness of shoppers [3]. In general, fruits and vegetables are widely consumed in human diets across the globe in which plants serve as a great caloric and restorative ingredient [4].

The traditional green vegetable, *Corchorus olitorius* belongs to the Malvaceae family [5]. Many agriculturally important species can be found in the genus, *Corchorus*, rich in medicinal properties. *C. olitorius* is made up of 40 species and about 30 species are native to Africa [6]. *C. olitorius* is known for having a large distribution and a lot of trans-diversity [7]. The leaflets are parallel, oval, lance-shade, and ridged, causing diverse morphotypes to be recognized [8;9]. The presence of various macronutrients, beta-carotene and folic acid in *C. olitorius*, makes this vegetable a major food component [10].

The leaves of *C. olitorius* have been applied in ethnomedicine in the management of gonorrhea, chronic bladder infections, soreness, flu, and malignancies [11]. Furthermore, *C. olitorius* extracts

Comment [M1]: Not the correct term

Comment [M2]: Why deposites? delete

Comment [M3]: Provide a picture

could also be used to cure a variety of illnesses, including typhoid, anaemia and ulcers [9]. As a result of its numerous ethnomedicinal uses, this study sets out to evaluate the proximate content, phytochemicals screening (qualitatively and quantitatively) and *in vitro* antioxidant properties of methanol extract of *C. olitorius* leaf.

#### **2 MATERIALS AND METHODS**

#### 2.1 Collection of plants sample

Corchorus olitorius leaves were obtained from a vegetable farm in Ugbowo, Benin City, Edo State. The leaves were identified in the Department of Plant Biology and Biotechnology (Herbarium Unit), University of Benin, Nigeria. The voucher specimen (voucher number: UBH-C558) was immediately deposited at the Herbarium of the department. The leaves were rinsed properly and air-dried at room temperature for 7 days, then hand crushed into coarse powder and weighed.

#### 2.2 Preparation of Corchorus olitorius extract

Corchorus olitorius (213.81 g) powdered leaves sample was soaked in 2.4 L of methanol respectively for 72 hr. Afterwards, the sample was filtered through a double-layered muslin cloth to obtain a filtrate which was placed in a rotary evaporator to dry off the solvent. The concentrate was stored in an airtight container at 4 °C to protect against sunlight and moisture [12].

### 2.3 Proximate Analysis of Corchorus olitorius

The proximate composition (namely, crude protein, crude fibre, crude carbohydrate, moisture, crude fat and ash contents) of *Corchorus olitorius* was carried out according to the Association of Official Analytical Chemists [13] methods.

## 2.4 Mineral Analysis of Corchorus olitorius

Five grams (5 g) of dried powdered leaf sample was weighed into a porcelain dish and further dried at 105 °C for 3 hr in an oven. The dish with content was transferred to a muffle furnace and ignited for 6 hr at 500 °C until free from carbon (residue appears greyish-white). This was removed from the oven and the ash was moistened with a few drops of water (to expose bits of unashed carbon). The ash was re-dried in the oven at 100 °C for 3 hr and re-ashed in the furnace at 500 °C for 1 hr. The content was removed from the muffle furnace, and placed in a desiccator until it cooled. The ash was dissolved in 10 % nitric acid and filtered. The filtrate was further made up to 100 mL. The concentration of the mineral elements (including calcium, potassium, iron, lead, copper, magnesium, zinc, manganese and phosphorus) in *C. olitorius* leaf was analysed using an Atomic Adsorption Spectrophotometer (AAS) [13].

## 2.5 Preparation of stock solution

The stock solution was prepared by adding 0.1 g leaf extract of *C. olitorius* into a beaker containing 100 mL of ethanol.

# 2.6 Phytochemical Screening of Corchorus olitorius

The extract of *Corchorus olitorius* were screened for the presence of phytochemicals namely; flavonoids, tannins, alkaloids, phenols, cardiac glycosides, saponins, steroids, terpenoids and quinones according to the method described by [14].

#### 2.7 Reducing power of Corchorus olitorius

The reducing power of *Corchorus olitorius* extracts were determined according to the method described by [15;16] Exactly 800  $\mu$ L of *C. olitorius* extract was mixed with 400  $\mu$ L phosphate buffer (0.2 M, pH = 6.6) and 800  $\mu$ L of 1 % potassium ferricyanide [K<sub>3</sub>Fe(CN)<sub>6</sub>]; the resulting mixture was incubated at 50 °C for 20 min. Thereafter, 800  $\mu$ L (10 %) of trichloroacetic acid (TCA) was added to the mixture and centrifuged for 10 mins (3000 r/t). The resulting supernatant (400  $\mu$ L) was mixed with 400  $\mu$ L of distilled water and 80  $\mu$ L FeCl<sub>3</sub> (0.1 %) and the absorbance was recorded at 700 nm. Increased absorbance of the reaction mixture showed increased reducing power. The results were expressed as  $\mu$ g ascorbic acid equivalent/mg extract.

**Comment [M4]:** Of what? – fresh leaf or dry leaf or extraxt or concentrate? specify

Comment [M5]: Specify model and make, place

**Comment [M6]:** Incorrect Should be concentrate

Comment [M7]: was

Comment [M8]: name the method

Comment [M9]: against a reagent blank

**Comment [M10]:** rewrite as "the reducing power was reported in comparison with ascorbic acid.

## 2.8 Total antioxidant capacity of of Corchorus olitorius

The total antioxidant capacity of *Corchorus olitorius* was estimated by phosphomolybdenum assay of [17]. *C. olitorius* extract (1 mg/mL) was added to 3 mL of molybdate. The tube was incubated at 95 °C for 90 min. After incubation, the tubes were normalized to room temperature for 30 min and the absorbance of the reaction mixture was measured at 695 nm. Ascorbic acid was used as the standard antioxidant compound.

#### 2.9 Hydrogen peroxide scavenging ability of Corchorus olitorius

Hydrogen peroxide was measured based on procedures described by[18]. Briefly, hydrogen peroxide solution (2 mM) was prepared in 50 mM phosphate buffer (pH 7.4). Ascorbic acid was used as standard antioxidant compound. The abilities to scavenge the hydrogen peroxide were calculated using the following equation:

Hydrogen peroxide scavenging activity (%) =  $\frac{A_0 - A_1}{A_0}$  × 100

Where:

 $A_0$  =Absorbance of control  $A_1$ =Absorbance of sample

# 2.10 Diphenyl picrylhydrazyl (DPPH) of Corchorus olitorius

The scavenging abilities of *C. olitorius* extracts against 1,1–diphenyl–2–picrylhydrazyl (DPPH) radicals were estimated by a slightly modified method of [19]. Exactly 0.5 mL of 0.1 mM DPPH solution in methanol was added to 2 mL of different concentrations (0.2 - 1.0 mg/mL) of *C. olitorius* extracts. The tubes were shaken and incubated for 15 min at room temperature in the dark. The absorbance was read at 517 nm. All tests were performed in triplicate. Ascorbic acid was used as standard antioxidant compound. A blank containing 0.5 mL of 0.3 mM DPPH and 2 mL methanol was prepared and treated as the test samples. The radical scavenging ability was calculated using the formula below:

DPPH radical scavenging activity (%) =  $\frac{A_0 - A_1}{A_0}$  × 100

Where;

A<sub>0</sub> =Absorbance of control A<sub>1</sub>=Absorbance of sample

## 2.11 Phenolic composition of Corchorus olitorius

An aliquot (5 mL) of *Corchorus olitorius* extract was injected through a conditioned solid-phase extraction tube at 5 mL/min. The tubes were placed under vacuum (60 kPa) until the resin was thoroughly dried after which the phenolic compounds were eluted with 1 mL of ethyl vials. The PPL tubes were conditioned by first passing 2 mL of ethyl acetate followed by 2 mL of water (pH < 2.0). Purified phenolic extracts (1 mL: 10:1 split) were analyzed for composition by comparison with phenolic standards and chromatography with standards on a waters 600 high performance liquid chromatograph LCD system equipped with waters 515 HPLC pump, waters 2487 UV/VIS detector, C18 column with dimensions 5 mm, 4.6-250 mm with Hamilton microliter syringe, and injection volume of 20 mL. The following conditions were employed per separation: wavelength, 280 nm; flow rate, 1.0 mL/min; gradient elution total run time of 31 min, having solvent A as acetonitrile, solvent B as 0.1 % phosphoric acid in de ionized water, which was started with 85 % A and held at this for 13 min. This was followed by 75 % eluent B for 10 min and then the concentration of B was increased to 85 % for another 8 min.

# 2.12 Statistical Analysis

The results of this study were analysed using the Minitab version 17 package and Microsoft excel statistical package.

# 3.0 RESULTS AND DISCUSSION

Comment [M11]: Extract or concentrate

Comment [M12]: Name of the procedure

Comment [M13]: Why plural?

Comment [M14]: was

**Comment [M15]:** Picrylhydrazyl assay or radical scavenging activity

Comment [M16]: Method name?

Comment [M17]: Instrument name?

 $\label{lem:comment} \textbf{Comment [M18]:} \ \mathsf{Give \ details-name \ \& \ conc.}$ 

**Comment [M19]:** This is the name of the instrument? Use first letter capitals and place? Waters 600E, MA, USA?

**Comment [M20]:** How were the results obtained and reported?

#### 3.1 Proximate composition, Mineral contents, Phytochemical screening and Phenolic composition of Corchorus olitorius

The proximate composition of Corchorus olitorius revealed that ash, fat, moisture, crude fibre and carbohydrate contents were 15.00 %, 9.60 %, 11.80 %, 4.50 % and 59.80 % respectively. Protein was not detected in the sample (Table 1). The proximate composition result of Corchorus olitorius in this study revealed appreciable amount of ash, crude fat and crude carbohydrate contents. However, moisture and crude fibre were relatively low while protein was not available. The absence of protein in the leaf extract may be attributed to the method used to assay for the protein among other factors. Nevertheless, the proximate content of C. olitorius as observed in this study were relatively low when compared to previous work carried out on four tropical leafy vegetables by [20]. The moisture content of the leaf does not concur with the reports of [21]. The relatively low moisture content suggests that the C. olitorius leaf has a good shelf-life, improved processing characteristics and texture [22]. On the contrary, high moisture content may result to increased activity of water soluble enzymes that participate in the metabolic activities of leafy vegetables [22]. Ash is the inorganic residue left after the water and organic matter have been removed by burning the extract. In this study, C. olitorius recorded an appreciable amount of inorganic residue and mineral content. The ash content result asserts to the findings of [23]. The low fat content of C. olitorius in this study corroborated with the findings of previous studies which showed that leafy vegetables are poor sources of lipids [24]. Hence, it's important to note that food substance supplying 1 - 2 % of its caloric energy as fat is considered to be adequate to human beings, as excess intake of fatty food could conduce to cardiovascular disorders such as atherosclerosis, cancer and aging [25]. Crude fibre measures the cellulose, hemicellulose and lignin content in a food sample [26]. In this study, the crude fibre content of C. olitorius was present in minute amount. Dietary fibres are known to enhance bowel motility, prevent constipation and reduce the risk of colon cancer [26].

Table 2 shows the results of the mineral analysis of Corchorus olitorius leaf. Calcium (Ca), potassium (K), iron (Fe), sodium (Na), copper (Cu), magnesium (Mg), zinc (Zn), manganese (Mn) and lead (Pb) were present. Values were expressed in milligrams per 100 g (mg/100 g). The mineral analysis of C. olitorius extract revealed a high level of potassium. Furthermore, appreciable concentrations of sodium, calcium, and iron were also seen. However, copper, magnesium, zinc, manganese and lead were present in trace amounts in the leaf extract. The presence of high amount of certain minerals could be linked to the cultivar, different experimental analysis conditions and soil type [9]. Calcium is an essential component in the structure of bones and teeth. Calcium is basic for blood coagulating, upkeep of pulse and a cofactor in enzymatic procedures [9]. Potassium and sodium are important for the regular functioning of the sensory system and also circulatory systems [9]. Zinc, iron, magnesium and manganese constitute the basic components of the immune system and are essential for the build-up of haemoglobin [9].

The phytochemical screening of Corchorus olitorius extract shows that flavonoids, tannins, cardiac glycosides, saponins, phenols and steroids were present in the leaf extract while quinones and terpenoids were undetected (Table 3) and the phenolic compositions of Corchorus olitorius shown in (Table 4).

High-performance liquid chromatography of methanol extract of Corchorus olitorius leaf is presented in Figure 1 while the quantities of the phytochemicals present are shown in Table 4. The chromatograph (Figure 1) showed that the leaf extract had high levels of some secondary metabolites especially quercetin (34.35 mg/100 g), kaempferol (15.93 mg/100 g), epicatechin (14.01 mg/100g), chlorogenic acid (13.93 mg/100 g), syringic acid (12.32 mg/100 g) and quercitrin (11.19 mg/100 g). The remaining metabolites, though of biological importance, occurred in small quantities (Table 4). The phytochemicals, including flavonoids, tannins, cardiac glycosides, saponins, phenols and steroids, present in the leaf extract of C. olitorius as observed in this study corroborates with the reports of [27;28]. Flavonoids help to regulate cellular activity and scavenge free radicals that cause oxidative stress [29]. Also, flavonoids have been reported to lower the risk of emerging chronic diseases [29]. Phenols regulate enzyme activity and cell receptors. Studies have shown the protective function of polyphenols such as quercitrin, quercetin, catechin, epicatechin, kaempferol and sinapinic acid in the management of cardiovascular and neurodegenerative conditions [30; 31; 32; 33]. These and other phytochemicals were quantified via HPLC in the leaf extract of C. olitorius.

Table 1: Proximate Composition of Corchorus olitorius Leaf

**Proximate Contents** Corchorus olitorius (%)

The figures can be give next to the name of the component. Comment [M22]: Was absent? – not correct

Comment [M21]: The units must be specified -

May be negligible !!! The whole leaf is rich in protein – from studies

Comment [M23]: Mention value

Comment [M24]: Value?

Comment [M25]: Value? Why minute amounts?

Comment [M26]: Value?

Comment [M27]: Give the value for each

Comment [M28]: This study was on methanolic extract of the leaf - is it not? How are you giving data for leaf?

Comment [M29]: Give the correct unit of expression. Just % is not sufficient

Ash	16.00 ± 0.00	
Fat	$9.60 \pm 0.33$	
Moisture	11.80 ± 0.59	
Crude fibre	$4.50 \pm 0.05$	
Crude protein	$10.15 \pm 0.00$	
Carbohydrate	$48.80 \pm 0.69$	

All values were expressed as mean ± SEM

Table 2: Mineral analysis of Corchorus olitorius leaf extract

Minerals	Corchorus olitorius (mg/100 g)
Ca	33.43
K	1715.69
Fe	16.78
Pb	0.21
Na	49.62
Cu	2.11
Mg	4.39
Zn	2.94
Mn	9.44

All values were expressed in mg/100 g

Table 3: Phytochemical screening of Corchorus olitorius extract

Phytochemicals	Corchorus olitorius
Flavonoids	++
Tannins	+
Cardiac glycosides	++
Quinones	•
Saponins	+
Alkaloids	++
Phenols	++
Steroids	+
Terpenoids	-

+ Present in low concentration

++ Present in moderate concentration - Not detected.

Comment [M30]: G or mL

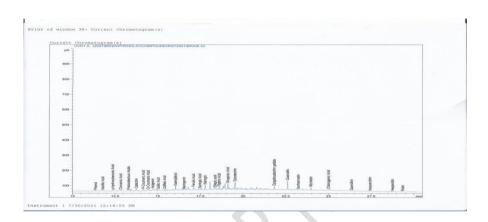


Figure 1: HPLC chromatogram of methanol extract of Corchorus olitorius

Table 4: Phenolic composition of Corchorus olitorius

Name	Amount (mg/100g)
Quercetin	34.34994
Myricetin	5.94739
Chlorogenic acid	13.92977
Quercitrin	11.19065
Catechin	7.06486

Comment [M32]: Leaf or extract?

Comment [M33]: 100g leaf or extract ?? mL??

**Comment [M34]:** Five digits after the decimal is not required – all figures in the table Two is sufficient

Gallic acid	1.56594
Caffeic acid	4.95559
Kaempferol	15.93058
Naringenin	4.90425
Ferulic acid	8.71090
Syringic acid	12.31687
Sinapinic acid	1.30345e- <sup>3</sup>
Epicatechin	14.00992
Epigallocatechin gallate	7.14122e- <sup>1</sup>
Isoquercitrin	3.45045e- <sup>1</sup>
Hesperidin	1.30183e- <sup>2</sup>

#### 3.2 In vitro Antioxidant potential of C. olitorius extract

Figure 2.1 to 2.4 represent the in vitro antioxidant potential of C. olitorius extract in terms of total antioxidant capacity, reducing power, 1, 1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging and hydrogen peroxide radical scavenging capacities. The results showed that the total antioxidant capacity of the extract was significantly (P = .05) low in contrast to the standard, ascorbic acid (Figure 2.1). The reducing power of the extract increased in a concentration dependent manner (P =.05) but was not as high as the standard antioxidant, ascorbic acid (Figure 2.2). In Figure 2.3, C. olitorius extract was able to significantly (P = .05) scavenge DPPH radicals in some cases at comparable levels as the standard ascorbic acid with IC50 for the extract and ascorbic acid being 1.19  $\mu$ g/mL and 1.10  $\mu$ g/mL, respectively. The ability for *C. olitorius* extract to scavenge  $H_2O_2$ radicals was high but not commensurate with that of the standard antioxidant, vitamin C (Figure 2.4). The antioxidant assays revealed that Corchorus olitorius leaf extract has appreciably high antioxidant potential especially in terms of its reducing power, total antioxidant capacity and hydrogen peroxide and DPPH radical scavenging abilities; though not as high as the standard antioxidant, ascorbic acid. The total antioxidant capacity assay is primarily based on the reduction of ferric ion to ferrous ion and molybdenum (VI) by the antioxidants in the samples, individually [28]. Thus, suggesting that Corchorus olitorius may possess the capacity of converting ferrous ion to their reduced form (ferric ion). The moderate reducing power of C. olitorius extract may translate to its ability to donate electrons in oxidative stress situation [34]. The relatively high hydrogen peroxide scavenging effect of C. olitorius extracts signifies reduction of such radicals as H<sub>2</sub>O<sub>2</sub> by making them less toxic. Therefore, the removal of hydrogen peroxide is critical for antioxidant defence in the cell [35].

## Total antioxidant capacity

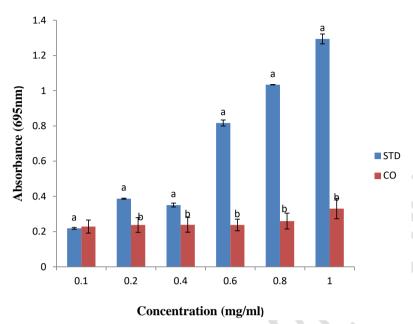
Comment [M35]: specify

Comment [M36]: what would you define as high or low?

Comment [M37]: ?

Comment [M38]: Define moderate

Comment [M39]: ??



**Figure 2.1** Total Antioxidant Capacity (TAC) analysis of *Corchorus olitorius* extracts Values were expressed as mean  $\pm$  SEM *where* n = 3; (P = .05) KEY: CO = *Corchorus olitorius*, STD = Standard

Reducing Power

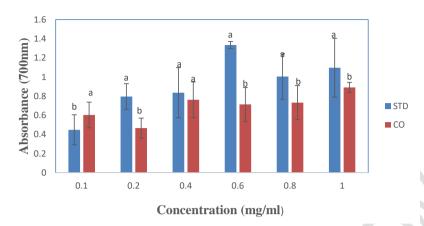
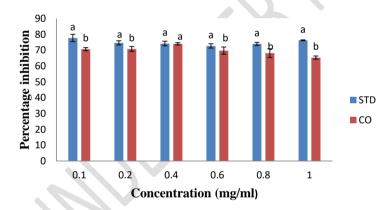


Figure 2.2 Reducing power of *Corchorus olitorius* extracts Values were expressed as mean  $\pm$  SEM where n = 3; (P = .05).

 $\mathsf{KEY} \colon \mathsf{CO} = \textit{Corchorus olitorius}, \, \mathsf{STD} = \mathsf{Standard}$ 

# **DPPH Radical Scavenging Ability**



**Figure 2.3:** DPPH radicals scavenging ability of *Corchorus olitorius* extracts Values were expressed as mean  $\pm$  SEM where n = 3; (P = .05). KEY: CO = *Corchorus olitorius*, STD = Standard

**Hydrogen Peroxide Radical Scavenging Ability** 

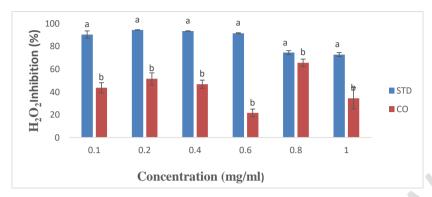


Figure 2.4: Hydrogen peroxide radical scavenging ability of Corchorus olitorius extracts

Values were expressed as mean  $\pm$  SEM where n = 3; (P = .05).

KEY: CO = Corchorus olitorius, STD = Standard

#### DPPH IC<sub>50</sub> value of Corchorus olitorius

Extract	Concentration (ug/mL)	
Ascorbic acid	1.10	
Corchorus olitorius	1.19	

In this study, the DPPH radical scavenging capacity of the extract was noticed to be almost at par with that of ascorbic acid standard suggesting that the extract may act more as a scavenger of free radicals generated and thereby inhibiting autoxidation of lipids in the cells [34].

# **4 CONCLUSIONS**

In conclusion, findings from this study reveal that methanol extract of *C. olitorius* leaf extract has appreciable nutritional values and could be considered as a rich source of antioxidants. The extract contained important secondary metabolites which may be of relevance in the management of some degenerative conditions.

## **REFERENCES**

- Handoussa H, Hanafi R, Eddiasty I, El-gendy M, El-Khatib A. Anti-inflammatory and cytotoxic activities of dietary phenolics isolated from *Corchorus olitorius* and *Vitis vinifera*. Journal of Functional Foods, 2013; 5(3): 1204-1216.
- Mulungulungu ND, Mpiana TP, Mbayo KM, Tshisand YP, Badibanga ML. Etude phytochimique de quelques légumes consommés dan le Haut-Katanga (RD Congo) et évaluation de leur activité antioxydante. International Journal of Innovation and Applied Studies,2015;10(1): 393-404.
- Lazou A, Kronida M. Functional properties of corn and corn-lenti extrudates. Food Research International, 2010; 43(2): 609-616.
- Dansi A, Adjatin A, Adoukonou-Sagbadja H, Faladé V, Yedomonhan H, Odou D, Dossou B. Traditional leafy vegetables and their use in the Benin Republic. Genetic Resources and Crop Evolution, 2008; 55: 1239-1256.
- Whitlock BA, Karol KG, Alverson WS. Chloroplast DNA sequences confirm the placement of the enigmatic *Oceano papaver* within *Corchorus* (Grewioideae: Malvaceal, formerly Tiliaceae). International Journal of Plant Sciences, 2003; 164(1): 35-41.

Comment [M40]: Specify extract

- Mbaye MS, Noba K, Sarr RS, Kane A, Sambou JM, Tidiane BA. Elements de précision sur la systématique d'espèces adventices dugenre *Corchorus* (Tiliaceae) au Sénégal. *African* Journal of Science and Technology,2001; 2(1): 51-64.
- 7. Benor S, Fuchs J, Blattner FR. Genome size variation in *Corchorus olitorius* (Malvaceae) and correlation with elevation and phenotypic traits. Genome, 2011, 54(7): 578-585.
- 8. Adebo HO, Ahoton LE, Quenum F, Ezin V. Agromorphological characterization of Corchorus olitorius cultivars of Benin. Annual Research and Review in Biology, 2015, 7: 229-240.
- 9. Adjatin A, Balogoun D, Loko L, Djengue W, Bonou-gbo Z, Yedomonhan H, Dansi A, Akoégninou A, Akpagana K. Phenotypic diversity, use and management of local varieties of *Corchorus olitorius* from central Benin. Journal of Biodiversity and Environmental Sciences, 2017; 11(1): 81-96.
- Choudhary SB, Sharma HK, Karmakar PG, Saha AR, Hazra P, Bikas SM, Anil-Kumar A. Nutritional profile of cultivated and wild jute (*Corchorus*) species. Australian Journal of Crop of Science, 2013; 7(13): 1973-1982.
- 11. Kumawat BK, Gupta M, Singh TY. Free radical scavenging effect of various extracts of leaves of *Balanites aegyptiaca* (L.) Delile by DPPH method. Asian Journal of Plant Science and Research, 2012; 2(3): 323-329.
- Sutharson L, Lila KN, Prasanna KK, Shila EB, Rajan VJ. Anti-inflammatory and antinociceptive activities of methanolic extract of the leaves of *Fraxinus floribunda* wallich. African Journal of Tradition, CAM, 2007; 4 (4): 411 – 416.
- 13. Association of Official Analytical Chemist AOAC. Official Methods of the Association of Official Chemists. Official Analytical Int, Arlington VA. change of Bacterial Community. Washington DC; 2000.
- Ogundipe A, Greenberg B, Braida W, Christodoulatos C, Dermatas D. Morphological characterization and spectroscopic studies of the corrosion behavior of tungsten heavy alloys. Corrosion Science, 2006, 48: 3281–3297.
- 15. Oyaizu M. Studies on product of browning reaction prepared from glucose amine. *Japan* Journal of Nutrition, 1989; 44: 307-315.
- 16. Hemalatha A, Girija K, Parthiban C, Saranya C, Anantharaman P. Antioxidant properties and total phenolic content of a marine diatom, *Navicula clavata* and green microalgae, *Chlorella marina* and *Dunaliella salina*. Advances in Applied Science Research, 2013, 4: 151-157.
- 17. Prieto P, Pineda M, Aguilar M.Spectrophotometric Quantitation of Antioxidant Capacity through the Formation of a Phosphomolybdenum Complex: Specific Application to the Determination of Vitamin E. Analytical B Biochemistry.1999,269: 337–341.
- Nabavi SM, Ebrahimzadeh MA, Nabavi SF, Fazelian M, Eslami B. In vitro antioxidant and free radical scavenging activity of *Diospyros lotus* and *Pyrus boissieriana* growing in Iran. Pharmacognosy Magazine. 2009, 5: 122-126.
- 19. Brand-Williams W, Cuvelier A, Berse M. Use of free radical methods to evaluate antioxidant activity. Food science and technology, 1995, 28(1):25-30.
- Obeng E, Kpodo FM, Tettey CO, Essuman EK, Adzinyo OA.Antioxidant total phenols and proximate constituents of four tropical leafy vegetables. Scientific African. 2020, 7: 2-2.
- 21. FAO. Food and Agriculture Organization. Annex 6. Requirements for fortification in food aid programmes. FAO Technical consultation on food fortification: Technology and Quality control Rome, Italy; 2006.
- Iheanacho K, Ubebani AC. Nutritional composition of some leafy vegetable consumed in Imo State, Nigeria. Journal of Applied Science and Environment Management, 2009;13: 35-38.
- 23. Morris A, Barnett A, Burrows O. Effect of processing on nutrient content of foods. Cajaericels, 2004, 37:130-164.
- Ejoh AR, Tchouanguep MF, Fokou E. Nutrient composition of the leaves and flowers of Colocasia esculenta and the fruits of Solanum melongena. Plant Food for Human Nutrition, 1996; 49: 107-112.
- Kris-Etherton PM, Hecker KD, Bonanome A, Coval SM, Binkoski AE, Hilpert KF, Griel AE, Etherton TD. Bioactive compounds in foods: their role in the prevention of cardiovascular disease and cancer. PubMed, 2000, 9: 71-88.
- Zakpaa HD, Mak-Mensah EE, Adubofour M. Production and characterization of flour produced from ripe 'apem' plantain (*Musa sapientum* L. var. *paradisiaca*; French horn) grown in Ghana. Journal of Agricultural Biotechnology and Sustainable Development, 2010; 2(6): 92-99.

**Comment [M41]:** Reference is incorrect The correct one is given below

Comment [M42]: Kris-Etherton, P.M., Hecker, K.D., Bonanome, A., Coval, S.M., Binkoski, A.E., Hilpert, K.F., Griel, A.E. and Etherton, T.D. (2002) Bioactive Compounds in Foods: Their Role in the Prevention of Cardiovascular Disease and Cancer. The American Journal of Medicine, 113, 71-88. http://dx.doi.org/10.1016/S0002-9343(01)00995-0

- Okunlola GO, Jimoh MA, Olatinji OA, Olowolaju ED. Comparative study of the phytochemical contents of *Corchorus olitorius* and *Amaranthus hybridus* at different stages of growth. Annales of West University of Timisoara. Series of Biology, 2017, 20 (1):43.
- Anthony OE, Ojeifo UP. Phytochemical screening and acute toxicity evaluation of *Telfairia* occidentalis aqueous extracts on rats. Pakistan Journal of Pharmaceutical Sciences, 2016; 29(3):13-19.
- 29. Jideani AO, Silungwe H, Takalani T, Omolola AO, Udeh HO, Anyasi TA. Antioxidant-rich natural fruit and vegetable products and human health. International Journal of Food Properties, 2021: 24(1): 41-67.
- Vauzour D, Rodriguez-Mateos A, Corona G, Oruna-Concha MJ, Spencer JP. Polyphenols and human health: prevention of disease and mechanism of action. Nutrients, 2010, 2:1106-1131
- 31. Cory H, Passarelli S, Szeto J, Tamez M, Mattei J. The role of polyphenols in human health and food systems: A mini-review. Frontier Nutrition, 2018,5:1-9.
- 32. Forni C, Facchiano F, Bartoli M, Pieretti S, Facchiano A, D'Arcangelo D, Norelli S, Valle G, Nisini R, Beninati S, Tabolacci C, Jadeja RN. Beneficial role of phytochemicals on oxidative stress and age-related diseases. BioMedical Research International 2019, 1-16.
- 33. Potì F, Santi D, Spaggiari G, Zimmetti F, Zanotti I. Polyphenol health effects on cardiovascular and neurodegenerative disorders: a review and meta-analysis. International Journal of Molecular Science, 2019; 20: 351.
- 34. Aliyu AB, Ibrahim MA, Musa MA, Musa AO, Kiplimo JJ, Oyewale AO. Free radical scavenging and total antioxidant capacity of root extracts of Anchomanes difformis Engl.(araceae). Acta Poloniae Pharmaceutical Drug Research, 2013, 70(1): 115-121.
- 35. Halliwell B. Reactive oxygen species in living systems: Source, biochemistry and role in human disease. American Journal of Medicine, 1991;91: 14-22.

Comment [M43]: Annals

Comment [M44]: 43-48