

## Original Research Article

### Appraisal the Effect of Nitrogen Phosphorus and Bio-fertilizers on Protein content in Seed of Garden Pea (*Pisum sativum* L.) cv. Arkel

#### Abstract

The field experiment was performed at Instructional Farm, College of Agriculture, Swami Keshwanand Rajasthan Agricultural University, Bikaner in the course of rabi year 2017-18. To study the effect of nitrogen, phosphorus and biofertilizers on the protein content of seeds of garden peas (*Pisum sativum* L.) cv. Arkel. The experiment was set up in RBD, with 11 treatments in three replications comparing nitrogen and phosphorus levels, as well as bio-fertilizers, to the control. A significant increase in protein content from 15.60% to 22.06% when  $20 \text{ kg ha}^{-1} \text{ N} + 40 \text{ kg ha}^{-1} \text{ P} + \text{Rhizobium} + \text{PSB}$  was applied ( $T_5$ ).

**Keywords:** Phosphorus, protein content, bio-fertilizers, pea, nitrogen

#### Introduction

Garden pea [*Pisum sativum* (L.)] is a popular vegetable crop farmed all over the world. In India, It is mostly grown as a cool-season crop in North India's plains and as a summer vegetable in the hills. It has chromosomal number  $2n=14$  and belongs to the leguminosae (Fabaceae) family. It is an annual herbaceous plant as dwarf as 20 cm and as tall as 1 to 1.25 meter or more. It is a self-pollinated crop, fruit which is eaten, known as pod. According to Choudhary, it is assumed to have originated in Ethiopia, a portion of Europe and Asia (1967)[4]. According to Thompson and Kelly, it is endemic to Europe and Western Asia (1957)[14]. Peas

are mostly used as a vegetable and as a pulse. It can be eaten fresh, canned, processed, or dehydrated (Thamburaj and Singh, 2005)[13].

In India, the pea occupies 540 thousand hectares and produces 5427 thousand tonnes of grain (Anonymous, 2017)[3]. It is grown on around 13831 hectares in Rajasthan, with an annual grain production of 36375 tonnes (Anonymous, 2016)[2]. It is primarily grown in Rajasthan's Jaipur, Baran, Bundi, Kota, and Bharatpur districts.

Peas are nutrient-dense, with a high proportion of digestible protein, as well as carbs and vitamins. It also has a lot of minerals. It has 7.2 percent protein, 19.8% carbs, and 0.8 percent mineral content in its pod. Dried pea grains, on the other hand, are high in vitamins A, B1, B2, and C and include 19.7% protein, 56.6 percent carbohydrate, 2.1 percent mineral matter, and 4.4 percent iron. 1967 (Choudhary)[4]. Its importance as a green manure crop has long been known because it is a nitrogen-fixing legume. As chemical fertilisers become less available and more expensive, people are becoming increasingly interested in pea as a soil-building crop. Pea is a plant that responds well to nitrogen fertiliser, especially when it is young. Nitrogen helps to encourage the growth of leaves, stems, and other vegetative growth. It also boosts the amount of protein in peas.

The pea is a prolific nodulating legume that requires little nitrogen. Nitrogen, on the other hand, plays an important function in a variety of plant metabolic activities. It is a component of proteins and chlorophyll, as well as nucleotides, phosphatides, alkaloids, enzymes, hormones, vitamins, and other chemicals important in plant metabolism. It gives plants a dark green colour, speeds up early growth, and boosts the capacity to fix atmospheric nitrogen symbiotically. Nitrogen administration at lower doses to legumes in the early stages is critical for a vigorous start.

Low yields are caused by growing pulses without using phosphatic fertiliser. Various researchers have reported that an appropriate supply of phosphorus is advantageous for legume growth and production, quality, and massive nodule development (Sammauria et al., 2009)[9]. It is an important structural component of cell membranes, chloroplasts, and mitochondria. It's found in energy phosphates like ADP and ATP, as well as nucleic acid, nucleoproteins, purines, pyrimidine, nucleotides, and a variety of co-enzymes. It is involved in the basic photosynthesis reaction. It is involved in cell division, carbohydrate breakdown for energy release, the transmission of hereditary traits, and the maturation of plants. Poor nodulation and low yield were noted in locations where legumes are

typically farmed without phosphorus. On all soil types, phosphorus shortage is probably the most important single reason for poor leguminous crop productivity. The added phosphorus is said to have a twofold effect, raising both the production of the legume and the yield of the following crop.

The usage of bio-fertilizers is critical for enhancing fertiliser efficiency. When pulse seedlings are inoculated with *Rhizobium* and planted in these soils. It boosts their population in the rhizosphere, increasing the quantity of microbiologically fixed nitrogen available for plant growth. Seed inoculation with the correct *Rhizobium* culture strain boosts seed yield above the uninoculated control. The combination of *Rhizobium* with pulse plants improves soil fertility and is a low-cost nitrogen fertiliser strategy for legumes.

Due to a lack of population and low activity of *Rhizobium* strains, inoculation with effective *Rhizobium* and P-solubilizing strains in the presence of N and P fertilisers may help to increase production. In recent years, eco-friendly and low-cost bio-fertilizers, both N-fixer and P-solubilizer, have emerged as a significant and integral component of crop production's integrated plant nutrients supply (IPNS) system.

Around 93-99 percent of total phosphorus is insoluble in soil and so unavailable to plants directly. During the application season, only about a quarter of the water soluble phosphate is taken up by plants, with the rest being transformed to insoluble (unavailable) forms (Verma, 1993)[15]. Phosphate solubilizing first appearance Solubilizing microorganisms in the rhizosphere of crops and soil increase the availability of phosphate from insoluble sources, desorption of fixed phosphates, and phosphatic fertiliser efficiency (Gaur, 1991)[5]. Phosphate solubilizing bacteria secrete acidic compounds that solubilize inaccessible soil phosphorus and make it available to plants after being inoculated. As a result, the culture can be used as a broad-spectrum bio-fertilizer, potentially increasing crop yields by 10-30% and supplementing phosphorus needs by up to 30 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>. PSB culture improves crop nodulation, growth, nutrient uptake, and yield (Shrivastava and Ahlawat, 1993)[10].

Increased nitrogen content in seed, possibly as a result of increased nitrogen availability to plants. Increased activity of the nitrate reductase enzyme could be another reason for the elevated nitrogen concentration. Because nitrogen is a fundamental component of amino acids, which form the basis of protein, increased nitrogen in seed is directly responsible for higher protein.

## **Materials and Methods**

The experimental study was conducted at the Instructional Farm, College of Agriculture, Swami Keshwanand Rajasthan Agricultural University, Bikaner, which is located 10 kilometres from Bikaner city on the Sri Ganganagar Road at an elevation of 234.7 metres above mean sea level, with a latitude of 28° 01' N and a longitude of 73° 22' E. According to the National Bureau of Soil Survey and Land Use Planning (NBSS & LUP), Bikaner is part of Agro ecological region No.2 (MgE1), which is characterised by deep, sandy and coarse loamy desert soils with low water holding capacity, a hot and arid climate, and annual precipitation of less than 300 mm. In this area, annual potential evapotranspiration (PET) ranges from 1500 to 2000 mm. Bikaner is located in the "Hyper Arid Partially Irrigated Western Agro-climatic Zone (IC)," according to NARP, Bikaner is part of the "Hyper Arid Partially Irrigated Western Plain Zone" Agro-climatic zone (IC). Bikaner is located in India's Agro-climatic Zone XIV (Western Dry Region), according to the National Planning Commission.

### **Weather and climate conditions**

Weather and climate conditions Weather characteristics play a vital part in determining the growth and development processes of crops, as well as the manifestations of various applied treatments, so it's crucial to present the climatic data collected during the experiment. The environment of this zone is often arid, with acidity in the atmosphere and salinity in the rhizosphere, harsh temperatures in both summer and winter, and temperature shifts marked by sudden drops and rises. Furthermore, during the summer and winter months, the mean maximum and minimum temperatures fluctuate widely. The region's yearly average rainfall is around 200-300 mm, with the most of it falling during the south-west monsoon season of July to September. The Meteorological Observatory at the Agricultural Research Station, Bikaner, recorded mean weekly data for temperature, relative humidity, rainfall, wet days, and mean evaporation.

### **Soil of experimental field**

To determine the physical and chemical parameters of soil, soil samples ranging from 0 to 30 cm in depth were collected at random from various locations around the experimental field, and a representative composite sample was generated. The

physico-chemical characteristics of the soil were determined using this composite sample.

### **Nitrogen content**

An electrical grinder was used to grind the dry seed samples taken during harvesting and winnowing. To eliminate the black colour, the samples were digested with sulphuric acid and hydrogen peroxide. The nitrogen concentration was calculated using a colorimetric approach that used Nessler's reagent to create colour (Snell and Snell, 1949)[11] and was given as a percentage.

### **Protein content in seed**

The protein content of dry seed was estimated by multiplying the nitrogen percentage in the seed by 6.25. (A.O.A.C., 1960)[1].

## **Result and Discussion**

### **Nitrogen content**

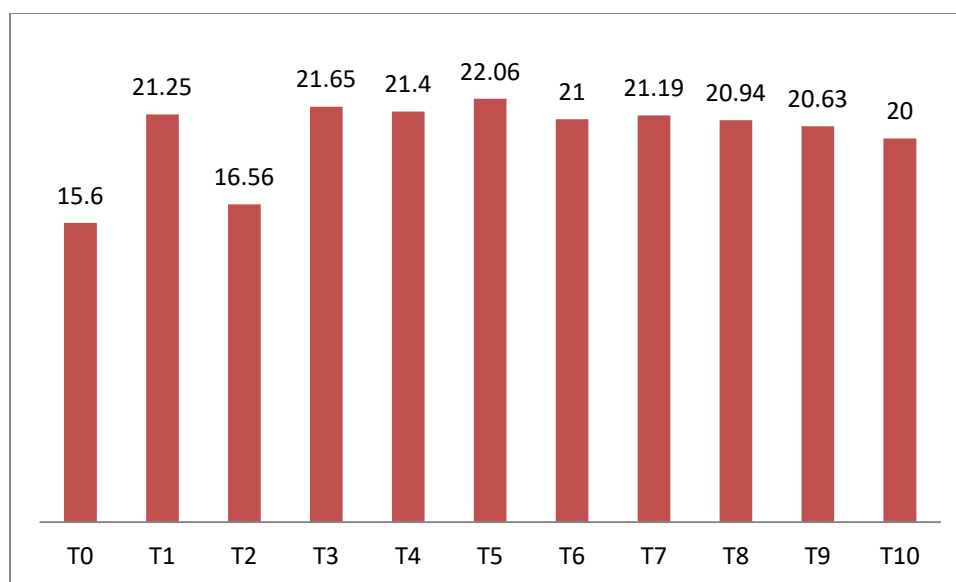
Table 2 shows that nitrogen content in plants varied greatly depending on nitrogen and phosphorus levels, as well as in conjunction with biofertilizers, as compared to the control. The method of treatment The maximal nitrogen concentration in plants was 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + Rhizobium + PSB (T<sub>5</sub>) (2.30 per cent). T<sub>5</sub> was statistically comparable to 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + Rhizobium (T<sub>3</sub>) and 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + PSB (T<sub>4</sub>), while the minimal nitrogen content in the plant (1.90%) was observed under control (T<sub>0</sub>) Table 2 also shows the nitrogen concentration of seeds. Because of the various treatments, the nitrogen content in seed differed dramatically. The maximal nitrogen concentration in seed was 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + Rhizobium + PSB (T<sub>5</sub>) (3.53 per cent). T<sub>5</sub> remained on par with 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P (T<sub>1</sub>), 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + Rhizobium (T<sub>3</sub>), 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + PSB (T<sub>4</sub>), 20 kg ha<sup>-1</sup> N + 20 kg ha<sup>-1</sup> P + PSB (T<sub>6</sub>), 20 kg ha<sup>-1</sup> N + 30 kg ha<sup>-1</sup> P + PSB (T<sub>7</sub>), 10 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + Rhizobium (T<sub>9</sub>). Under control, the minimal nitrogen content in seed was found to be 2.50. (T<sub>0</sub>).

## Protein content in seed

Protein content in seed was found to vary dramatically under different nutritional levels and in combination with biofertilizers, as shown in table 3. The method of treatment The greatest protein content in seed was found at 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + Rhizobium + PSB (T<sub>5</sub>) (22.06 per cent). T<sub>5</sub> was statistically equivalent to 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P (T<sub>1</sub>), 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + Rhizobium (T<sub>3</sub>), 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + PSB (T<sub>4</sub>), 20 kg ha<sup>-1</sup> N + 20 kg ha<sup>-1</sup> P + PSB (T<sub>6</sub>), 20 kg ha<sup>-1</sup> N + 30 kg ha<sup>-1</sup> P + PSB (T<sub>7</sub>), 10 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + Rhizobium (T<sub>0</sub>).

Due to enhanced phosphorus solubility and better nitrogen fixation in nodules, the combination inoculation of seeds with Rhizobium + PSB was more advantageous in improving all of the above characteristics, resulting in increased nitrogen and phosphorus availability. These findings back up those of Vimla and Natarajan (2000)[16] and Tanwar et al (2003)[12].

Because of increased nitrogen content in seed, which could be the result of enhanced nitrogen availability to plants, a significant rise in protein content from 15.60% in control to 22.06% with application of 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + Rhizobium + PSB (T<sub>5</sub>) was observed in the current study. Increased activity of the nitrate reductase enzyme could be another reason for the elevated nitrogen concentration. Because nitrogen is a fundamental component of amino acids, which form the basis of protein, increased nitrogen in seed is directly responsible for higher protein. Kasturikrishna and Ahlawat (2000)[7], Pandya and Bhatt (2007)[8], and Gupta et al. (2009)[6] all came to similar conclusions .



**Fig-1 Protein content (%) in seed**

## Conclusion

Based on the results of a one-year experiment, it can be concluded that 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + Rhizobium + PSB (T<sub>5</sub>) recorded highest protein content in seed (22.06%), and that this treatment was significant compared to 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + Rhizobium (T<sub>3</sub>), 20 kg ha<sup>-1</sup> N + 40 kg ha<sup>-1</sup> P + PSB (T<sub>4</sub>), 20 kg ha<sup>-1</sup> N + 20 kg ha<sup>-1</sup> P + PSB (T<sub>0</sub>). These findings are simply indicative, and more research is required to gain a more consistent and conclusive conclusion.

**Table 1. Details of the treatments with their symbols**

S.N.	Treatments	Symbols
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1.	Control	T <sub>0</sub>
2.	20 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P	T <sub>1</sub>
3.	<i>Rhizobium</i> + PSB	T <sub>2</sub>
4.	20 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P + <i>Rhizobium</i>	T <sub>3</sub>

	Treatments	Nitrogen content (%) in plant	Nitrogen content (%) in seed
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5.	20 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P + PSB	T <sub>4</sub>
6.	20 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P + <i>Rhizobium</i> + PSB	T <sub>5</sub>
7.	20 kg ha <sup>-1</sup> N + 20 kg ha <sup>-1</sup> P + PSB	T <sub>6</sub>
8.	20 kg ha <sup>-1</sup> N + 30 kg ha <sup>-1</sup> P + PSB	T <sub>7</sub>
9.	10 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P + <i>Rhizobium</i>	T <sub>8</sub>
10.	10 kg ha <sup>-1</sup> N + 30 kg ha <sup>-1</sup> P + <i>Rhizobium</i> + PSB	T <sub>9</sub>
11.	10 kg ha <sup>-1</sup> N + 20 kg ha <sup>-1</sup> P + <i>Rhizobium</i> + PSB	T <sub>10</sub>

Table 2. Effect of nitrogen, phosphorus and bio-fertilizers on nitrogen content in plant and seed of garden pea

T <sub>0</sub>	Control	1.90	2.50
T <sub>1</sub>	20 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P	2.15	3.40
T <sub>2</sub>	<i>Rhizobium</i> + PSB	1.91	2.65
T <sub>3</sub>	20 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P + <i>Rhizobium</i>	2.25	3.46
T <sub>4</sub>	20 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P + PSB	2.20	3.42
T <sub>5</sub>	20 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P + <i>Rhizobium</i> + PSB	2.30	3.53
T <sub>6</sub>	20 kg ha <sup>-1</sup> N + 20 kg ha <sup>-1</sup> P + PSB	2.16	3.36
T <sub>7</sub>	20 kg ha <sup>-1</sup> N + 30 kg ha <sup>-1</sup> P + PSB	2.18	3.39
T <sub>8</sub>	10 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P + <i>Rhizobium</i>	2.10	3.35
T <sub>9</sub>	10 kg ha <sup>-1</sup> N + 30 kg ha <sup>-1</sup> P + <i>Rhizobium</i> + PSB	2.00	3.30
T <sub>10</sub>	10 kg ha <sup>-1</sup> N + 20 kg ha <sup>-1</sup> P + <i>Rhizobium</i> + PSB	2.08	3.20
SEm±		0.04	0.08
CD at 5%		0.11	0.24

**Table 3. Effect of nitrogen, phosphorus and bio-fertilizers on protein content in seed of garden pea**

	<b>Treatments</b>	<b>Protein content (%)</b>
T <sub>0</sub>	Control	15.60
T <sub>1</sub>	20 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P	21.25
T <sub>2</sub>	<i>Rhizobium</i> + PSB	16.56
T <sub>3</sub>	20 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P + <i>Rhizobium</i>	21.65
T <sub>4</sub>	20 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P + PSB	21.40
T <sub>5</sub>	20 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P + <i>Rhizobium</i> + PSB	22.06
T <sub>6</sub>	20 kg ha <sup>-1</sup> N + 20 kg ha <sup>-1</sup> P + PSB	21.00
T <sub>7</sub>	20 kg ha <sup>-1</sup> N + 30 kg ha <sup>-1</sup> P + PSB	21.19
T <sub>8</sub>	10 kg ha <sup>-1</sup> N + 40 kg ha <sup>-1</sup> P + <i>Rhizobium</i>	20.94
T <sub>9</sub>	10 kg ha <sup>-1</sup> N + 30 kg ha <sup>-1</sup> P + <i>Rhizobium</i> + PSB	20.63
T <sub>10</sub>	10 kg ha <sup>-1</sup> N + 20 kg ha <sup>-1</sup> P + <i>Rhizobium</i> + PSB	20.00

SEm <sub>±</sub>	0.51
CD at 5%	1.51

## References

1. A.O.A.C. 1960. Official Method of Analysis, 18<sup>th</sup> Edn. Association of Official Agricultural Chemists, 1608. Broadnon Drive, Champaign, Illinois, USA.
2. Anonymous, 2016. Directorate of Horticulture, room no. 361B, Third floor, Pant Krishi Bhawan, Jaipur.
3. Anonymous 2017. Indian Horticulture Database, National Horticulture Board, Gurgaon
4. Choudhary, B. 1967. Vegetable (1<sup>st</sup> Ed.) National Book Trust, India. pp. 113.
5. Gaur, A.C. 1991. Phosphate solubilizing micro-organism and bio-fertilizer. Omega Scientific Publishers, New Delhi, pp. 176.
6. Gupta, S.C., Singh, R.P. and Verma, R. 2009. Response of chickpea to P levels from different sources with various PSB species. *Legume Research*, **32** (3) : 212-214.
7. Kasturikrishna, S. and Ahlawat, I.P.S. 2000. Effect of moisture stress and phosphorus, sulphur and zinc fertilizers on growth and development of pea (*Pisum sativum* L.). *Indian Journal of Agronomy*, **45** (2) : 353-356.
8. Pandya, C.B. and Bhatt, V.R. 2007. Effect of different nutrient levels on yield and nutrient content of fodder cowpea. *Legume Research*, **30** (3) : 218-220.
9. Sammauria, R., Yadav, R.S. and Nagar, K.C. 2009. Performance of clusterbean (*Cyamopsis tetragonoloba*) as influenced by nitrogen and phosphorus fertilization and biofertilizers in Western Rajasthan. *Indian Journal of Agronomy*, **54** (3) : 319-323.
10. Shrivastava, T.K. and Ahlawat, I.P.S. 1993. Response of pea (*Pisum sativum* L.) to phosphours, molybdenum and bio-fertilizers. *Indian Journal of Agronomy*, **40** : 630-635.
11. Snell, P.D. and Snell, G.T. 1949. *Colorimetric methods of analysis*, 3<sup>rd</sup> Edn. II D Van Nostrand Co. Inc., New York.

12. Tanwar, S.P.S., Sharma, G.L. and Chahar, M.S. 2003. Effect of phosphorus and biofertilizer on yield, nutrient concentration and uptake by blackgram (*Vigna mungo* L. Hepper). *Legume Research*, **26** (1) : 39-41.
13. Thamburaj, S. and singh, N. 2005. Vegetables Tuber Crops and Spices (3<sup>rd</sup> Edition), ICAR, Publication, New Delhi, pp. 196-221.
14. Thompson, H.C. and Kelly, W.C. 1957. Vegetable Crops. Publs. McGraw-Hill Book Company (5<sup>th</sup> Ed.), New York. pp: 460.
15. Verma, L.N. 1993. Organics in soil health and crop production, Ed. (Thampan, P.K.) *Tree crop Development foundation*, cochin : 151-184.
16. Vimla, B. and Natarajan, S. 2000. Effect of nitrogen, phosphorus and bio-fertilizers on pod characters, yield and quality in pea (*Pisum sativum* L. spp. hortense). *South Indian Horticulture*, **48** : 60-63.