Effects of Copper and Zinc Doses on Haematological, Renal and Liver Indices in Wistar

Rats

ABSTRACT

Copper and zinc are essential elements for various physiological and biochemical functions. Nevertheless, exposure to these heavy metals could also be detrimental to some vital organs such as the liver and kidney of the human body. There has been an increased use of agrochemicals on plants in the agriculture sector in many African countries, with the potential of bioaccumulation of these trace elements in both humans and animals as they feed on these plants. The present study, therefore, investigated the effects of these trace elements on haematological, renal, and liver indices in Wistar rats upon exposure.

A total of twenty-five two-weeks old Wistar rats (185 g - 250 g) were recruited for the study. Animals were grouped into five and administered feeds containing low and high doses of Copper and Zinc two weeks post-acclimatization. Laboratory investigations on haematological, renal, and liver indices were assessed after three weeks of feeding.

Both copper and Zinc doses significantly (P <0.05) elevated the counts for RBC, Hb, HCT, WBC, and Platelets with no significant effect on their weight gain compared to the control group of rats. Zinc doses elevated the levels of Alanine transaminase (ALT), with a low dosage having a significant effect on AST and ALP. The effect of copper and Zinc doses significantly (P <0.05) reduced creatinine levels, with no significant effect on urea concentrations.

The elevation of ALT and Platelets as a result of Copper and Zinc exposure suggests their deteriorating effect on the liver and other organs of the body. Long-term exposure to these trace elements can lead to a lot of pathologies.

Keywords: Copper, Zinc, haematological, renal, liver indices

1. INTRODUCTION

Copper and Zinc play functional roles in diverse physiological and biochemical activities in the body (Engwa et al., 2019; Shukla et al., 2019). Their effects on the improvement of enzymatic activities, protein synthesis, and stabilization of DNA and RNA have made them ideal for consumption (Dehkhodaei et al., 2018; Nishito and Kambe, 2018; Tabelin et al., 2018). Nonetheless, extreme exposure could be detrimental to vital organs of the human body (Fang et al., 2019; Tabelin et al., 2018). Long-term exposure to these metals has been associated with the slow progression of physical, muscular, and neurological degenerative processes that mimic

diseases like Parkinson's disease and Alzheimer's disease (Eom et al., 2020; Satarug et al., 2018).

Anthropogenic activities such as mining, smelting, and the use of agrochemicals in farming have immensely contributed to the release of these metals into the environment (Lü et al., 2018; Semu et al., 2019). Although chemical fertilizers, pesticides, and weedicides increase plant yield, their application has been associated with the accumulation of trace elements such as Copper and Zinc in the leaves, stems, and roots of crops (Garg and Payasi, 2020; Sundari et al., 2019). Consumption of leafy vegetables may therefore expose humans to these heavy metals (Chaoua et al., 2019; El-Kady et al., 2018).

Available reports suggest increased human exposure to heavy metals, a result of rapid growth and applications in agriculture, industrial, domestic, and technology (Proshad et al., 2018). While there have been studies on heavy metal exposure, these have largely been limited to assessing their levels and concentrations in environmental matrices (soil, water, and vegetables) (Engwa et al., 2019; Rai et al., 2019; Sandeep et al., 2019; Masindi and Muedi, 2018). The few available reports however evaluated the concentration of heavy metals in the animal rearing system and the administration of Copper Sulphate on markers of renal functions and feeding patterns of Wistar rats (Staniek, 2019; Ashajyothi and Handral, 2018).

Recent studies point to increasing concentrations of heavy metals in both medicinal plants and herbal distillates, leading to the production of reactive oxygen species upon human consumption and exposure (Fu and Xi, 2020; Kohzadi et al., 2019; Tamakhina et al., 2018). Based on these reports, we suspect that exposure to heavy metals (Copper and Zinc) may result in

pathophysiological effects in humans. Therefore, we qualitatively assessed the effects of these heavy metals on the haematological, renal, and liver indices of Wistar rats.

2. MATERIALS AND METHODS

2.1. Study Design

Twenty-five (25) male and female Wistar rats (185g – 250g) aged two weeks old were randomly selected for the experiment. These rats were obtained from the Pharmacological farm of Kwame Nkrumah University of Science and Technology (KNUST), Ghana.

2.2. Study Site

Animal housing and treatment were carried out at the laboratory of the Department of Science Education, University of Education, Winneba, Asante Mampong Campus. Hematological, Renal, and Liver tests were done at EDEP Laboratory service, Santasi –Kumasi, Ghana.

2.3. Maintenance of animals

Rats were housed in a one-tier hutch during the study. They were kept in animal rooms at the laboratory where room temperature was maintained at 26±3°C under a 12-hour light-dark cycle.

The rats were thoroughly examined and any abnormalities and/or ailments were recorded before the research began. Two hundred and twenty-five grams (225 g) of concentrate feed was weighed with a Camry dial spring scale (ISO9001 APPROVED) every morning and given to the animals as their daily feed. Feeding and water troughs were cleaned every morning before feeding. All animal experiments, procedures, and techniques used in this study were conducted in compliance with the National Institute of Health Guidelines for Care and Use of Laboratory Animals, with ethical approval from the Institutional Review Board, University of Cape Coast, Ghana (ethical clearance number: UCC IRB/CHAS/2017).

2.4. Preparation of feed

After two weeks of acclimatization, experimental rats were purely subjected to concentrate feed. The feed was formulated and prepared at the University of Education, Winneba, Mampong campus (UEW-M) laboratory. Ingredients were made up of, wheat brand (5 kg/30 kg); maize (15 kg/30 kg); concentrate (10 kg/30 kg); Copper Sulphate and Zinc tablet. They were weighed according to their required quantities with the Camry dial spring scale (ISO9001 APPROVED). The measured quantities were thoroughly mixed by spade, bagged, and then stored in the animals' feed room.

2.5. Treatments and preparations

A total of 25 experimental rats were used to assess the effect of Copper and Zinc on the haematological, renal, and liver indices. Animals were divided into five (5) groups and housed under controlled environmental conditions.

Copper sulphate (CuSO₄) and Zinc tablet used for the study were obtained from UEW-M laboratory and Laugh Pharmacy (Mampong-Ashanti) respectively. Five treatments were used for the study which included, treatment 1 (wheat brown maize concentrate), treatment 2 (wheat brand maize concentrate with low zinc conc.), treatment 3 (wheat brand maize concentrate with high zinc conc.), treatment 4 (wheat brand maize concentrate with high CuSO₄ conc.) and treatment 5 (wheat brand maize concentrate with low CuSO₄ conc.).

In the preparation of the wheat brand maize concentrate with CuSO₄ treatment, an amount of 3.929 g and 0.982 g of CuSO₄ powder was weighed for the high and low concentrations respectively using an electronic scale. It was then dissolved in 200ml distilled water and was uniformly mixed with 5 kg of the feed concentrate. Also, for the zinc treatment, 200 g and 50 g

of zinc tablets were dissolved in 200 ml distilled water for the high and low doses respectively. It was uniformly mixed with 5 kg of the feed concentrate. The mixture was bagged and stored in the animals' feed room.

Each treatment was administered as a feed to each group three times a day throughout the experimental period. Group 1 (control) was given treatment 1, Group 2 (treatment 2), Group 3 (treatment 3), Group 4 (treatment 4) and Group 5 (treatment 5). Each group was given an equal amount of feed throughout the experimental period. Weekly measurements of weight were recorded.

2.6. Haematological and biochemical parameters

Experimental rats were sacrificed and blood was dispensed into EDTA tubes and then analyzed for haematological indices such as red blood cells (RBCs), haemoglobin (Hb), hematocrit (HCT), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH), mean corpuscular haemoglobin concentration (MCHC) using an automated haematology analyzer (MSLAB 12, China). Another portion of blood was collected in empty vacutainer tubes and centrifuged at 13000 rpm for 5 mins. The serum was then retrieved and biochemical parameters such as AST, ALT, ALP, TB, TP, creatinine, and urea concentrations in serum were analyzed using commercially available kits and a standard BS-120 Mindray Chemistry Analyzer.

2.7. Statistical Analysis

GraphPad Prism Version 5.0 for Windows (GraphPad Software, San Diego, CA, USA) was used for data analysis. Data were presented as mean \pm SEM online graphs and tables. For comparing the control and treatment groups, independent samples t-tests for the significance of differences were used. To compare the biological effects of the treatment, an analysis of variance (ANOVA)

was used. Newman Keuls Posthoc analysis was also used to identify the source of the variation. p values of less than 0.05 were considered statistically significant.

3. RESULTS

3.1 Effect of Copper and Zinc doses on the weight of Wistar rats

Treated groups showed a steady weight gain after two weeks of acclimatization, although it was not statistically significant (P < 0.05) from the control group (figure 2).

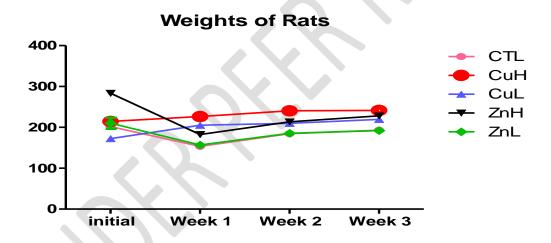


Figure 1. Effect of CuH (3.929g), CuL (0.982g), ZnH (200g), ZnL (50g) on weight.

3.2. Effect of Copper and Zinc doses on haematological parameters

The effects of copper and zinc dosages on haematological parameters were investigated and compared with similar parameters of normal control rats. Following the feed administration with

variously defined dosages, haematological parameters were significantly increased (P < 0.05) compared to the control group.

Table 1. Effect of **CuH** (3.929g), **CuL** (0.982g), **ZnH** (200g), and **ZnL** (50g) on levels of haematological indices.

	RBC	Hb	НСТ	MCV	МСН	МСНС	MPV	PLATELETS	WBC
CT							9.06±0.		
L	5.6 ± 0.1	10.6±0.1	35.00±1.0	59.1±0.7	19.32±0.3	31.78±0.3	4	26.18±2.9	2.36±0.2
Cu	8.3±0.2••	13.98±0.3•	51.46±1.3•	62.96±1.	17.12±0.3•			897.6±111.3•	
Н	•	••	••	2	•	27.2±0.5•••	9.8±0.2	•	8.82±0.2•••
Cu	8.1±0.2••	14.04±0.3•	50.52±1.5•		17.34±0.3•		9.42±0.	779.2±67.2••	
L	•	••	••	62.4±1.6		27.9±0.2•••	3	•	5.69±0.7••
Zn	9.0±	15.22±0.6•		60.78±0.	16.88±0.4•	27.64±0.4•	9.20±0.	805.3±53.6••	13.25±1.1•
Н	0.5•••	••	56.3±2.2•••	8			2	•	••
Zn	7.4±0.2••	13.06±0.2•	44.82±1.6•	60.64±1.	17.70±0.4•		8.70±0.	767.8±45.9••	11.71±0.6•
L	•	••		1	•	29.2±0.9••	3	•	••

Each column represents the Mean±SEM. (n= 5). •p <0.05, ••p <0.001, •••p <0.0001 compared to control group (One way ANOVA Followed by Newman-Keuls test)

3.3. Effect of Copper and Zinc doses on liver enzymes

Zinc low doses significantly (P <0.05) elevated the levels of the liver enzymes (AST, ALT, and ALP) whereas Zinc high doses only showed a significant (P <0.001), increase in ALT compared to the control. Comparing the different doses of copper, though not significant, the elevation of the biochemical indices was higher in CuL than in CuH as compared to the control group (table 2).

Table 2. Effect of **CuH** (3.929g), **CuL** (0.982g), **ZnH** (200g), **ZnL** (50g) on levels of enzymes and proteins associated with the liver (ALT, AST, ALP, GLB, ALB, TP).

	ALT	AST	ALP	GLOBULIN	ALBUMIN	TOTAL PROTEIN
CTL	100.5± 1.99	172.4±25.7	235.9±20.3	50.2±1.8	28.4±1.7	5.5±0.1
CuH	79.6±4.56	195.0±21.7	339.5±72.9	56.24±1.3	28.8±1.4	5.8±0.3
CuL	102.6±7.6	235.8±9.4	388.5±52.4	48.74±1.2	29.2±0.4	6.2±0.2
ZnH	157.6±26.9••	227.1±12.6	299.1±55.8	57.94±6.6	28.12±0.6	4.4±0.6
ZnL	147.3±12.3•	259.8±25.1•	783.2±201•	63.8±6.2	27.36±1.8	5.6±0.7

Each column represents the Mean±SEM. (n= 5). •p <0.05, ••p <0.001 compared to control group (One way ANOVA Followed by Newman-Keuls test)

3.4. Effect of Copper and Zinc doses on renal function

Doses of all treatment groups showed a significant (P <0.05) reduction of creatinine levels compared to the control. Zinc high doses showed an increase in urea levels, however, urea levels in copper doses (low and high) and zinc low doses were statistically not significant (figure 2).

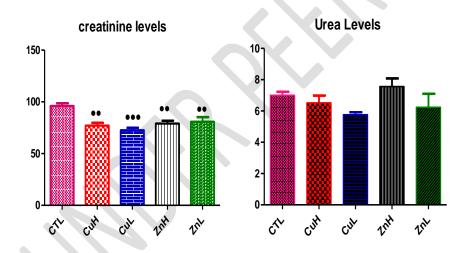


Figure 2. Effect of **CuH** (3.929g), **CuL** (0.982g), **ZnH** (200g), **ZnL** (50g) on creatinine and urea levels. Each column represents the Mean±SEM. (n= 5). •p <0.05, •p <0.001 compared to control group (One way ANOVA Followed by Newman-Keuls test).

4. DISCUSSION

The present study investigated the effects of Copper and Zinc on haematological, renal, and liver indices in Wistar rats. The effects of these trace elements administered as feeds significantly (P <0.05) elevated the haematological indices of the treated rats compared to the control. This agrees with Yahya et al., (2019) who reported significant (P <0.05) changes in haematological parameters, especially white blood cells (WBC) following the administration of copper oxide and/or zinc oxide nano-particles on rats. Also, a study by Zamaneh (2018) showed a significant (P <0.05) increase in red blood cells (RBCs), haemoglobin (Hb), and hematocrit (HCT) following zinc supplements.

The significant elevation of the haematological parameters such as RBC, Hb, WBC, and platelets counts, as shown in Table 1 following the supplementation of these trace elements may be a result of bone marrow deficiency leading to left shift production of these indices (Willis et al., 2005). Also, high platelets count suggests reactive thrombocythemia as a result of bone marrow suppression. This is in line with studies that suggest that persistent hyperglycemia predisposes individuals to thrombogenesis with platelet aggregation as a result of acute zinc supplements thereby prolonging bleeding time (Dimitrova et al., 2010; Hughes and Samman, 2006).

The high elevation of RBC and Hb following Copper sulphate exposure suggest vascular oxidative stress which can lead to impaired oxygen delivery hence the increment in its count for homeostatic balance. This is supported by findings from Lucas and Rifkinds (2013) who reported a pronounced increase in RBC and Hb as a result of vascular oxidative stress following copper bound amyloid- β peptide (CuA β) exposure.

The observed significant difference between treatment and control groups in some of the enzymes associated with liver function (ALT) following zinc doses (high and low)

administration suggests organ damage, especially in the liver. This agrees with Schultheiss et al., 2002 who reported on high liver zinc leading to histologic inflammatory lesions in dogs.

Serum alanine transaminases (ALT) are not only measured to determine possible liver damage but also useful in monitoring the general health status (Vazquez et al., 2020; Joni et al., 2020). This is an enzyme that is mainly found in the cytosol of the hepatocytes and plays a role in gluconeogenesis (Moriles and Azer 2020). ALT is released as a result of liver injury from the injured liver cells into the serum, which causes the levels to increase significantly in the serum (Ghosh et al., 2020; Smith et al., 2020). Also, only low doses of zinc treatment significantly (p <0.05) affected most of the liver enzymes (ALT, AST, ALP) indicating the essential role of zinc and hence it's right proportional supply to the body. Hepatic damage following Zinc deficiency sides with other studies that report on zinc deficiency as a predisposing factor for acute and chronic liver disease (Himoto and Massaki, 2018; Gruengreiff et al., 2016). Zinc supplementations have therefore been found to protect against toxin-induced liver damage and recommended as a therapy for hepatic encephalopathy in patients' refractory to standard treatment (Kamath and Leise, 2018; Elwir and Rahimi, 2017).

Copper doses, both high and low did not have any significant effect on the enzymes associated with liver function. This can be correlated to the significant reduction of creatinine levels following copper treatment. The observable insignificant effect of copper doses on renal function may be due to the body's homeostatic mechanism of regulating excess copper via bile secretion by the liver (Linder, 2020; To and Schilsky, 2018).

Urea is a principal Nitrogenous waste product of metabolism as a result of protein deamination (Higgins, 2016), its physiological role in the detoxification of ammonia and conservation of water in the kidney has made it ideal as a biomarker for renal function (Wang et al., 2019). Even

though statistically, no significant differences were observed in the treatment groups compared to control regarding blood urea levels, its slight increment following high doses of zinc treatment cannot be overlooked. The insignificant differences observed may be because blood urea levels are only affected following an extreme reduction of the Glomerular filtration rate (Elamin et al., 2017).

It was observed that the total protein, Gamma-glutamyltransferase (GGT), indirect bilirubin and total bilirubin levels remain in the normal range. This indicates that there was no effect of the treatments on the rats in this regard. Albumin and direct bilirubin levels were found to be very low as compared to their normal ranges. Bilirubin acts as an antioxidant that protects tissues throughout the body from damage by substances that can cause the breakdown of cells. (Jewell, 2018). Given this, lower levels of bilirubin indicate a possibility of certain body parts being vulnerable to damage. Several medical conditions are associated with lower levels of bilirubin in the body. These conditions include ulcerative colitis, brain lesions and diabetic retinopathy (Schieffer et al., 2017; Res, 2017 & Higuchi et al., 2018). On the other hand, lower levels of albumin are associated with inflammation, shock and the inability of the body to absorb and digest protein

5. CONCLUSION

The high elevation of the haematological, liver and renal indices from the study suggests that Copper and Zinc exposure have a deteriorating effect on the blood, liver and other organs of the body, therefore, long-term exposure to these trace elements can lead to a lot of physiological and pathological defects.

5.1. Recommendations

Based on the findings of this current study, it is recommended that;

- further studies should be carried out to ascertain the histopathological effects of copper and zinc on the liver and kidney.
- whenever Cu and Zn treatments are administered orally to rats, there may be a high propensity of the gut of the rats being affected, it is therefore recommended that further investigation be made to study the effects of Cu and Zn on the gut of rats as well as possible microbiota present in the gut.
- the Food and Agriculture Organization (FAO) and the Ministry of Food and Agriculture (MOFA) must ensure strict compliance to permissible levels of trace elements in pesticides used in crop production.
- farmers must be educated on the dangers of breaching the permissible levels of trace elements on-farm produce as well as in the supplementation of animal diets.

COMPETING INTERESTS DISCLAIMER:

- Authors have declared that no competing interests exist. The products used for this
 research are commonly and predominantly use products in our area of research and
 country. There is absolutely no conflict of interest between the authors and producers of
 the products because we do not intend to use these products as an avenue for any
 litigation but for the advancement of knowledge. Also, the research was not funded by
 the producing company rather it was funded by personal efforts of the authors.
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REFERENCES

- Ashajyothi, C., & Handral, H. K. (2018). A Comparative In Vivo Scrutiny of Biosynthesized Copper and Zinc Oxide Nanoparticles by Intraperitoneal and Intravenous Administration Routes in Rats. *Nanoscale research letters*, *13*(1), 93.
- Chaoua, S., Boussaa, S., El Gharmali, A., & Boumezzough, A. (2019). Impact of irrigation with wastewater on the accumulation of heavy metals in soil and crops in the region of Marrakech in Morocco. *Journal of the Saudi Society of Agricultural Sciences*, 18(4), 429-436.
- Dehkhodaei, M., Sahihi, M., Rudbari, H. A., & Momenbeik, F. (2018). DNA and HSA interaction of Vanadium (IV), Copper (II), and Zinc (II) complexes derived from an asymmetric bidentate Schiff-base ligand: multi spectroscopic, viscosity measurements, molecular docking, and ONIOM studies. *JBIC Journal of Biological Inorganic Chemistry*, 23(2), 181-192.
- Dimitrova, A., Russeva, A., Atanasova, M., & Strashimirov, D. (2010). Effects of zinc supplementation on some haematological parameters of spontaneously hypertensive rats. *Trakia Journal of Sciences*, 8(2), 61-65.
- El-Kady, A. A., & Abdel-Wahhab, M. A. (2018). The occurrence of trace metals in foodstuffs and their health impact. *Trends in Food Science & Technology*, 75, 36-45.
- Engwa, G. A., Ferdinand, P. U., Nwalo, F. N., & Unachukwu, M. N. (2019). Mechanism and health effects of heavy metal toxicity in humans *Poisoning in the Modern World-New Tricks for an Old Dog?*: IntechOpen.
- Elwir, S., & Rahimi, R. S. (2017). Hepatic encephalopathy: an update on the pathophysiology and therapeutic options. *Journal of clinical and translational hepatology*, *5*(2), 142.
- Eom, S.-Y., Yim, D.-H., Huang, M., Park, C.-H., Kim, G.-B., Yu, S.-D., . . . Kim, H. (2020). Copper–zinc imbalance induces kidney tubule damage and oxidative stress in a population exposed to chronic environmental cadmium. *International Archives of Occupational and Environmental Health*, 93(3), 337-344.
- Fang, A. P., Chen, P. Y., Wang, X. Y., Liu, Z. Y., Zhang, D. M., Luo, Y., . . . Zhou, Z. G. (2019). Serum copper and zinc levels at diagnosis and hepatocellular carcinoma survival in the Guangdong Liver Cancer Cohort. *International journal of cancer*, *144*(11), 2823-2832.

- Fu, Z., & Xu, S. (2020). The effects of heavy metals on human metabolism. *Toxicology Mechanisms and Methods*, 30(3), 167-176.
- Hughes, S., & Samman, S. (2006). The effect of zinc supplementation in humans on plasma lipids, antioxidant status, and thrombogenesis. *Journal of the American College of Nutrition*, 25(4), 285-291.
- Garg, D., & Payasi, D. K. (2020). Nanomaterials in Agricultural Research: An Overview *Environmental Nanotechnology Volume 3* (pp. 243-275): Springer.
- Ghosh, A., Onsager, C., Mason, A., Arriola, L., & Mubayi, A. (2020). The Role of Oxygen Intake and Liver Enzyme on The Dynamics of Damaged Hepatocytes}:\\Implications to Ischaemic Liver Injury via A Mathematical Model. *bioRxiv*.
- Gruengreiff, K., Reinhold, D., & Wedemeyer, H. (2016). The role of zinc in liver cirrhosis. *Annals of hepatology*, 15(1), 7-16.
- Joni, S. S., Cheshmavar, M., Shoureshi, P., Zamani, Z., Taoosi, N., Akbari, M., & Afzali, M. (2020). Effects of fingolimod treatments on alanine transaminase and aspartate transaminase levels in patients with multiple sclerosis. *International Journal of Physiology, Pathophysiology and Pharmacology*, 12(3), 88.
- Higgins, C. (2016). Urea and the clinical value of measuring blood urea concentration. *Acutecaretesting*. *Org*, 1-6.
- Himoto, T., & Masaki, T. (2018). Associations between zinc deficiency and metabolic abnormalities in patients with chronic liver disease. *Nutrients*, *10*(1), 88
- Kohzadi, S., Shahmoradi, B., Ghaderi, E., Loqmani, H., & Maleki, A. (2019). Concentration, source, and potential human health risk of heavy metals in the commonly consumed medicinal plants. *Biological trace element research*, 187(1), 41-50.
- Kamath, P. S., & Leise, M. D. (2018). Latest Concepts in Inpatient Hepatic Encephalopathy Management. In *Diagnosis and Management of Hepatic Encephalopathy* (pp. 77-97). Springer, Cham.
- Linder, M. C. (2020). Copper Homeostasis in Mammals, with Emphasis on Secretion and Excretion. A Review. *International Journal of Molecular Sciences*, 21(14), 4932

- Lucas, H. R., & Rifkind, J. M. (2013). Considering the vascular hypothesis of Alzheimer's disease: effect of copper associated amyloid on red blood cells. In *Oxygen Transport to Tissue XXXIV* (pp. 131-138). Springer, New York, NY.
- Lü, J., Jiao, W.-B., Qiu, H.-Y., Chen, B., Huang, X.-X., & Kang, B. (2018). Origin and spatial distribution of heavy metals and carcinogenic risk assessment in mining areas at You'xi County southeast China. *Geoderma*, *310*, 99-106.
- Masindi, V., & Muedi, K. L. (2018). Environmental contamination by heavy metals. *Heavy Metals*, 19, 2019.
- Moriles, K. E., & Azer, S. A. (2020). Alanine Amino Transferase (ALT). In *StatPearls* [*Internet*]. StatPearls Publishing.
- Nishito, Y., & Kambe, T. (2018). Absorption mechanisms of iron, copper, and zinc: an overview. *Journal of nutritional science and vitaminology*, 64(1), 1-7.
- Proshad, R., Kormoker, T., Mursheed, N., Islam, M. M., Bhuyan, M. I., Islam, M. S., & Mithu, T. N. (2018). Heavy metal toxicity in agricultural soil due to rapid industrialization in Bangladesh: a review. *International Journal of Advanced Geosciences*, 6(1), 83-88.
- Rai, P. K., Lee, S. S., Zhang, M., Tsang, Y. F., & Kim, K.-H. (2019). Heavy metals in food crops: Health risks, fate, mechanisms, and management. *Environment international*, 125, 365-385.
- Sandeep, G., Vijayalatha, K., & Anitha, T. (2019). Heavy metals and their impact on vegetable crops. *International Journal of Chemical Studies*, 7(1), 1612-1621.
- Satarug, S., Nishijo, M., Ujjin, P., & Moore, M. R. (2018). Chronic exposure to low-level cadmium-induced zinc-copper dysregulation. *Journal of Trace Elements in Medicine and Biology*, 46, 32-38.
- Semu, E., Tindwa, H., & Singh, B. (2019). Heavy Metals and Organopesti-cides: Ecotoxicology, Health Effects and Mitigation Options with Emphasis on Sub-Saharan Africa. *J Toxicol Cur Res*, *3*(010).
- Schieffer, K. M., Bruffy, S.M., Rauscher, R., Koltun, W. A., Yochum, G.S., Gallagher C.J. (2017)

 Reduced total serum bilirubin levels are associated with ulcerative colitis. PLoS ONE 12(6)
 e0179267.https://doi.org/10.1371/journal.pone.0179267

- Shukla, P. K., Misra, P., Maurice, N., & Ramteke, P. W. (2019). Heavy Metal Toxicity and Possible Functional Aspects of Microbial Diversity in Heavy metal-contaminated Sites *Microbial Genomics in Sustainable Agroecosystems* (pp. 255-317): Springer.
- Smith, A. K., Ropella, G. E., McGill, M. R., Krishnan, P., Dutta, L., Kennedy, R. C., ... & Hunt, C. A. (2020). Contrasting model mechanisms of alanine aminotransferase (ALT) release from damaged and necrotic hepatocytes as an example of general biomarker mechanisms. *PLOS Computational Biology*, 16(6), e1007622
- Staniek, H. (2019). The combined effects of Cr (III) supplementation and iron deficiency on the copper and zinc status in Wistar rats. *Biological trace element research*, 190(2), 414-424.
- Sundari, S. K., Prakash, A., Yadav, P., & Kumari, A. (2019). Plant Growth-Promoting Microbes as Front-Runners for On-site Remediation of Organophosphate Pesticide Residues in Agriculture Soils *Phyto and Rhizo Remediation* (pp. 249-285): Springer.
- Tabelin, C. B., Igarashi, T., Villacorte-Tabelin, M., Park, I., Opiso, E. M., Ito, M., & Hiroyoshi, N. (2018). Arsenic, selenium, boron, lead, cadmium, copper, and zinc in naturally contaminated rocks: A review of their sources, modes of enrichment, mechanisms of release, and mitigation strategies. Science of the Total Environment, 645, 1522-1553.
- Tamakhina, A. Y., Dzakhmisheva, I. S. e., Beslaneev, E. V., & Gadieva, A. A. e. (2018). Bioaccumulation of Heavy Metals by Medicinal Plants of the Inula Genus. *Journal of Pharmaceutical Sciences and Research*, 10(5), 1263-1266.
- Yahya, R. A., Attia, A. M., El-Banna, S. G., El-Trass, E. E., Azab, A. E., Jbireal, J. M., & Shkal, K. E. M. (2019). Hematotoxicity induced by copper oxide and/or zinc oxide nanoparticles in male albino rats. *J Biotechnol Bioeng*, *3*(4), 1-7.
- To, U., & Schilsky, M. L. (2018). Introduction to Copper Metabolism and Wilson Disease. In *Management of Wilson Disease* (pp. 1-16). Humana Press, Cham.
- Tsukada, N. (2018) Low Bilirubin Levels Indicate a High Risk of Cerebral Deep White Matter Lesions in Apparently Healthy Subjects. Accessed on: 24th May 2020.Retrieved from: www.nature/scientific report.com

- Zamaneh, H. T. (2018). Effects of zinc supplement on haematological parameters following sixweek endurance exercise in male rats. *International Journal of Green Pharmacy* (*IJGP*), 11(04).
- Wang, Y., Pasparakis, C., Mager, E. M., Stieglitz, J. D., Benetti, D., & Grosell, M. (2019). Ontogeny of urea and ammonia transporters in mahi-mahi (Coryphaena hippurus) early life stages. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, 229, 18-24.
- Willis, M. S., Monaghan, S. A., Miller, M. L., McKenna, R. W., Perkins, W. D., Levinson, B. S., ... & Kroft, S. H. (2005). Zinc-induced copper deficiency: a report of three cases initially recognized on bone marrow examination. *American journal of clinical pathology*, 123(1), 125-131.
- Vazquez, J. H., Clemens, M. M., Allard, F. D., Yee, E. U., Kennon-McGill, S., Mackintosh, S.G., ... & McGill, M. R. (2020). Identification of Serum Biomarkers to Distinguish Hazardous and Benign Aminotransferase Elevations. *Toxicological*

Sciences, 173(2), 244-254.