

### **Wildlife Forensics: A Case Study on Golden Jackal *Canis aureus* in India using canid head and tailor-made musk pod**

#### **Abstract**

The escalating trend of illegal trade on wildlife and wildlife products is a serious threat to the existence of wildlife globally. Effective enforcement of the laws to contain this requires accurate identification of the seizures which is often difficult due to fake lookalike articles. The articles thus pose challenging complexity and necessitate the usage of different approaches in identifying the species. Species identification by the use of molecular technology is widely recognized to be a definitive method. In this report, we describe the identification of seized canid head, suspected to be of jackal and a pod suspected to be of musk deer using the DNA-based approach. The mitochondrial regions of the query samples were amplified using ribosomal markers, 12S rRNA and 16S rRNA. The sequence reads were compared with the homologous hits extracted from the NCBI database. The phylogenetic analysis was carried out with the aligned sequences to identify species with 500 bootstrap replications. Based on the genetic analysis of both the seizures, the tested samples were identified to be belonging to the species 'Golden Jackal'.

**Key words:** Illegal wildlife trade, Golden Jackal, Musk pod, Molecular forensics

#### **1. Introduction**

Use and abuse of wild animals for the benefit of humankind has been prevalent since ancient times. Apart from live animal trafficking, the illegal trade of wildlife products is contributing to a significant portion of wildlife seizure records (Scheffers et al. 2020). Such illicit trade is a complex and lucrative industry worldwide (Rosen & Smith 2010). Severe threats are posed to both flora and fauna due to illicit trade. Though most of the wildlife species are under protection on a national and international scale, only charismatic species like Tiger (*Panthera tigris*) or Asian elephant (*Elephas maximus*) gains greater concern from the public, the lesser known or

less charismatic species do not garner enough attention (Douglas & Alie 2014). Crime on such species generally suffers a low conviction rate and are under-punished (Chawla 2020).

One such species is Golden Jackal (*Canis aureus*), this animal with stable population and wide geographic range is protected under Schedule II Part II of the Wildlife (Protection) Act 'WPA' (1972), 'Least Concern' in IUCN red list, and is listed in CITES Appendix III only for India (Hatlauf et al 2021). Having their major distribution outside the protected areas, Jackals face drastic decline in their population size due to anthropogenic activities (Pillay et al. 2011). Such activities cause survival threats to the animal in the form of hunting and poaching for their body parts like skin, teeth, tail, and specifically for jackal 'siyarsinghi' which translates to 'Jackal horns' (Chawla 2020). It is used in black magic and other superstitious practices in various parts of India (Srinivas & Jhala 2021).

The species identification from the evidence obtained from wildlife crimes usually present a tricky challenge for morphological examinations. Since samples such as dried or processed skin is difficult to analyze, the species identification through DNA-based techniques are more applicable and are well accepted as the fail proof evidence by law (Shen et al. 2013). The DNA barcoding technology is being utilized as a potential tool in wildlife forensics, wherein short regions of mitochondrial DNA (mtDNA) are amplified and sequenced to barcode or identify the species with utmost specificity and distinction from other species (Kress & Erickson 2008). Here, we report on wildlife forensic case study undertaken to ascertain species identity by conducting the molecular analysis using 16S and 12S mtDNA markers on excessively dried and processed skin obtained from the seizures of a canid head and a suspected musk pod sample presented by the State Forest department officials for implementation of Wildlife (Protection) Act, 1972 of India.

## **2. Materials & Methods**

### **2.1 Case history**

The State Forest department officials confiscated two different seizures in Tamil Nadu. The investigating team had collected the evidences of a canid head and a suspected musk pod from the crime sites. The same was forwarded to the Centre for Wildlife Forensic Sciences, Advanced Institute for Wildlife Conservation Institute (AIWC), Vandalur, Chennai. The cases

were registered to undertake molecular analysis for species identification. The canid head sample was marked as 'Seizure 1' and the suspected musk pod was marked as 'Seizure 2' (Fig 1).

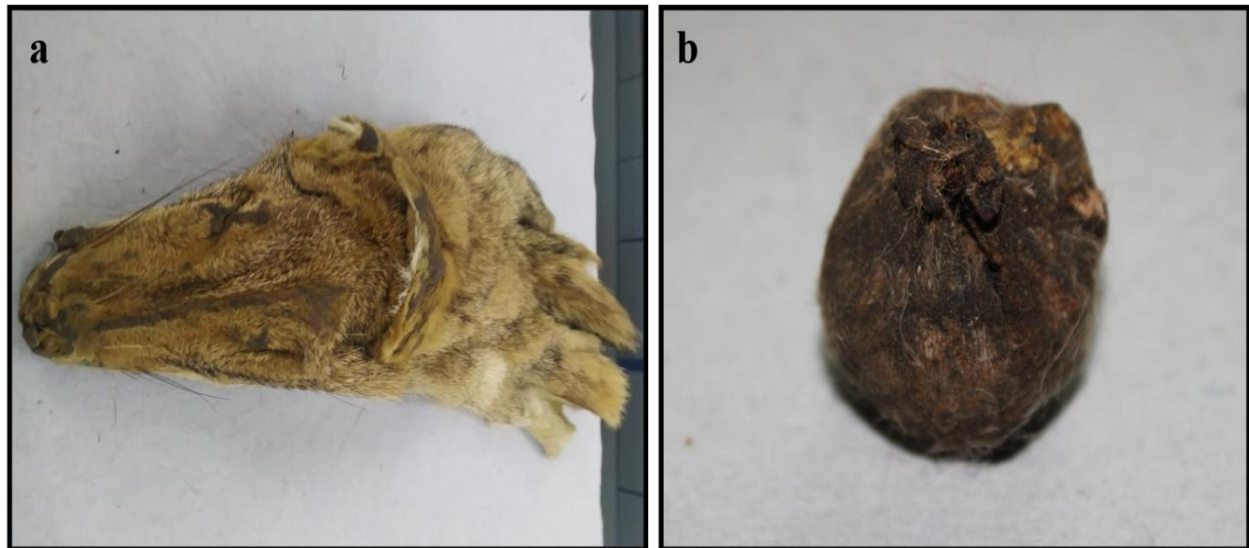


Fig 1: Photographs of seized a. Canid head and b. suspected musk pod

## 2.2 Laboratory procedures

### 2.2.1. Washing

One sq.cm of skin sample from Seizure 1 and 2 were taken in separate 2 ml microcentrifuge tubes and subjected to washing with autoclaved distilled water for 5 minutes as samples were excessively dry (seizure 1) and was suspected to have charcoal contamination as filling material in pod (seizure 2). Upon washing, the samples were transferred to fresh tubes for DNA extraction. In case of seizure 2, both interior and exterior skin portions were taken for examination.

### 2.2.2. DNA extraction, PCR amplification and Sequencing

DNA was extracted following the commercially available Qiagen DNeasy Blood & Tissue kit procedure (Qiagen, Germany) with an extraction blank. The samples were kept under extraction digestion with digestion buffer and proteinase K (KAPA Biosystems, SIGMA), for the period of 48 hours in total, of which 24 hours was in room temperature and 24 hours in circulating water-bath at 56 °C, in order to achieve utmost digestion of the samples. The DNA

extracted from the samples were subjected to amplification of mitochondrial regions of 16S rRNA (Guha & Kashyap, 2006) and 12S rRNA (Kitano et al., 2007) yielding 600bp and 215bp respectively (Supplementary table 1). Independent PCR for both the target genes was carried out in Eppendorf Nexus GSX1 Mastercycler. The reaction tubes of 10  $\mu$ L total volume were set up containing 1X Taq Buffer (KAPA Biosystems, SIGMA), 0.25 mM dNTPs, 0.4  $\mu$ M of both forward and reverse primer, 2.5 mM  $MgCl_2$ , 0.25 U Taq DNA Polymerase (KAPA Biosystems, SIGMA) and 1  $\mu$ L of template DNA. Cycling conditions consisted of 5 min. of initial denaturation at 95 °C, followed by 35 cycles of 30 seconds of denaturation at 95 °C, 30 seconds of annealing at 57°C, 45 seconds of extension at 72 °C and final extension at 72°C for 10 min. The PCR reactions were set with positive and non-template controls. Amplicons were visualized in 2% agarose gel with novel juice DNA stain (SIGMA) and documented using BioRad XR+ gel doc system. The PCR products were purified using QIAquick gel extraction kit (Qiagen, Germany) and sequenced bi-directionally by Sanger sequencing in ABI 3730 DNA Analyzer (Applied Biosystems, USA) using ABI Big Dye TM Terminator Cycle sequencing kit (Applied Biosystems, USA). The forward and reverse sequences of each sample were aligned and trimmed at both ends and assembled using MEGA X software.

### 2.2.3 Data Analysis

The sequences of each sample were searched against the GenBank database using BLASTn. The top hits of homologous sequences with a percentage match of greater than 98% were downloaded from GenBank and subjected to multiple sequence alignment with the query sequence using CLUSTALW. Genetic distances/ evolutionary divergence matrix and phylogenetic analysis by Neighbour Joining method using the Kimura-2-Parameter model with 500 bootstrap replications were computed using MEGA X software. The sequences including similar and closely related species with 3 hits for each species were selected with respect to each gene. For 16S rRNA region, the sequences extracted from the NCBI database are of Golden jackal (*Canis aureus*), Domestic dog (*Canis lupus familiaris*), Grey wolf (*Canis lupus*) and Japanese wolf (*Canis lupus hodophilax*) for analysis. In the similar fashion, the species picked for 12S rRNA regions are Golden jackal (*Canis aureus*), Coyote/ American Jackal (*Canis latrans*), grey wolf (*Canis lupus*), and Red wolf (*Canis rufus*). The difference in the selected species for 16S rRNA and 12S rRNA is based on the top hits or entries obtained upon search of

the query sequences against the NCBI database. For analysis purpose all the query sequences and the extracted sequences from NCBI database were trimmed to same length.

### 3. Results & discussion

Genomic DNA concentration of the isolated samples ranging from 10 - 40 ng/μL were obtained. The extraction control was eliminated from further processing, as no PCR amplification was observed. The mitochondrial genes (16S rRNA and 12S rRNA) from both the seizures were successfully amplified and sequenced. The 12S rRNA assembled sequences of seizure 1 and seizure 2 were 100% similar to the Indian Golden Jackal (*Canis aureus*). Similarly, 16S rRNA sequences of seizure 1 and seizure 2 were 99.54 % similar to the Indian Golden Jackal (*Canis aureus*). The query samples of each primer were submitted to NCBI database with the accession numbers OL960585, OL989144, OL989145, OL960565. To validate the BLAST search results, evolutionary divergence matrix, neighbour-joining (NJ) tree was constructed for each gene region (Tamura et al. 2004). In evolutionary distance matrix construction, the number of base substitutions per site between sequences are displayed (Kumar 2018). The distance computed between the query sequences and the best hit species from the NCBI database ranged from 0.000 to 0.013 for 16S rRNA sequences. The least divergence from the query sequences was observed in *Canis lupus familiaris* with 0.010 and most divergence 0.013 was observed with the species *Canis lupus* and *Canis lupus hodophilax* (Table 1). The distances between 12S rRNA sequences of query samples and NCBI database entries ranges from 0.000 to 0.005. It is observed from the matrix that query sequences does not have any divergence with closely related species such as *Canis latrans* and *Canis lupus*. The divergence was observed only with *Canis rufus* (0.005) (Table 2).

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Table 1

Estimates of evolutionary divergence between related species of 16S rRNA gene region. The standard error estimations are shown above the diagonal within the table. The analysis was carried out using Kimura two-parameter model.

SPECIES	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
Seizure_1_16S		0.000	0.000	0.000	0.000	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005
Seizure_2_16S	0.000		0.000	0.000	0.000	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005
KT448274.1 <i>Canis aureus</i>	0.000	0.000		0.000	0.000	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005
MZ433379.1 <i>Canis aureus</i>	0.000	0.000	0.000		0.000	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005
MN647063.1 <i>Canis aureus</i>	0.000	0.000	0.000	0.000		0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005
KJ139388.1 <i>Canis lupus familiaris</i>	0.011	0.010	0.011	0.011	0.011		0.000	0.000	0.000	0.002	0.003	0.000	0.002	0.002	0.002
MW916051.1 <i>Canis lupus familiaris</i>	0.011	0.010	0.011	0.011	0.011	0.000		0.000	0.000	0.002	0.003	0.000	0.002	0.002	0.002
MW916049.1 <i>Canis lupus familiaris</i>	0.011	0.010	0.011	0.011	0.011	0.000	0.000		0.000	0.002	0.003	0.000	0.002	0.002	0.002
EU789655.1 <i>Canis lupus familiaris</i>	0.011	0.010	0.011	0.011	0.011	0.000	0.000	0.000		0.002	0.003	0.000	0.002	0.002	0.002
MN699617.1 <i>Canis lupus</i>	0.013	0.012	0.013	0.013	0.013	0.002	0.002	0.002	0.002		0.002	0.002	0.000	0.000	0.000
DQ480506.1 <i>Canis lupus</i>	0.011	0.012	0.011	0.011	0.011	0.004	0.004	0.004	0.004	0.002		0.003	0.002	0.002	0.002
KF661039.1 <i>Canis lupus</i>	0.011	0.010	0.011	0.011	0.011	0.000	0.000	0.000	0.000	0.002	0.004		0.002	0.002	0.002
LC520095.1 <i>Canis lupus hodophilax</i>	0.013	0.012	0.013	0.013	0.013	0.002	0.002	0.002	0.002	0.000	0.002	0.002		0.000	0.000
LC520094.1 <i>Canis lupus hodophilax</i>	0.013	0.012	0.013	0.013	0.013	0.002	0.002	0.002	0.002	0.000	0.002	0.002	0.000		0.000
LC520093.1 <i>Canis lupus hodophilax</i>	0.013	0.012	0.013	0.013	0.013	0.002	0.002	0.002	0.002	0.000	0.002	0.002	0.000	0.000	

Estimates of evolutionary divergence between related species of 12S rRNA gene region. The standard error estimations are shown above the diagonal within the table. The analysis was carried out using Kimura two-parameter model.

Species	1	2	3	4	5	6	7	8	9	10	11	12	13
Seizure_1_12S	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.005	0.004	0.005
Seizure_2_12S	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.005	0.004	0.005
KT448274.1 <i>Canis aureus</i>	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.005	0.004	0.005
MZ433379.1 <i>Canis aureus</i>	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.005	0.004	0.005
KT448277.1 <i>Canis latrans</i>	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.005	0.004	0.005
KT448275.1 <i>Canis latrans</i>	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.005	0.004	0.005
KT448276.1 <i>Canis latrans</i>	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.005	0.004	0.005
MZ042364.1 <i>Canis lupus</i>	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.005	0.004	0.005
MZ042363.1 <i>Canis lupus</i>	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.005	0.004	0.005
MZ042362.1 <i>Canis lupus</i>	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.005	0.004	0.005
MZ367921.1 <i>Canis rufus</i>	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.006	0.006	0.006
MZ367912.1 <i>Canis rufus</i>	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.009	0.009	0.006
MZ367903.1 <i>Canis rufus</i>	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.009	0.009	0.009



The NJ tree analysis based on 16S ribosomal RNA region of mitochondrial gene showed the query sample of both the seizure 1 & 2 clustered with the *Canis aureus* with 99 % bootstrap value and other related species such as *Canis lupus familiaris*, *Canis lupus*, and *Canis lupus hodophilax* were grouped separately (Fig. 2). Likewise, 12S ribosomal RNA mitochondrial gene-based tree displayed clustering of query sequences with *Canis aureus* with 89% bootstrap value. The other species included for the analysis grouped one within the other (Fig. 3).

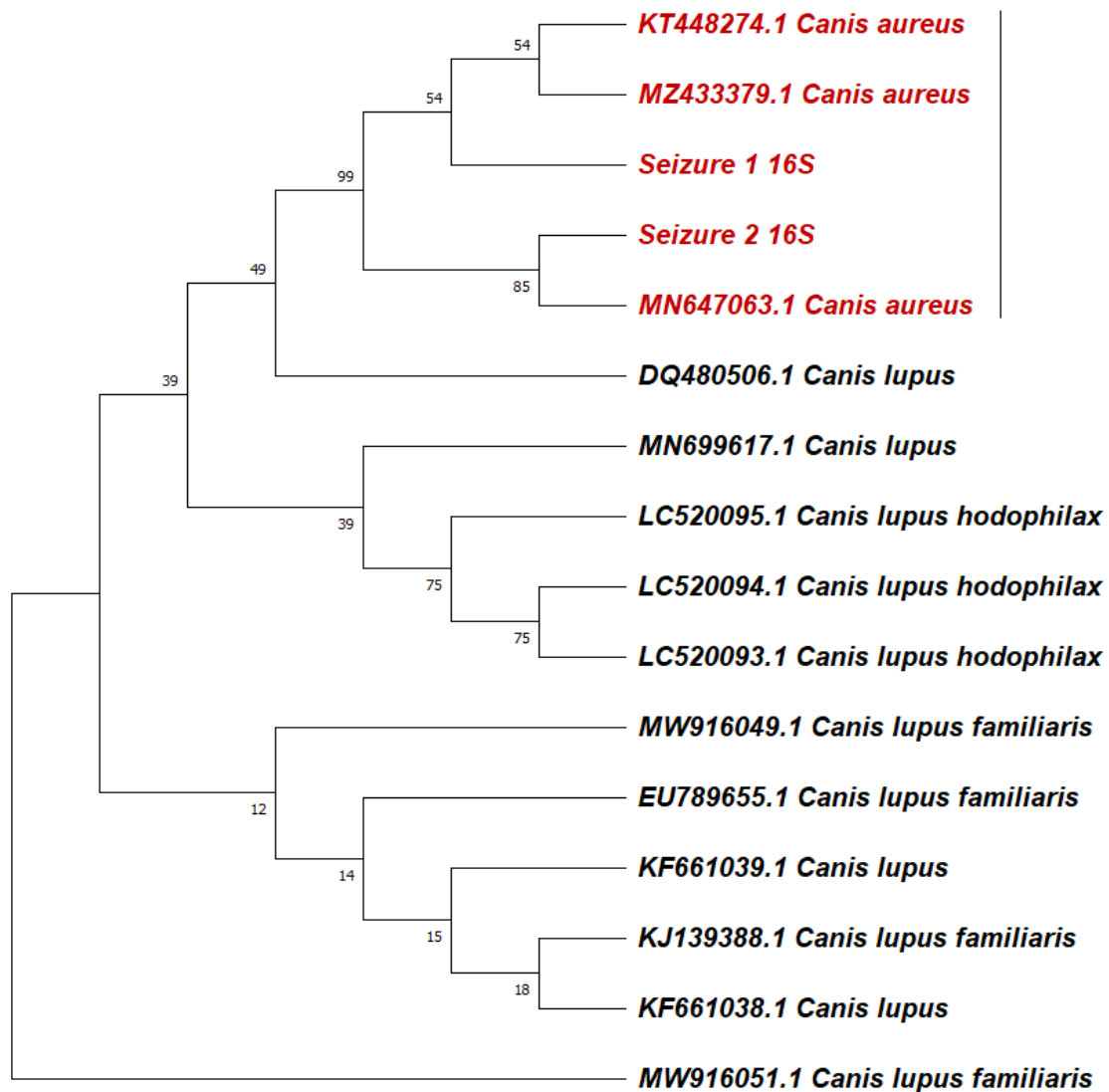


Fig. 2. 16S rRNA gene region-based tree topology displaying the query sequences as Seizure 1 & 2 with the most similar species obtained from the NCBI database through BLAST search.

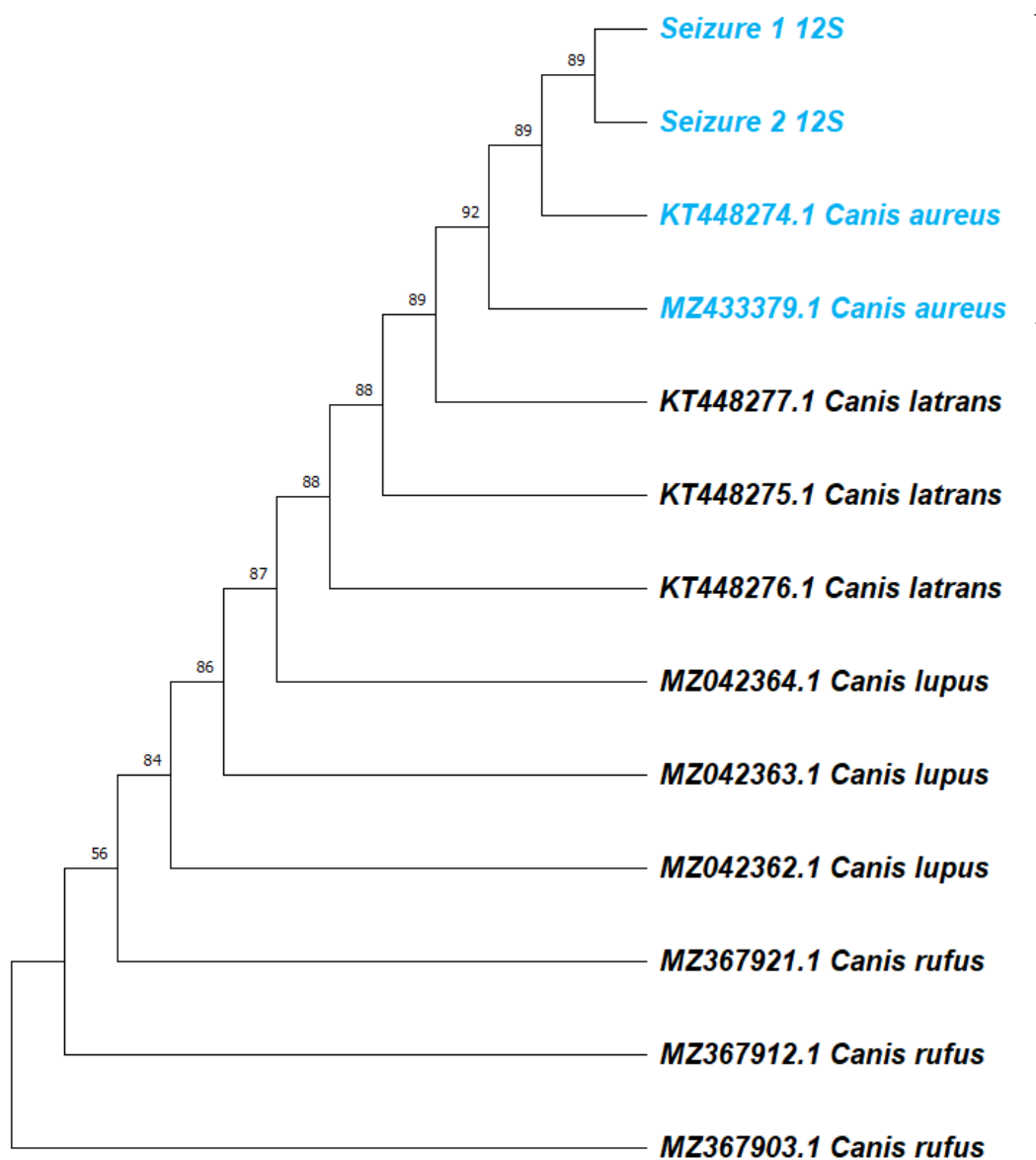


Fig. 3. 12S rRNA gene region-based tree topology displaying the query sequences as Seizure 1 & 2 with the most similar species obtained from the NCBI database through BLAST search.

Generally, the primary choice of markers is either cytochrome b or COI for species identification of mammals (Dalton et al. 2020). Since, the query samples failed in amplification of aforementioned markers, ribosomal markers of mitochondria are used for the study. As far as the employed marker provides a convenient, and accurate species identification, any candidate marker can be used for examination. From the above analysis undertaken on query sequences of

mammalian head and suspected musk pod, it is visibly observed that the examination with a single marker is insufficient as closely related species are most likely to produce similar results within a truncated sequence amplification range (Ghosh et al. 2019). In the analysis carried out on 12S rRNA region, it is observed that our query sequences belonging to *Canis aureus* has 100% similarity with other related species of study *Canis latrans* and *Canis lupus* with 0.000 genetic distances. It shows variation only with *Canis rufus* by 0.005 divergence. Though, 16S query sequences displays discrimination with self and related species, the identity confirmation was achieved based on the results of 2 markers. Hence, we advise the use of more than one molecular marker to confirm the species identity, as mentioned in other studies (Kumar et al. 2021).

The present work targeting the Golden Jackal (*Canis aureus*) underscores the need for more attention to control its prohibited trade as it is typically neglected because of its 'common status' in India. Illicit trade of such a lesser-charismatic species receives hardly any attention causing their decline or loss (Douglas & Alie 2014). The prevalent paucity of attention on jackal hunting provides favourable ground to the poachers or traffickers for the practice of religious or superstitions beliefs involving wildlife articles such as Jackal horns, skin, and skull. Particularly, in the black-magic world, the demand for the supply of musk-pod pushes the sale of fake pods made of jackal skin (Khan et al. 2006; Dangol 2015).

#### **4. Conclusion**

The increasing incidence of wildlife crimes involving not-so attractive wildlife populations poses an unknown threat to the ecosystem globally. Identification of wildlife artefacts by means of physical examination using morphometry usually suffers from lack of reference samples for comparison and also often fails the test of legal scrutiny. DNA examination of wildlife crime samples is considered to be failsafe evidence for species identification. On completion of DNA analysis, we observed that *Canis aureus* was the species of origin for both canid head and the suspected pod, which was confirmed to be a tailor-made pod, reflective of fake pods' circulation in the illegal market. Hence, we recommend the state forest department to conduct capacity-building programs for the forest personnel guarding wildlife, and awareness camps for the public to raise awareness on the Golden Jackal and its parts circulating in the illegal wildlife market.

## COMPETING INTERESTS DISCLAIMER:

Authors have declared that they have no known competing financial interests OR non-financial interests OR personal relationships that could have appeared to influence the work reported in this paper.

## References

American Association for the Advancement of Science. (2020). Erratum for the Research Article:“Global wildlife trade across the tree of life,” by BR Scheffers, BF Oliveira, I. Lamb, DP Edwards. Science, 369(6502), DOI: 10.1126/ science.abd 8164

Rosen, G. E., & Smith, K. F. (2010). Summarizing the evidence on the international trade in illegal wildlife. EcoHealth, 7(1), 24-32, <https://doi.org/10.1007/s10393-010-0317-y>

Chawla, M. M., Srivathsa, A., Singh, P., Majgaonkar, I., Sharma, S., Punjabi, G., & Banerjee, A. (2020). Do wildlife crimes against less charismatic species go unnoticed? A case study of Golden Jackal *Canis aureus* Linnaeus, 1758 poaching and trade in India. Journal of Threatened Taxa, 12, 15407-15413, <https://doi.org/10.11609/jott.5783.12.4.15407-15413>

Douglas LR, Alie K (2014) High-value natural resources: linking wildlife conservation to international conflict, insecurity, and development concerns. Biol Cons 171:270–277, <https://doi.org/10.1016/j.biocon.2014.01.031>

Kress, W. J., & Erickson, D. L. (2008). DNA barcodes: genes, genomics, and bioinformatics. Proceedings of the National Academy of Sciences, 105(8), 2761-2762, <https://doi.org/10.1073/pnas.0800476105>

Shen, Y. Y., Chen, X., & Murphy, R. W. (2013). Assessing DNA barcoding as a tool for species identification and data quality control. PloS one, 8(2), e57125, 10.1371/journal.pone.0057125.

Dalton, D. L., de Bruyn, M., Thompson, T., & Kotzé, A. (2020). Assessing the utility of DNA barcoding in wildlife forensic cases involving South African antelope. Forensic Science International: Reports, 2, 100071, <https://doi.org/10.1016/j.fsir.2020.100071>

Dangol, B. R. (2015). Illegal wildlife trade in Nepal: a case study from Kathmandu Valley (Master's thesis, Norwegian University of Life Sciences, Ås), <http://hdl.handle.net/11250/296693>

Ghosh, A., Basu, S., Khatri, H., Chandra, K., & Thakur, M. (2019). Ascertaining species of origin from confiscated meat using DNA forensics. *Mitochondrial DNA Part B*, 4(1), 329-331, <https://doi.org/10.1080/23802359.2018.1544041>

Hatlauf, J., Bayer, K., Trouwborst, A., & Hackländer, K. (2021). New rules or old concepts? The golden jackal (*Canis aureus*) and its legal status in Central Europe. *European Journal of Wildlife Research*, 67(2), 1-15, <https://doi.org/10.1007/s10344-020-01454-2>

Khan, A. A., Qureshi, B. U. D., & Awan, M. S. (2006). Impact of musk trade on the decline in Himalayan musk deer *Moschus chrysogaster* population in Neelum Valley, Pakistan. *Current Science*, 696-699, <https://www.jstor.org/stable/24094396>

Guha S, Kashyap VK. (2006). Molecular identification of lizard by RAPD & FINS of mitochondrial 16S rRNA gene. *Leg. Med.* 8(1):5-10, <https://doi.org/10.1016/j.legalmed.2005.07.01>

Kitano, T., Umetsu, K., Tian, W., & Osawa, M. (2007). Two universal primer sets for species identification among vertebrates. *International journal of legal medicine*, 121(5), 423-427, <https://doi.org/10.1007/s00414-006-0113-y>

Tamura K., Nei M., and Kumar S. (2004). Prospects for inferring very large phylogenies by using the neighbor-joining method. *Proceedings of the National Academy of Sciences (USA)* 101:11030-11035, <https://doi.org/10.1073/pnas.0404206101>

Kumar S., Stecher G., Li M., Knyaz C., and Tamura K. (2018). MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. *Molecular Biology and Evolution* 35:1547-1549, doi: 10.1093/molbev/msy096

Kumar, V. P., Rajpoot, A., Gupta, A., & Rasaily, S. S. (2021). Forensic investigation of a hunting incident of Indian porcupine (*Hystrix indica*) in Uttarakhand: A study to help rein in biodiversity

loss. Forensic Science International: Animals and Environments, 1, 100002,  
<https://doi.org/10.1016/j.fsiae.2021.100002>

Srinivas Y, Jhala Y (2021) Morphometric variation in wolves and golden jackal in India (Mammalia, Carnivora). Biodiversity Data Journal 9: e67677.  
<https://doi.org/10.3897/BDJ.9.e67677>

Pillay, R., Johnsingh, A. J. T., Raghunath, R., & Madhusudan, M. D. (2011). Patterns of spatiotemporal change in large mammal distribution and abundance in the southern Western Ghats, India. Biological Conservation, 144(5), 1567-1576.  
<https://doi.org/10.1016/j.biocon.2011.01.026>

Supplementary Table 1: List of primers used in the study

mtDNA regions	Primer name	Sequence (5'-3')	Ta (°C)	Amplicon size (bp)
<i>16S rRNA</i> (Guha & Kashyap, 2006)	16L Fwd 16L Rv	CGCCTGTTTATCAAAAACAT CTCCGGTTTGAAGTCAGATC	57	600
<i>12S rRNA</i> (Kitano et al., 2007)	12SrRNAL1085 - F 12SrRNAH1259 - R	CCCAAAGTGGGATTAGATACCC GTTTGCTGAAGATGGCGGTA	57	215