

## Original Research Article

### HEPATOPROTECTIVE EFFECT OF POLY-HERBAL FORMULA (PHF5) ADMINISTRATION ON ALLOXAN-INDUCED DIABETIC WISTAR RAT LIVER FUNCTION ENZYMES

#### ABSTRACT

**Purpose:** Protective action of *Ocimum gratissimum*, *Gnetum africanum*, *Gongronema latifolium*, *Vernonia amygdalina* and *Aloe Barbadensis* leaf extracts (obtained by maceration) was evaluated in an animal model of hepatotoxicity induced by alloxan.

**Methods:** Thirty (30) Wistar male rats were assigned into six (6) groups, with five (5) animals each. Group 1 served as normal control animals; group 2 had the diabetic rats treated with PHF5 (75 mg/kg bw); group 3, diabetic rats treated with PHF5 (150 mg/kg bw); group 4, diabetic rats treated with PHF5 (300 mg/kg bw); group 5, diabetic rats not given any intervention, group 6 diabetic animals treated with Glibenclamide (5 mg/kg bw). The induction of diabetes was done intraperitoneally using alloxan monohydrate (100 mg/kg bw). PHF5 administration was done orally for eight weeks, after which rats were sacrificed by cervical dislocation under light ether anaesthesia. Blood was collected for biochemical evaluation using standard techniques (Randox kits). Fasting blood glucose level was checked at a 7-day interval.

**Results:** Acute toxicity studies of PHF5 did not show any toxic symptoms in animals that received the PHF5 at up to 1000 mg/kg bw dose. The elevated liver (TP, AST, ALT and ALP) markers in the diabetic animals were significantly ( $p < 0.05$ ) lowered in the PHF5 treated animals and found to be within the normal range.

**Conclusions:** The present observations suggested treating *Ocimum gratissimum*, *Gnetum africanum*, *Gongronema latifolium*, *Vernonia amygdalina*, and *Aloe Barbadensis* leaf extracts in different doses of mg/kg enhanced protection against alloxan-induced hepatic damage.

**Keywords:** Alloxan monohydrate, Diabetes, Liver Function Poly-herbal formula,

#### 1. INTRODUCTION

Diabetes mellitus (DM) is one of the most common forms of metabolic disease that affect the world globally [1]. It is a chronic disease caused by changes in the metabolism of carbohydrates, proteins, and lipids due to absolute or moderate insulin deficiency [2,3]. Diabetes mellitus (DM) leads to a risk of several other diseases, many of which are debilitating and followed by an increased risk of dying [3]. Abnormalities observed in DM patients could progress to lesions that include nephropathy, neuropathy, retinopathy and angiopathy [3]. Early-stage symptoms of diabetes are hyperglycemia with hyperinsulinemia as a result of the insensitivity of tissues to insulin. DM is characterized by physiological and cellular changes that result in the demise of beta ( $\beta$ ) cells due to the progression of the disease [4]. Failure of  $\beta$  -cells result from glucose toxicity and lipid toxicity, and an excessive glucose uptake causes glucose toxicity by the islet beta ( $\beta$ )-cells [4]. Elevated sugar levels trigger glycation reactions and

reactions in the electron transport chain, resulting in an imbalance in the cell's antioxidant capacity due to an increase in the production of reactive oxygen species [5]. The consequent oxidative stress leads to a decrease in insulin production and secretion, initiating a sequence of cellular events that ultimately lead to death [5]. In addition to the burden of living with diabetes, it is associated with high morbidity and mortality rates and therefore accounts for a large part of the public health care expenses. Intensified diabetes research is therefore required to improve our knowledge and ability to prevent and treat diseases.

Evaluation of plant products to treat diabetes is of growing interest as they contain many bioactive substances with therapeutic potential [6]. Several plants are efficient ameliorators of stress associated with diabetes. Although many medicinal plants have already been tested for their hepatoprotective properties, these effects remain to be investigated in various other medicinal plants [6].

*Ocimum gratissimum*, also known as basil from the Family of Lamiaceae, is a culinary herb with a pungent sweet smell [7]. The leaves are often used fresh in cooking or added at the last minutes, as cooking destroys the flavour quickly [8]. Studies have established that basil oil has potent antioxidants, anticancer, antiviral and antimicrobial properties [9]. The leaves extract has also been reported to possess antidiabetic properties in alloxan-induced diabetic rats [10,11].

In the South-Western and South-Eastern parts of Nigeria, *Gnetum africanum* (eru or African jointfir), *Gongronema latifolium*, commonly called "utazi" and "arokeke," and *Vernonia amygdalina*, commonly called bitter leaf" in English because of its bitter taste, are mainly used as a vegetable for soups and stews [12,13]. *Gnetum africanum* is a good source of both essential and non-essential amino acids, which contains high levels of leucine, aspartic acid, and glutamic acid, with low levels of histidine and cysteine. The leaves can treat nausea, sore throats, or dressing for warts [14].

Cold concoctions of *Vernonia amygdalina* are used to treat malaria, intestinal parasites, diarrhoea, and stomach pain [14]. A concoction of this plant is also used for malarial fever, schistosomiasis, amoebic dysentery, and many other intestinal parasites and stomach pains in many African ethnic groups. [15].

Traditionally folk medicine uses *Gongronema latifolium* [16]. Digestive problems such as dyspepsia, anorexia, colic and stomach pain, constipation, dysentery, intestinal worms, hyperglycemia, and hypertension are commonly treated with an infusion or decoction of the whole plants (the leaves, stems,

and roots) [17, 18, 19,20, 21]. Studies by Aka et al. [22] showed the antidiabetic activity of aqueous extract (AE) and methanol extracts and fractions of *G. latifolium* in alloxan-induced diabetic rats.

Emphatically, medicinal plants including *Ocimum gratissimum*, *Gnetum africanum*, *Gongronema latifolium*, *Vernonia amygdalina*, and *Aloe Barbadensis* were selected for the preparation of poly-herbal formulation, used to investigate their hepaprotective properties in alloxan-induced diabetic rats.

## **2. MATERIALS AND METHODS**

### **2.1 COLLECTION AND PREPARATION PLANT MATERIALS**

The leaves of *Vernonia amygdalina* (VA), *Gongronema latifolium* (GL), *Ocimum gratissimum* (OG), *Gnetum Africanum* (GA) and *Aloe barbadensis* (AB) were purchased from a local market, Oriugba market in Umuahia Abia state and were authenticated by a Taxonomist (Dr Ibe K. Ndukwe) from the forestry department, College of Natural Resources and Environment Management (CNREM), Michael Okpara University of Agriculture Umudike. The leaves were peeled, washed with distilled water and then air-dried (at a temperature of 27 °C) to a constant weight. After one week, the leaves were milled into a powdered form using a mechanical homogenizer. The powders of the different plants were stored in clean containers and labelled accordingly.

### **2.2 PREPARATION OF PHF5**

The powders of the different plants VA, GL, OG, AS and AB were mixed in the ratio of 3:3:3: 2:1 respectively to derive the poly-herbal formula (PHF) used for this study. The PHF was dissolved in hot distilled water and filtered after 2 minutes. The filtrate was freshly prepared for each administration and was used to treat the animals in this study.

### **2.3 EXPERIMENTAL ANIMALS**

Thirty (30) male Wistar rats (80-120g) were purchased from an animal farm at the University of Nigeria Nsukka and used for this study. The animals were housed in aluminium cages (6 animals per cage) in clean conditions at an ambient temperature of 25°C with a 12-hour light /dark cycle. They were fed standard feed and water *ad libitum*. The animals were acclimatized for 7 days before the commencement of the experiment. The Principles of Laboratory Animal Care (NIH, 1985) were followed throughout this

study. All experimental procedures were conducted according to the animal ethics committee. The induction of diabetes was done intraperitoneally using alloxan monohydrate at the dose of 100 mg/kg bw

## **2.4 INDUCTION OF DIABETES**

Alloxan monohydrate was dissolved in normal saline and administered to the animals at a dose level of 100mg/kg intraperitoneally to induce Type 2 diabetes. The animals were evaluated for fasting blood glucose levels 72 hours after administration of the drugs to confirm induction. A fasting blood glucose level of 140 mg/dl was the criterion for selecting diabetic rats.

## **2.5 DOSE SELECTION**

The PHF was administered to the animals daily for 8 weeks. The doses were calculated for humans and modified for rats using the method of Paget and Barnes [23].

## **2.6 EXPERIMENTAL DESIGN**

A total of 30 rats were used, and they were divided randomly into 6 different groups of 5 animals per cage and treated as follows:

Group 1 – Normal control.

Group 2 – 100mg/kg bw of Alloxan + low dose (75 mg/kg bw) of PHF5

Group 3 – 100mg/kg bw of Alloxan + medium dose (150 mg/kg bw) of PHF5

Group 4 – 100mg/kg bw of Alloxan + high dose (300 mg/kg bw) of PHF5

Group 5 – 100mg /kg bw of Alloxan only

Group 6 – Glibenclamide (Glanil) (5 mg/kg bw) + 100mg /kg bw Alloxan

The administration of PHF5 was done orally, and the fasting blood glucose level was checked at a 7-day interval using a glucometer during the treatment period by collecting blood from the tail vein of the animals. Bodyweight variations were monitored weekly for all the experimental animals.

## **2.7 ACUTE ORAL TOXICITY**

The acute oral toxicity of the poly-herbal formulation was carried out following the Organization for Economic Cooperation and Development (OECD) guidelines. Three animals per dose were used for the experiment. Overnight fasted rats were orally fed with PHF5 in dose levels of 200, 400, 800, and 1000 mg/kg body weight, respectively. The animals were continuously observed for their behavioural (alertness, restlessness and irritability), touch response, pain response and spontaneous activity, and

autonomic (defecation and urination) profiles for 24 h. After 24 h, the animals were observed for 14 days for mortality.

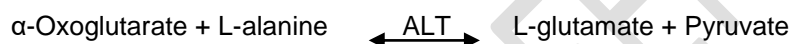
## 2.8 COLLECTION AND PREPARATION OF SAMPLES

At the end of the administration period, the animals were anaesthetized, and blood samples were collected via cardiac puncture. The blood samples were stored in clean vacutainer tubes and centrifuged at 4000 g for 15 minutes. The serum was used for the estimation of biochemical markers such as Alanine aminotransferase (ALT), Aspartate aminotransferase (AST) and Alkaline phosphatase (ALP) using Randox Diagnostic kits.

## 2.9 DETERMINATION OF LIVER FUNCTION MARKERS

### 2.9.1 Assay of serum alanine aminotransferase (ALT) activity

**Principle:** Serum ALT activity was estimated by Reitman and Frankel [24] as outlined in Randox KIT. The method is based on pyruvate production by the transamination activity of ALT reacted with 2,4-dinitrophenylhydrazine (DNPH), which gives a brown-coloured hydrazone that can be measured colourimetrically at 550 nm.



### 2.9.2 Assay of serum aspartate aminotransferase (AST) activity

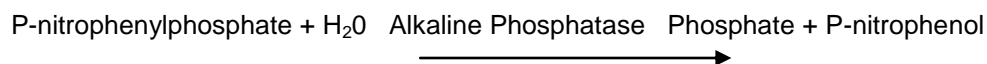
**Principle:** Serum AST activity was estimated according to the method of Reitman and Frankel [24], as outlined in Randox Kit. Oxaloacetate reacts with AST, which decarboxylates it spontaneously to pyruvate measured by hydrazone formation after pyruvate reacts with 2, 4-dinitrophenyl hydrazine (DNPH), which gives a brown-coloured hydrazone that can be measured colourimetrically at 510nm.



### 2.9.3 Assay of serum alkaline phosphatase (ALP) activity

The activity of alkaline phosphatase (ALP) was assayed using the method of Kochmar and Moss [25].

**Principle:** In the presence of magnesium and zinc ions, p-nitrophenol phosphate is hydrolyzed by phosphatase to form phosphate and p-nitrophenol. The p-nitrophenol released is proportional to the alkaline phosphatase (ALP) activity and can be measured photometrically.



### 2.9.4 Serum total protein

Total protein estimation was assayed using the direct Biuret method [26].

**Principle:** This method's principle is that serum proteins react with copper sulphate in sodium hydroxide to form a violet complex called the Biuret complex. The intensity of the violet colour is proportional to the concentration of the protein.

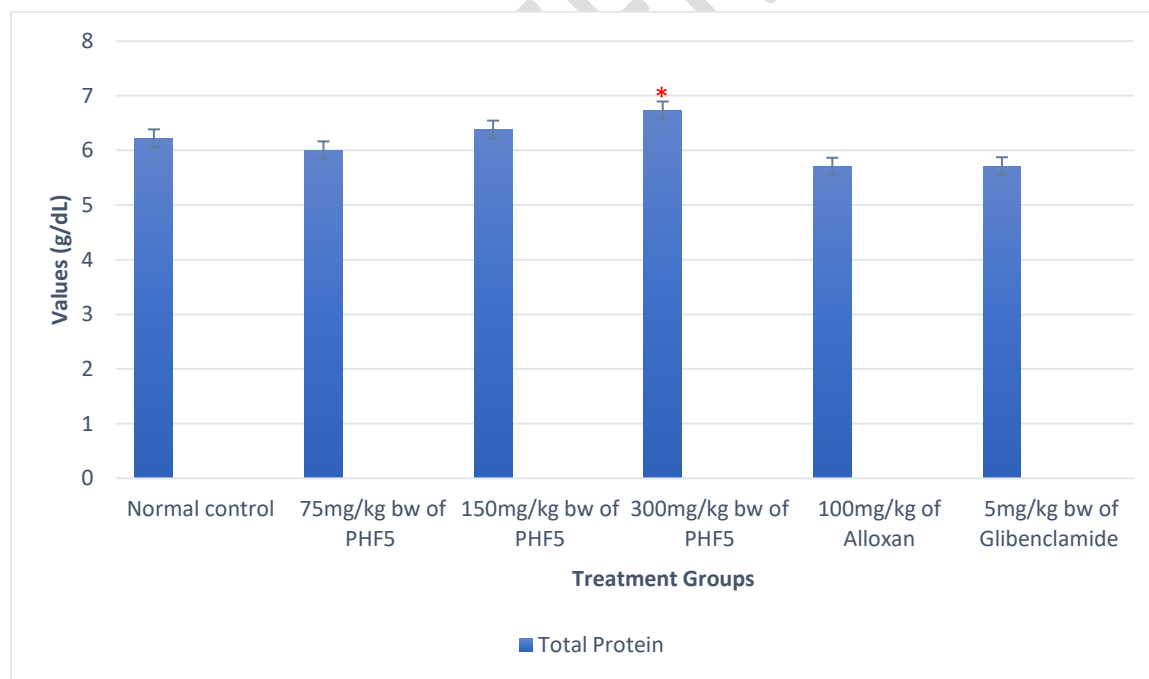
## 2.10 STATISTICAL ANALYSIS

Data obtained was expressed as mean  $\pm$  SD and statistically analyzed using one-way analysis of variance (ANOVA) with Turkey's multiple comparison post hoc tests to compare the level of significance between the test groups. The values of  $p < 0.05$  were considered significant.

## 3. RESULT

### 3.1 Toxicity study of PHF5

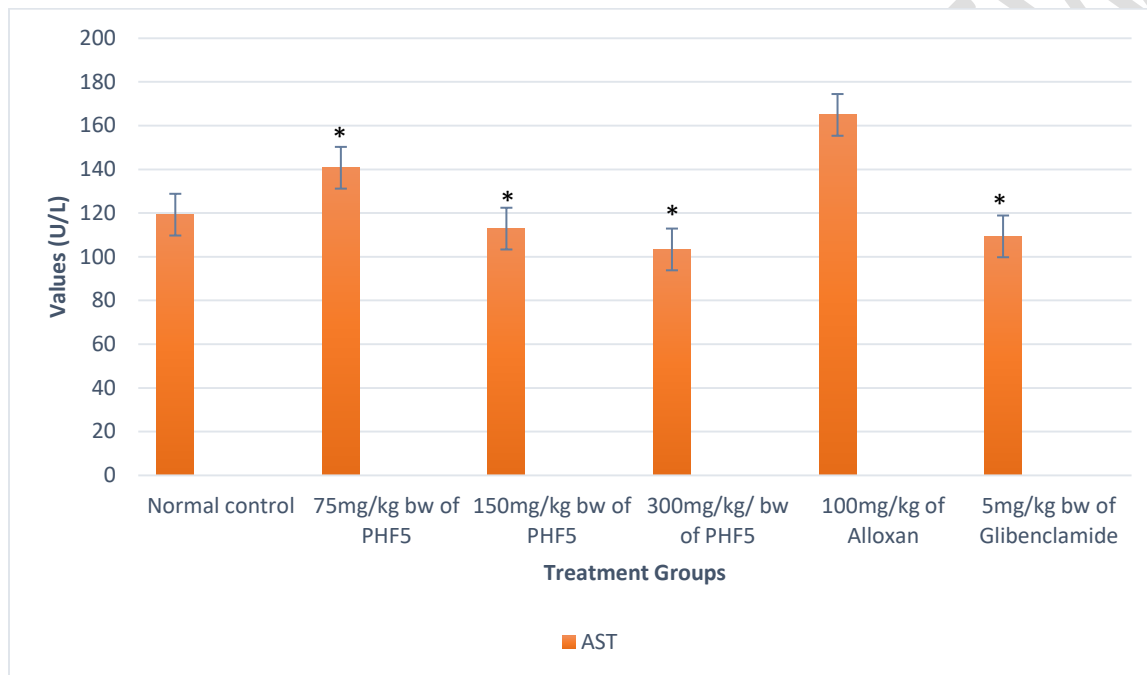
Acute toxicity studies showed no mortality up to 2000 mg/kg given as single oral administration mg/kg. The study was done at three different dose levels (75, 150, and 300 mg/kg).



**Fig.1 Effect of PHF5 Extract on Total protein**

PHF5: Polyherbal formula. Values are expressed as mean  $\pm$  SEM (n=6). \* $p < 0.05$  when compared with the negative control TP: Total Protein.

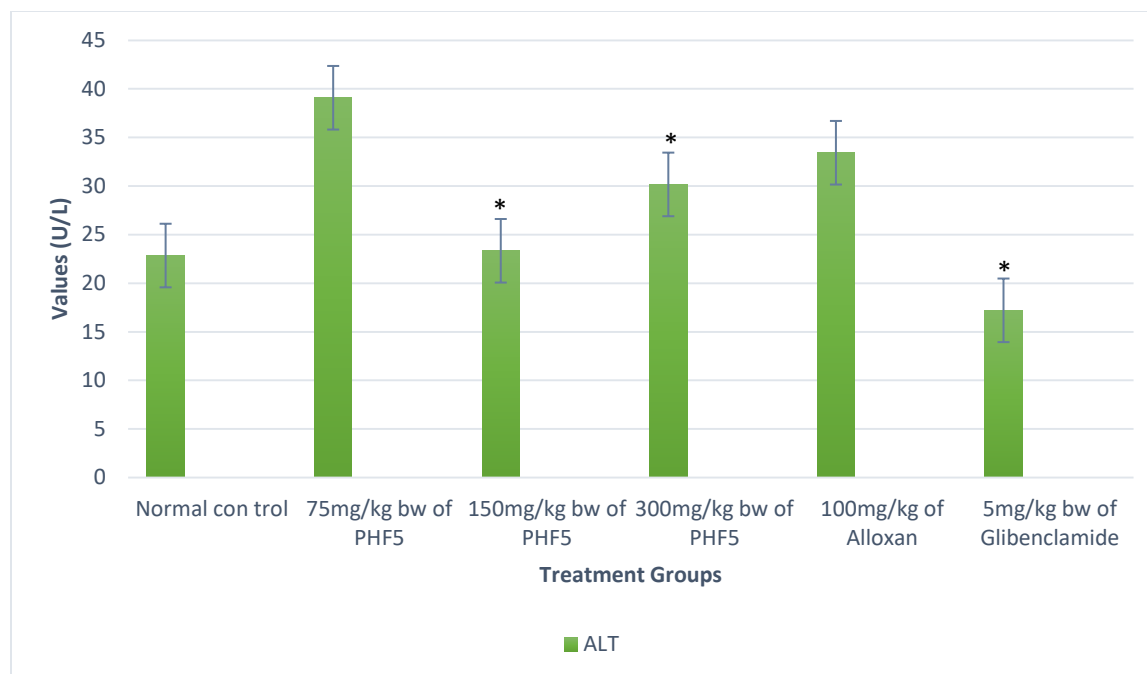
The results of the total protein activities of the studied rats are presented in Fig.1. As shown in the Figure, there was a statistically significant ( $p<0.05$ ) decrease in the total protein of the diabetic control group when compared to the normal control. However, at 300 mg/kg, the PHF5 was able to significantly ( $p<0.05$ ) increase total protein in the animals compared to normal control.



**Fig. 2. Effect of PHF5 Extract on Aspartate Amino Transferase (AST) serum enzymes**

PHF5: Polyherbal formula. Values are expressed as mean $\pm$ SEM (n=6). \* $p<0.05$  when compared with the negative control. AST: Aspartate Amino Transferase

Data for the AST activities were significantly ( $p<0.05$ ) increased in the diabetic control animals compared to the normal animals. At 75 mg/ kg, 150 mg/kg and 300 mg/kg, the PHF5 significantly decreased AST compared to the normal control.

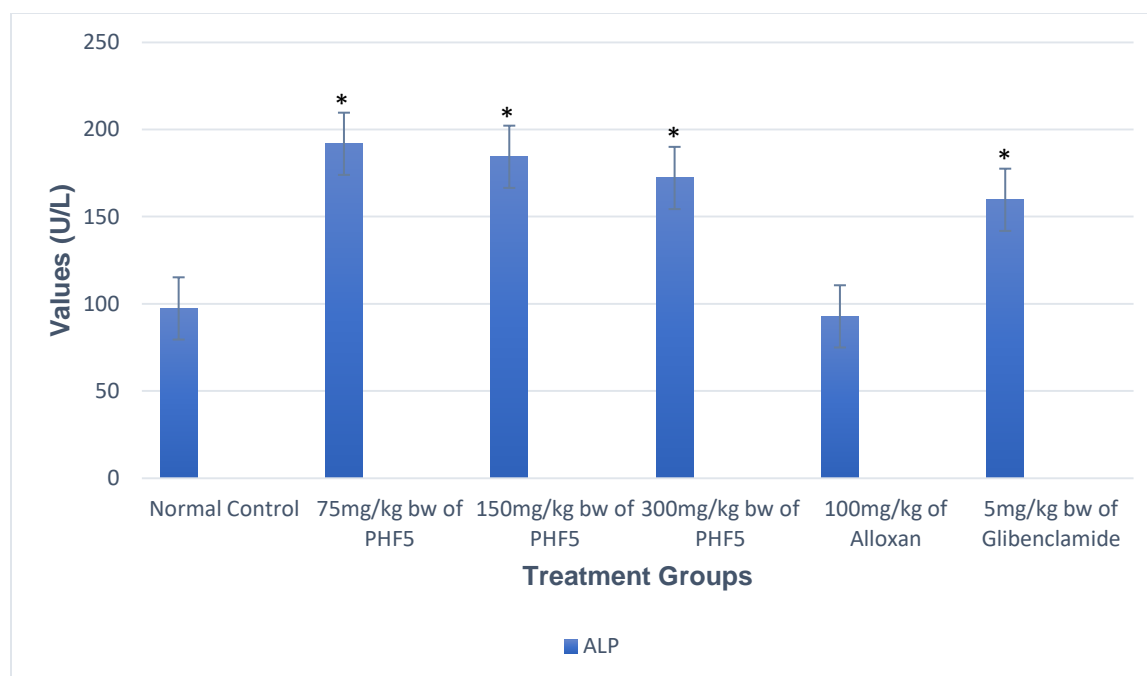


**Fig. 3. Effect of PHF5 Extract on Alanine Amino Transferase (ALT) serum enzymes**

PHF5: Polyherbal formula. Values are expressed as mean  $\pm$  SEM (n=6). \* $p < 0.05$  when compared with the negative control. ALT: Alanine Amino Transferase

Data for the ALT enzyme activities were significantly ( $p < 0.05$ ) increased in the diabetic control animals compared to the normal animals. At 150 mg/kg and 300 mg/kg, the PHF5 significantly decreased AST compared to the normal control.





**Fig. 3. Effect of PHF5 Extract on Alkaline Phosphatase (ALP) serum enzymes**

PHF5: Polyherbal formula. Values are expressed as mean  $\pm$  SEM (n=6). \* $p<0.05$  when compared with the negative control. ALP: Alkaline Phosphatase

Data for the ALP enzyme activities were significantly ( $p<0.05$ ) reduced in the diabetic control animals compared to the normal animals. Supplementation of poly-herbal formula at 150 mg/kg and 300 mg/kg significantly elevated ALP activity compared to the diabetic control animals. The Glibenclamide-treated animals also showed significantly higher ALP activity than the diabetic control.

## DISCUSSION

Herbal formulations have gained much attention in treating diseases, owing to their effectiveness with minor known side effects and easy access [27]. According to studies, polyphenolic compounds effectively prevent the development of long-term diabetes and its complications [28]. This present study investigated the hepatoprotective effect of poly-herbal formula (PHF5) administration on alloxan-induced diabetic Wistar rat liver function enzymes.

In the present study, hyperglycemia caused by alloxan resulted in a significant ( $p<0.05$ ) decrease in total protein, ALP and an increase in AST and ALT plasma levels. The decrease in total protein level could indicate a decrease in the rate of protein synthesis or an increase in protein breakdown, following the previous studies by Suriawinata and Thung [29]. Recently, decreased protein synthesis has been accord

for microproteinuria, which has been accounted to precede the development of overt nephropathy in diabetes mellitus [30].

The levels of alanine transaminase, aspartate transaminase, and alkaline phosphatase are essential indicators for assessing the severity of the injury. The effects of hyperglycaemia-induced oxidative stress on the liver are primary organs susceptible to it [31]. The increase in activities of these enzymes can be attributed to alloxan toxicity which leads to liver damage. However, the dose-dependent (75 mg/ kg, 150 mg/kg and 300 mg/kg) treatment with poly-herbal formula (PHF5) for eight weeks was able to attenuate the damage caused to the liver as evidenced by the reduced enzyme activity in the animals. This result was in agreement with the previous of Thapa and Anuj [32], who reported that ALT (10 – 55  $\mu$ /L), AST (10 – 40  $\mu$ /L), and ALP (45 – 115  $\mu$ /L) are the standard range of accepted values for liver function tests, beyond which liver disease can be suspected. From our findings, liver function parameters remained within normal range after administering a polyherbal formula (PHF5) in all the treated groups, suggesting the hepatoprotective properties of the alloxan-induced Wistar rats, which is in tandem with Thapa and Anuj [32].

## **CONCLUSION**

Using these results, we demonstrate the hepatoprotective value of the polyherbal formulation given to alloxan-induced diabetic animals at doses 75, 150 and 300 mg/kg. Decreased TP, ALT, AST, and ALP levels demonstrated the polyherbal formulation's hepatoprotective properties. PHF5-treated animals had the same effect as Glibenclamide, a standard antidiabetic drug.

## **ETHICAL APPROVAL**

The study was conducted following the National Institute of Health guidelines, the USA, as approved by the College of veterinary medicine, Michael Okpara University of Agriculture, Umudike. The ethical committee's reference number is: MOUAU/CVM/REC/202015

## REFERENCES

1. Prince OA, Chinwe EO, Chiemeziem AO, Chimaraoke O, Peter, OE. and George C.N. Hypoglycemic effect of methanol extract of pawpaw (*Carica papaya*) leaves in alloxan-induced diabetic rats. *Intl. J Innovat. Sci. Res. Tech.* 2022; 7(1); 627-631. <https://doi.org/10.5281/zenodo.5995940>
2. Dib SA, Russo EMK, Chacra AR. 2012. Tratado de endocrinologia clínica. São Paulo: Editora Rocca.
3. Arkkila PE, Koskinen PJ, Kantola IM, Rönnemaa T, Seppänen E, Viikari JS. Diabetic complications are associated with liver enzyme activities in type 1 diabetes. *Diabetes Res. Clin. Pract.* 2001; 52:113–8.
4. Rosalki B, McIntyre N. Biochemical investigations in the management of the liver disease. Oxford textbook of clinical hepatology, 2nd edition New York; *Oxford. Univer. press*, 2009; 503-521.
5. Kaneto H, Katakami N, Kawamori D, Miyatsuka T, Sakamoto K, Matsuoka T A, Matsuhisa M, Yamasaki Y. Involvement of oxidative stress in the pathogenesis of diabetes. *Antioxid. Redox Signal.* 2007; 9, 355–366 10.1089/ars.2006.1465
6. Singh JS. The biodiversity crisis: A multifaceted review. *Curricular Science.* 2012; 82(6):638-47.
7. Ehiagbonare J E. Micropropagation of *Ocimum gratissimum* L: A multipurpose medicinal plant in Nigeria. *Afri. J. Biotech.* 2004; 6 (1), pp. 13-14.
8. Giami SY, Achinewhu SC, Ibaakee C. Cookies' quality and sensory attributes supplemented with basil (*Ocimum gratissimum*). *J. Food Sci. Tech.* 2005; 40: 613-620.
9. Essawi T, Srour M. Screening of some Palestinian medicinal plants for antibacterial activity. *J. Ethnopharm.* 2000; 70: 343-349.
10. Adebolu T T, Salau A O. Antimicrobial activity of leaf of *Ocimum gratissimum* on selected diarrhoea causing bacteria in South-Western Nigeria. *Afri. J. Biotech.* 2005;4 (7): 682-684.
11. Rabelo M, Souza EP, Mirada AV, Matos FJA. Criddle, Antinociceptive properties of the essential oil of *Ocimum gratissimum*, L. (Labiatae) in mice. Brazilian. *J. medi. and bio. res.*, 2003; 36: 521-524.
12. Ijeh II, Ejike ECC. Current perspectives on the medicinal potential of *Vernonia amygdalina* Del. *J. Medi. Plant Res.* 2011; 5 (7): 1051–1061.

13. Egedigwe CA. Effect of dietary incorporation of *Vernonia amygdalina* and *Vernonia colorata* on blood lipid profile and relative organ weights in albino rats (Thesis). *Department of Biochemistry, MOUAU, Nigeria*. 2010; 20-32.
14. Asante D. Anti-inflammatory, anti-nociceptive and antipyretic activity of young and old leaves of *Vernonia amygdalina*. *Biomed. Pharmacol.*, 2019; 111: 1187–1203.
15. Asante D, Valgimigli L, Pratt DA. Antidiabetic Effect of Young and Old Ethanolic Leaf Extracts of *Vernonia amygdalina*: A Comparative Study. *J. Diab. Res.* 2016; 82 (52) 741 – 745.
16. Okpala B. Benefits of *Gongronema latifolium* (utazi). Global foodbook. [www.globalfoodbook.com/benefitsofG.L\(utazi\)](http://www.globalfoodbook.com/benefitsofG.L(utazi)) accesses 10th November (2015).
17. Morebise O, Fafunso MA, Makinde JM, Olajide O.A. Evaluation of bioactivity of *Gongronema latifolium* leaf extract in rodents. *Science Focus*. 2006; 11(1): 27-30.
18. Nwinyi OC, Chinedu NS, Ajani OO. Evaluation of antibacterial activity of *Pisidium guajava* and *Gongronema latifolium*. *J. Med. Plant Res.* 2008; 2: 189-192.
19. Oliver-Bever B. Medicinal plants in tropical West Africa. Camb. Univ. Press., London. 1986; 89-90.
20. Owu DU, Nwokocha CR, Obembe AO, Essien AD, Ikpi DE, Osim EE. Effect of *Gongronema latifolium* ethanol leaf extract on gastric acid secretion and cytoprotection in streptozotocin-induced diabetic rats. *West Ind. Med. J.* 2012; 6(9): 853-860.
21. Ugochukwu NH, Babady NE. Antihyperglycemic effect of aqueous and ethanolic extracts of *Gongronema latifolium* leaves on glucose and glycogen metabolism in livers of normal and streptozotocin-induced diabetic rats. *Life Science*. 2003; 73: 1925-1938.
22. Aka PA, Uzodinma SU, Okolo CE. Antidiabetic activity of aqueous and methanol extract and fraction of *Gongronema latifolium* (Aselepidaceae) leaves in Alloxan diabetic rats. *J. Appl. Pharm. Sci.* 2011;1(9): 99- 102.
23. Paget GE, Barnes JM. Evaluation of Drug Activities, Academic Press, Massachusetts. 1964; 135-166. <https://doi.org/10.1016/B978-1-4832-2845-7.50012-8>
24. Reitman S, Frankel AS. *J. Clin Pat*, 1957;28; 56-63
25. Kochmar JF, Moss DW. Fundamentals of clinical chemistry, NW Tietz (ed). WB Saunders and Company, Philadelphia, PA, 1976; 604.

26. Gornall AG, Bardawill CS, David MM. Determination of serum proteins utilizing the biuret reactions. *J Bio. Chemi.* 1948; 177:551.
27. Zhang Y, Zhen W, Maechler P and Liu D: Small molecule kaempferol modulates PDX-1 protein expression and promotes pancreatic  $\beta$ -cell survival and function via CREB. *J NutrBiochem.* 2013; 24: 638-646, 2013.
28. Bahadoran Z, Mirmiran P and Azizi F: Dietary polyphenols as potential nutraceuticals in management of diabetes: A review. *J Diabetes Metab. Disord.* 2013; 12: 43, 2013. 35.
29. Suriawinata AA, Thung SN. Liver pathology an Atlas and concise guide Latest Edition 2011
30. Bakris GL, Molitch M. Microalbuminuria as a risk predictor in diabetes: the continuing saga. *Diabetes Care.* 2014; 37(3):867-875.
31. Longo DL, Fauci AS, Kasper DL, Hauser SL, Jameson JL, Loscalzo J. Harrison's Principles of Internal Medicine. 18th ed. Vol. 2. New York, USA: McGraw-Hill; 2011.
32. Thapa BR, Aniy W. Liver function tests and their interpretation. *Ind. J. Pediat.* 2007; 74 (7): 663 – 671.