

Original Research Article

A Comparative Analysis of Phytoconstituents, Cytotoxicity and Antidiabetic Activity of Three Regional *Piper betle* Varieties: First Report From Bangladesh

Abstract

Aim: *Piper betle* is one of the significant species of Piperaceae family that exhibit remarkable medicinal potentials including anticancer and antidiabetic activity. The present study aimed to conduct a comparative study on different regional betel leaves focusing on identification of active phytochemicals and evaluating cytotoxic and antidiabetic activity of leaf extracts.

Methodology: The *P. betle* plant leaves of Moheshkhali, Shaplapur, and Rajshahi regions were included in the study. Active phytochemicals in leaves of different varieties were screened. In addition, brine shrimp lethality assay and α -glucosidase inhibitory assay were conducted for investigating cytotoxicity and antidiabetic activity respectively of the leaf extracts.

Results: We found alkaloid, saponin, tannin, glycoside, phenol, steroid, terpenoid, and flavonoid except anthocyanin in phytochemical screening of three *P. betle* varieties. In cytotoxicity assay, the dichloromethane extract of Rajshahi variety showed the highest cytotoxic effect (100% mortality) with LC₅₀ at 2.91 μ g/mL in comparison with Moheshkhali (LC₅₀ at 30.47 μ g/mL) and Shahplapur variety (LC₅₀ at 79.98 μ g/mL). In antidiabetic assay, the highest (100%) α -

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glucosidase inhibition activity was observed by the n-hexane extract of Rajshahi variety with IC₅₀ at 0.001 µg/mL followed by 0.004 µg/mL and 0.015 µg/mL of Shaplapur and Moheshkhali variety respectively.

Conclusion: Collectively, our study identified a number of active phytochemical constituents among the three varieties of *P. betle* leaf extracts as well as potent ability of different leaf extracts in cytotoxicity and antidiabetic activity. We hope the *P. betle* leaf extracts of Moheshkhali variety can play significant role as potential medicinal use in treating cancer and diabetes.

Keywords *P. betle*; Phytoconstituents; Cytotoxicity; Antidiabetic activity; Moheshkhali variety

1. Introduction

Plants are a valuable source of natural products which are used as pharmaceuticals, flavors, fragrances, colors, bio-pesticides and food additives [1]. Plant-based natural products can be isolated from any part of the plant like bark, leaves, roots, fruits, seeds, fruit rind, etc. Any part may contain active component [2]. The use of plants as medicine in treating diseases is as old as human being. Medicinal plants have been used for centuries with a strong belief in their effectiveness in curing diseases. Although modern medicine is well developed, large sections of the populations in developing countries still rely on medicinal plants and herbal medicines in primary health care [3]. More than 80% of the world's population relies on plant-based medicine for primary healthcare, a system that developed over time by dynamic interactions between

people and their environment [4]. Several modern drugs inspired by the traditional plant medicine system are now in use, prescribed by the formal doctrines, in the treatment of various diseases. Since ancient times researchers are trying to find solutions to various diseases using the effective role of alkaloids, i.e., phytochemicals from medicinal plants which can also be used as candidates for the viral vaccine [5,6].

Piper betle is one of the significant species of Piperaceae family that exhibit remarkable medicinal potentials. They are dioecious, semi-woody, and perennial climbers with strongly flavored leaves which are commonly used as mouth fresheners [7,8,9]. The betel chewing criteria is famous specially in south-east Asia and the Indian subcontinent and its popularity includes Melanesia to Tikopia in the east, Africa to Madagascar in the west, Papua New Guinea in the south and southern China in the north [10]. However, *P. betle* contains a wide variety of phytoconstituents. Terpene-like bodies and phenols are the reason behind the strong pungent aromatic flavors of betel leaves [11]. Betel leaves are found to contain sugars, diastases, starch, and an essential oil composed of allyl pyrocatechol monoacetate, safrole, terpinen-4-ol, eugenol, eugenyl acetate, etc. [12,13]. The terpenoids include 1,8-cineole, camphene, cadinene, caryophyllene, pinene, limonene, chavicol, carvacrol, allyl pyrocatechol, chavibetol, eugenol, and safrole [14-16].

The betel leaf extracts have remarkable hypoglycemic activity. The suspension of leaf can significantly lower the blood glucose level and glycosylated hemoglobin and decrease the activity of liver fructose-1,6-bisphosphatase and glucose-6-phosphatase, but in Streptozocin (STZ) diabetic rats, liver hexokinase increased in comparison with the untreated diabetic rats [17-19]. Besides, studies evaluated that *P. betle* leaf extracts act as potent cytotoxic and possible anticancer property. The cytotoxic compounds in leaf extract also show potential antitumor or

anticancer property [20]. The brine shrimp cytotoxicity assay is considered as a convenient probe for preliminary assessment of toxicity [21].

P. betle is a locally available and widely used plant in Bangladesh. From early times, this plant has been used as a mouth freshener and a chewing agent. Without knowing the benefits of the plant, many local practitioners used to give the sap of betel leaves as remedies in treating several diseases. This point of view has attracted the researchers and made them inquisitive to study the bioactive compounds of betel leaves that are playing the role in the treatment. Though studies have been done on medicinal properties of *P. betle* in Bangladesh, comparative study using different varieties of this potential medicinal plant has not been done yet in our country. So, the present study aimed to conduct a comparative study for the first time on different regional betel leaf plants of Bangladesh focusing on identification of active phytochemicals and evaluating cytotoxic activity antidiabetic and of leaf extracts.

2. Materials and Methods

2.1 Sample collection and processing

The *Piper betle* plant leaves were collected from three specific regions of Bangladesh, i.e., Moheshkhali, Shaplapur, and Rajshahi. After collection, plant leaves were washed followed by air-dried for 3 days, and ground into fine powder. The powder was packed and stored at -20°C for analysis.

2.2 Preparation of plant extract

P. betle leaf extracts were prepared with n-hexane, dichloromethane ethyl acetate and ethanol for increasing polarity. Approximately, 180 gm of powdered leaves of each variety was soaked in 850 ml each organic solvents, followed by filtering. Each filtrate was collected and stored at 4°C till analysis.

2.3 Phytochemical screening

Screening of pharmacologically active substances in the plant extracts was done by applying following tests:

Test for Tannin: 20 ml of distilled water was added to 0.5 g of dried leaf powder, followed by boiling the mixture. Few drops of 0.1% ferric chloride were added to the mixture and a brownish-green color indicated the presence of tannin in the plant extract [22].

Test for Saponin: 2 g of dry leaf powder was soaked in 20 ml of distilled water. The solution was boiled and filtered. 3 drops of olive oil were added to 10 ml of the filtrate and formation of emulsion indicated the presence of saponin [22].

Test for Flavonoid: Leaf powder was soaked in 10 ml of ethyl acetate. The mixture was kept in a steam bath for 3 minutes and then filtered. 1 ml of dilute ammonia was added to 4 ml of filtrate and formation of yellow color indicated the presence of the flavonoid [22].

Test for Steroid: 0.5 g of ethanol extract was dissolved into 2 ml of acetic anhydride. 2 ml of H₂SO₄ was added to the mixture and formation of green color indicated the presence of steroids [22].

Test for Terpenoid: 2 mg of ethanol extract was dissolved in 1 ml of chloroform. 1.5 ml of concentrated H_2SO_4 was added to the mixture and formation of red-brown color at the interface indicated the presence of terpenoid [23].

Test for Anthocyanin: 2 ml of 2 N HCl and ammonium solution was added to 2 ml of aqueous plant extract. The pink-red to blue-violet color indicated the presence of anthocyanin [24].

Test for Alkaloid: In two separate test tubes, a small portion of ethanolic crude extract was acidified with 2 ml of 1% HCl. Each of the solutions was heated. Then few drops of Mayer's reagent were added to one test tube and few drops of Wagner's reagent were added to the other and formation of turbidity indicated the presence of alkaloids [25].

Test for Glycosides: 4 ml of glacial acetic acid and 1 drop of 2% FeCl_3 were added to 10 ml of aqueous plant extract. 1 ml of concentrated H_2SO_4 was then added to the mixture and formation of a brown ring between layers indicated the presence of glycoside [26].

2.4 Cytotoxic activity

The cytotoxicity of the plant extracts was measured by using brine shrimp lethality bioassay [27]. A stock concentration of 400 mg/ml in DMSO was prepared for each extract and subsequently used to make dilutions of 200 mg/ml, 100 mg/ml, 50 mg/ml and 25 mg/ml. Brine shrimp eggs were kept over 24 h to hatch in sea water inside a covered chamber of a duo compartment plastic container. Active nauplii were separated from the eggs and used for the cytotoxicity assay. Nauplii viability was measured in each extract by adding 10 ml of each dilution to bikers containing 10 ml of sea water and 20 nauplii. After 24 h, the surviving nauplii in each tube were counted, and the lethality percentage was calculated for each dilution of each extract by using the following formula

$$\text{Mortality (\%)} = [(Total\ nauplii - Alive\ nauplii) / Total\ nauplii] \times 100$$

2.5 Antidiabetic activity

α -glucosidase inhibitory assay was carried out in 96-well plate followed by Shai et al., 2011 [28]. with some modifications. A reaction mixture of 50 μ L of phosphate buffer, 10 μ L of an enzyme, and 20 μ L of varying concentrations of extract of three different cultivars (400 μ g/ml, 200 μ g/ml, 100 μ g/ml, 50 μ g/ml, 25 μ g/ml and 12.5 μ g/ml) were incubated at 37°C for 15 min. Then, 20 μ l of 5 mM substrate (p-nitrophenyl alpha-D-glucopyranoside) was added and incubated at 37 °C for 20 min. Finally, 0.1 M of Na₂CO₃ (50 μ l) was added to stop the reaction. The released p-nitrophenol absorbance was measured (405 nm) by a microplate reader. DMSO was used as a control [29].

The inhibition percentage was assessed by the following formula:

$$\% \text{ of Inhibition} = [1 - \{(As - As_o) / (Ac - Ac_o)\}] \times 100$$

Where,

As: absorbance of plant extract + enzyme + phosphate buffer + substrate

As_o: absorbance of plant extract + sample + phosphate buffer+ substrate

Ac: absorbance of phosphate buffer + enzyme + substrate + 1% DMSO

Ac_o: absorbance of phosphate buffer + substrate + 1% DMSO

3. Results

3.1 Screening of phytochemical compounds

The phytochemical screening of leaf powder and ethanol extracts of *P. betle* revealed the presence of some secondary metabolites shown in Table 1.

Table 1 Phytochemical constituents of *P. betle* varieties

Tests	<i>P. betle</i> varieties		
	Moheshkhali	Rajshahi	Shaplapur
Steroid	+++	+++	+++
Terpenoid	+++	+++	+++
Alkaloid (Wagner's)	++	+	+++
Alkaloid (Mayer's)	+++	+++	+++
Flavonoid	+++	+++	+++
Saponin	++	-	+++
Tannin	+++	+++	+++
Glycosides	++	+++	+
Anthocyanin	-	-	-

Here, (-) means Absence; (+) means Present; (++) means Moderately present; (+++) means Appreciable amount

3.2 Cytotoxicity of *P. betle* extracts

The cytotoxicity analysis of three varieties of *P. betle* extracts is summarized in Fig. 1. *P. betle* from Moheshkhali region with dichloromethane extract showed an appreciative cytotoxic effect with an LC₅₀ value at 30.47 µg/mL compared to the other three extracts. The LC₅₀ values of n-hexane, ethyl acetate, and ethanol extracts were 82.70 µg/mL, 43.06 µg/mL, and 95.33 µg/mL respectively. The plant extracts showed comparatively lower cytotoxic effect than the positive

control (Vincristine sulphate: LC₅₀ was 10.96 µg/mL). No mortality was found in negative control (DMSO) group.

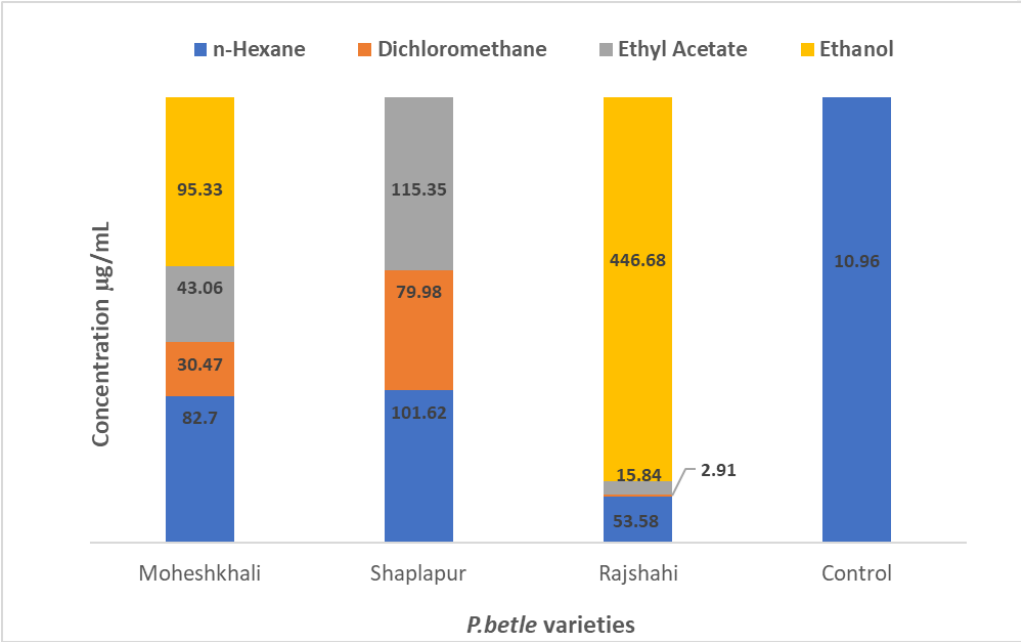


Fig. 1: Cytotoxic effect of different *P. betle* extracts from three regions

Of Shaplapur variety, the dichloromethane extract showed a better cytotoxic effect than n-hexane and ethyl acetate extract with LC₅₀ at 79.98 µg/mL, 101.62 µg/mL and 115.35 µg/mL respectively and each extract showed lower cytotoxic activity than the positive control. The ethanol extract didn't show cytotoxic effect. No mortality was found in the negative control group (Fig. 1).

The dichloromethane and ethyl acetate extract of Rajshahi region showed outstanding cytotoxic effect with LC₅₀ value at 2.91 µg/mL and 15.84 µg/mL respectively compared to n-hexane and ethanol extract. In addition, the dichloromethane extract showed better cytotoxic effect compared to the positive control. No mortality was found in the negative control group (Fig. 1).

3.3 Antidiabetic activity (α -glucosidase inhibitory assay)

The analysis of antidiabetic activity of three *P. betle* varieties is summarized in Fig. 2. The IC₅₀ values of n-hexane, dichloromethane, ethyl acetate, and ethanol extracts of Moheshkhali variety were 0.015 µg/mL, 1.04 µg/mL, 9.49 µg/mL, 9.10 µg/mL respectively. In the contrary, the IC₅₀ of positive control (acarbose) was 26.44 µg/mL. No inhibition was found in the negative control.

The Shaplapur variety also exhibited good inhibition of the α -glucosidase enzyme. The IC₅₀ values of n-hexane, ethyl acetate, ethanol, and dichloromethane extracts were 0.004 µg/mL, 4.69 µg/mL, 10.54 µg/mL, 14.92 µg/mL respectively and all extracts showed better inhibition than the positive control. No inhibition was found in negative control (Fig. 2).

Of Rajshahi variety, the leaf extracts highest inhibition of the α -glucosidase enzyme. n-hexane extract exhibited the best inhibitory effect with IC₅₀ at 0.001 µg/mL. The ethanol and ethyl acetate had almost similar IC₅₀ at 5.02 µg/mL and 5.06 µg/mL respectively. But the dichloromethane extract showed the lowest activity with LC₅₀ of 20.72 µg/mL. No inhibition was found in the negative control (Fig. 2).

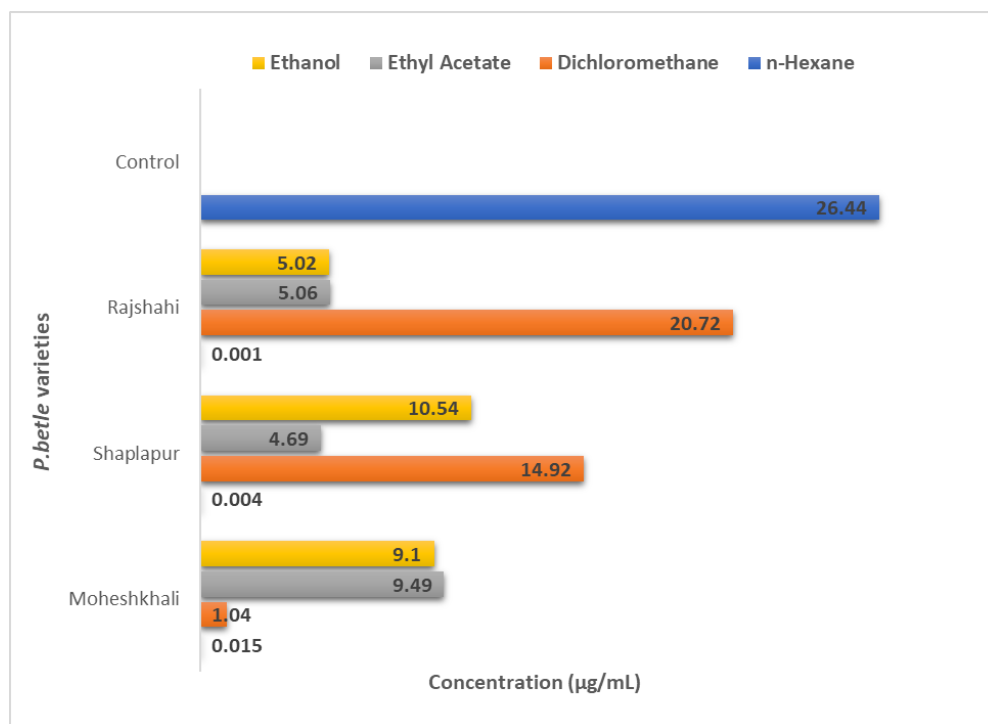


Fig. 2 Anti-diabetic activity of different *P. betle* extracts from three regions

4. Discussion

4.1 Phytochemical analysis

Many polyphenolic compounds, triterpenoids and other chemical compounds exist in leaf extract of plants may account for the observed anti-diabetic effect. *P. betle* leaves are used in much traditional medicines, the anti-tumor property of phytochemicals like polyphenols and alkaloids [30]. The present study identified a number of biologically active phytochemical compounds in the studied three varieties of *P. betle* leaves collected from Moheshkhali, Rajshahi and Shaplapur.

Steroid, terpenoid, alkaloid, flavonoid, saponin, tannin, glycosides were identified among the three varieties except anthocyanin. Among them, steroid, terpenoid, alkaloid, flavonoid, saponin and tannin were tested positive with more intensity in Shaplapur variety. On the other hand, glycosides were higher but saponin was absent in Rajshahi variety (Table 1). In general, the bioactive properties of phenolic compounds are considered as anticarcinogenic, antimutagenic, antioxidant, anti-inflammatory and inducible for apoptosis by cell cycle arrest [31]. A study showed that terpenoid decreases triglycerides and plasma cholesterol in hyperlipidemic rats, and hyperlipidemia plays significant role in developing heart disease [32]. Flavonoids and tannins are phenolic compounds and plant phenolics are a major group of compounds that act as primary antioxidants [33]. In addition, alkaloids exhibit a variety of medicinal properties including anti-hyperglycemic and anti-hyperglycemic activities extracted from piperaceae family [34]. The present study showed that the Shaplapur variety of *P. betle* leaf extract could be considered as an effective source of biologically active phytochemical compounds for designing any drugs for treating diseases.

4.2 Cytotoxicity assay

The study examined the cytotoxic effect of three *P. betle* varieties (Moheshkhali, Rajshahi, and Shaplapur) using Brine shrimp lethality assay and found significant results. The dichloromethane extract from Moheshkhali variety showed better cytotoxic effect with LC_{50} at 30.47 $\mu\text{g/mL}$ compared to ethyl acetate, n-hexane, and ethanol extracts. Similar result was found for Shahplapur variety whereas the dichloromethane extract exhibited cytotoxicity with LC_{50} at 79.98 $\mu\text{g/mL}$ in comparison with n-hexane and ethyl acetate extract, but ethanol extract did not show any effect. Compared to Moheshkhali variety and Shahplapur variety, *P. betle* extracts of

Rajshahi region showed outstanding cytotoxic effect. The dichloromethane extract of Rajshahi variety showed the highest cytotoxicity having LC_{50} at 2.91 $\mu\text{g/mL}$ in comparison with Moheshkhali variety (30.47 $\mu\text{g/mL}$) and Shahplapur variety (79.98 $\mu\text{g/mL}$). However, the ethanol extract of Rajshahi variety found to be the lowest cytotoxicity (Fig. 1). So, the extracts, in order of most cytotoxic to the least cytotoxic, were dichloromethane>ethyl acetate>n-hexane>ethanol. This comparative analysis revealed the dose-dependent cytotoxic effect Rajshahi variety could be considered as antitumor or anticancer potential.

Similar studies reported that ethyl acetate and hexane extract of *P. betle* leaf exhibited a dose-dependent inhibitory effect on the MCF-7 human breast cancer cells with IC_{50} at 65 $\mu\text{g/mL}$ and 163 $\mu\text{g/mL}$ respectively [30]. In addition, methanol extract showed significant activity toward Epstein-Barr virus (EBV) activation in Raji cells at IC_{50} values of 40 $\mu\text{g/mL}$ [35]. A study showed that dichloromethane extract of *P. betle* demonstrated strong cytotoxic effect on MDA-MB-468 and MDA-MB-231 breast cancer cell lines with IC_{50} value of 11.26 ± 0.01 $\mu\text{g/mL}$ and 19.76 ± 2.87 $\mu\text{g/mL}$ respectively [36]. However, a previous study from the country demonstrated that *Piper betle* ethanol extract at LC_{50} value of 274.6 $\mu\text{g/mL}$ showed promising cytotoxic effect in brine shrimp lethality bioassay [37]. Another study showed that ethanol and methanol extract of betel leaf had 50% mortality at 23.65 $\mu\text{g/mL}$ and 85.50 $\mu\text{g/mL}$ respectively [38]. So, *P. betle* leaf extracts exhibited cytotoxic potential and can be used for the treatment of several diseases. The potential lethality of *P. betle* extract against brine shrimp is a sign of the presence of potent cytotoxic components found in the plant. The cytotoxic potential of a compound does not always mean its direct toxic effect, but it can also suggest its antitumor or anticancer potential [39]. The cytotoxicity result of our study indicates the anticancer property of *P. betle* leaf extract which also corroborates the other studies that betel leaf is not carcinogenic.

Scientific studies have also shown that betel leaf does not have mutagenic and/or carcinogenic effects. A study reported for the very first time that aqueous extract of betel leaf failed to cause any tumor in both C17 mice and Swiss albino mice which proved betel leaf was devoid of any carcinogenic compound [40].

4.3 Anti-Diabetic assay

In the present study, α -glucosidase inhibition activity was studied to examine the antidiabetic properties of three *P. betle* leaf varieties (Moheshkhali, Rajshahi and Shaplapur). All the four extracts e.g. ethanol, ethyl acetate, dichloromethane and n-hexane of three different varieties showed significant antidiabetic properties. Based on the IC_{50} obtained in the antidiabetic assay of different extracts, the Moheshkhali variety exhibited the best inhibition followed by the Rajshahi and Shaplapur variety. The highest α -glucosidase inhibition activity was observed by the n-hexane extract with the following IC_{50} at 0.001 $\mu\text{g/mL}$, 0.004 $\mu\text{g/mL}$ and 0.015 $\mu\text{g/mL}$, in Rajshahi, Shaplapur and Moheshkhali varieties respectively. Followed by n-hexane extract, ethyl acetate extracts of the three leaf varieties had better inhibition activity compared to ethanol and dichloromethane. Interestingly, the dichloromethane extract of Moheshkhali variety exhibited the highest inhibition activity with LC_{50} value of 1.04 $\mu\text{g/mL}$ in comparison with Rajshahi (20.72 $\mu\text{g/mL}$ and Shaplapur (14.92 $\mu\text{g/mL}$) variety (Fig. 2).

However, several studies have repeatedly shown the antidiabetic property in *P. betle* extracts [41]. In a study, orally administered *P. betle* (75 and 150 mg/kg of body weight) was found to lower blood glucose and improve body weight at a significant level. The reduction of body weight in STZ (streptozotocin) induced diabetic rats is clear evidence of the degradation of structural proteins caused by diabetes. The structural proteins generally contribute to the body

weight [42]. The protection from huge bodyweight loss seems to be correlated to the ability of *P. betle* to reduce diabetes [43]. Another study reported that blood glucose in rats was significantly reduced by the administration of *P. betle* extract [44]. Similar study on antidiabetic potential of different variety of *P. betle* leaf extracts showed that 50% of α -glucosidase enzyme inhibition by methanolic extracts at 96.8 $\mu\text{g/mL}$, 157.7 $\mu\text{g/mL}$, and >200 $\mu\text{g/mL}$ in Sanchi, Kapoori, and Bangla variety respectively [29]. For the treatment of type 2 diabetes mellitus, α -glucosidase inhibitors are administrated orally as anti-diabetic medications [45]. In diabetes, these pharmacological enzyme inhibitors exhibit short term effect to decrease high blood glucose levels in blood [46]. The difference in total flavonoid content, phenolic content in different cultivars of *P. betle* can be the reason for different antidiabetic activities [47].

The present study aimed to investigate the inhibitory potentials of different varieties of *P. betle* leaf extracts on α -glucosidase, one of the key enzymes involved in carbohydrate hydrolysis. This study revealed that the n-hexane extract of different *P. betle* leaf extracts exhibited the highest role in α -glucosidase inhibition which can be considered as a potential pharmaceutical therapeutic to treat diabetes in Bangladesh. We hope this comparative study among the three *P. betle* varieties can be beneficial with further explorative study in choosing a natural therapeutic alternative for the treatment of cancer and diabetic patients with minimal side effects.

Conclusion

The present study identified a number of active phytochemical constituents among the three varieties of *P. betle* leaf extracts as well as potent ability of different leaf extracts in cytotoxicity and antidiabetic activity. In addition, the *P. betle* leaf extracts of Moheshkhali variety can play a significant role as potential medicinal use in treating cancer and diabetes. Moreover, the

quantification of individual phytoconstituents as well as pharmacological profile based on *in-vivo* studies of Rajshahi, Shaplapur and Moheskhali variety should be further investigated.

References

1. Al-Snafi AE (2013) Chemical constituents and pharmacological activities of Ammi majus and Ammi visnaga. A review. Int J Phar Indus Res 3(3):257-65.
2. Chanda S, Baravalia Y (2011) Brine shrimp cytotoxicity of *Caesalpinia pulcherrima* aerial parts, antimicrobial activity and characterisation of isolated active fractions. Natur Prod Res 25(20):1955-64.
3. Elvin-Lewis M (2000) Should we be concerned about herbal remedies? J Ethnophar 75:141–164.
4. Laird SA (2003) Medicinal plants in international trade: conservation and equity issues. J Ethnophar 2:1-5.
5. Ahmad M, Sultana S, Fazl-i-Hadi S, Ben Hadda T, Rashid S, Zafar M, Khan MA, Khan MP, Yaseen G (2014) An ethnobotanical study of medicinal plants in high mountainous region of Chail valley (District Swat-Pakistan). J Ethnobi Ethnomed 10:1-8.
6. Bailey CJ and Day C (1989) Traditional plant medicines as treatments for diabetes. Diabe Care 12(8): 553-564.
7. Hewageegana HG, Arawwawala LD, Arambewela LS, Ariyawansa HS (2011) *Piper betle* Linn: as a remedy for diabetes mellitus. Int J Rese Ayurv Pharm 2:1601-3.
8. Datta A, Ghoshdastidar S and Singh M (2011) Antimicrobial property of Piper betel leaf against clinical isolates of bacteria. Int J Pharm Sci Rese 2(3):104-109

Comment [SG3]: Pls. italicize all botanical names throughout manuscript

9. Chahal J, Ohlyan R, Kandale A, Walia A, Puri S (2011) Introduction, phytochemistry, traditional uses and biological activity of genus Piper: A review. *Int J Cur Pharma Rev Rese* 2(2):130-44.
10. Rooney DF (1995) Betel Chewing in South-East Asia. *Les plantes du plaisir et de la convivialité en Asie* 15.
11. Pradhan D, Suri KA, Pradhan DK, Biswasroy P (2013) Golden heart of the nature: Piper betle L. *J Pharmacog Phytochem* 1(6):147-67.
12. Annamalai SJ, Subashini SR, Priya JR, Naik R (2016) Betel vine. In *Leafy medicinal herbs: botany, chemistry, postharvest technology and uses* Wallingford UK: CABI.
13. Muhamad II, Hassan ND, Mamat SN, Nawi NM, Rashid WA, Tan NA (2017) Extraction technologies and solvents of phytocompounds from plant materials: physicochemical characterization and identification of ingredients and bioactive compounds from plant extract using various instrumentations. *Ingrd Extrac Physicochem Meth Food* 523-560
14. Kumar N, Misra P, Dube A, Bhattacharya S, Dikshit M and Ranade S (2010) Piper betle Linn. a maligned Pan-Asiatic plant with an array of pharmacological activities and prospects for drug discovery. *Cur Sci* 922-932.
15. Chaurasia S, Kulkarni GT, Shetty LN (2010) Phytochemical studies and in vitro cytotoxicity screening of Piper betle leaf (PBL) extract. *Int Rese J Phar* 1(1):384-391.
16. Yee SL, Myo TT (2020) Study on preliminary phytochemical screening, antibacterial and antioxidant activities of Piper betle L. (betel vine). *J Myan Acad Arts Sci* 18:1B.
17. Sheikh BA, Pari L, Rathinam A, Chandramohan R (2015) Trans-anethole, a terpenoid ameliorates hyperglycemia by regulating key enzymes of carbohydrate metabolism in streptozotocin induced diabetic rats. *Biochimie* 112:57-65.

18. Arambewela LS, Arawwawala LD, Ratnasooriya WD (2005) Antidiabetic activity of aqueous and ethanolic extract of Piper betle. *J Ethanopharmacol* 102(2): 239-45.
19. Ramji N, Ramji N, Iyer R, Chandrasekaran S (2002) Phenolic antibacterials from Piper betle in the prevention of halitosis. *J Ethnophar* 83(1-2):149-152.
20. Sharma RK, Goyal AK, Bhat RA (2013) Antifertility activity of plants extracts on female reproduction: A review. *Int J Pharm Biol Sci* 3(3):493-514.
21. Manilal A, Sujith S, Seghal KG, Selvin J, Shakir C (2009) Cytotoxic potentials of red alga, *Laurencia brandenii* collected from the Indian coast. *Glob J Pharmacol* 3:90-94.
22. Doss A (2009) Preliminary phytochemical screening of some Indian medicinal plants. *Anci Sci Life* 29(2):12.
23. Ayoola GA, Coker HA, Adesegun SA, Adepoju-Bello AA, Obaweya K, Ezennia EC, Atangbayila TO (2008) Phytochemical screening and antioxidant activities of some selected medicinal plants used for malaria therapy in Southwestern Nigeria. *Trop J Pharma Rese* 7(3):1019-24.
24. Savithramma N, Rao ML, Suhrulatha D (2011) Screening of medicinal plants for secondary metabolites. *Middle-East J Sci Rese* 8(3):579-84.
25. Shrestha P, Adhikari S, Lamichhane B, Shrestha BG (2015) Phytochemical screening of the medicinal plants of Nepal. *IOSR J Env Sci Toxi Food Tech* 1(6):11-7.
26. Gul R, Jan SU, Faridullah S, Sherani S, Jahan N (2017) Preliminary phytochemical screening, quantitative analysis of alkaloids, and antioxidant activity of crude plant extracts from *Ephedra intermedia* indigenous to Balochistan. *Sci World J* 13: 2017.
27. Sarah QS, Anny FC, Misbahuddin M (2017) Brine shrimp lethality assay. *Bangladesh J Pharmacol* 12(2):186-9.

28. Mogale MA, Lebelo SL, Thovhogi N, De Freitas AN, Shai LJ (2011) α -Amylase and α -glucosidase inhibitory effects of *Sclerocarya birrea* [(A. Rich.) Hochst.] subspecies *caffra* (Sond) Kokwaro (Anacardiaceae) stem-bark extracts. *African J of Biotec* 10(66):15033-9.
29. Yogeswari S, Bindu KH, Kamalraj S, Ashokkumar V, Jayabaskaran C (2020) Antidiabetic, Antithrombin and Cytotoxic bioactive compounds in five cultivars of Piper betle L. *Env Tech Innov* 20:101140.
30. Abraham NN, Kanthimathi MS, Aziz AA (2012) Piper betle shows antioxidant activities, inhibits MCF-7 cell proliferation and increases activities of catalase and superoxide dismutase. *BMC Comple Altern Med* 12:220.
31. Huang WY, Cai YZ, Zhang Y (2009) Natural phenolic compounds from medicinal herbs and dietary plants: potential use for cancer prevention. *Nutr Can* 62(1):1-20.
32. Vallianou L, Peroulis N, Pantazis P, Hadzopoulou-Cladaras M (2011) Camphene, a plant-derived monoterpene, reduces plasma cholesterol and triglycerides in hyperlipidemic rats independently of HMG-CoA reductase activity. *PLoS One* 6 (11):1–11.
33. Polterait O (1997) Antioxidants and free-radical scavengers of Natural Origin. *Current Org Chem* 1:415-440.
34. Qiu S, Sun H, Zhang AH, Xu HY, Yan GL, Han Y, Wang XJ (2014) Natural alkaloids: basic aspects, biological roles, and future perspectives. *Chin J Nat Med* 12(6):401–406.
35. Murakami A, Ali AM, Mat-Salleh K, Koshimizu K, Ohigashi H (2000) Screening for the in vitro anti-tumor-promoting activities of edible plants from Malaysia. *Biosci Biotech Biochem* 64(1):9-16.

36. Sriwiriyan S, Ninpesh T, Sukpondma Y, Nasomyon T, Graidist P (2014) Cytotoxicity screening of plants of genus *Piper* in breast cancer cell lines. *Trop J Pharma Rese* 13(6):921-8.
37. Uddin MF, Uddin SA, Hossain MD, Manchur MA (2015) Antioxidant, cytotoxic and phytochemical properties of the ethanol extract of *Piper betle* leaf. *Int J Pharma Sci Rese* 6(10):4252-8.
38. Del Socorro MM, Bendoy CP, Dacayana CM (2014) Cytotoxic effects of betel vine, *Piper betle* Linn. leaf extracts using *Artemia salina* leach (Brine Shrimp Lethality Assay). *J Multidi Studies* 3(1).
39. Elumba ZS, Teves FG, Malaluan RM (2013) DNA-binding activity and in vivo cytotoxicity of *Ganoderma applanatum* (Pers.) Pat. supercritical-CO₂ extracts. *African J Micro Rese* 7(3):202-10.
40. Bhide SV, Shivapurkar NM, Gothoskar SV, Ranadive KJ (1979) Carcinogenicity of betel quid ingredients: feeding mice with aqueous extract and the polyphenol fraction of betel nut. *British J Can* 40(6):922-926.
41. Nair SS, Kavrekar V, Mishra A (2013) In vitro studies on alpha amylase and alpha glucosidase inhibitory activities of selected plant extracts. *Euro J Exp Bio* 3(1):128-132.
42. Rajkumar L, Srinivasan N, Balasubramanian K, Govindarajulu P (1991) Increased degradation of dermal collagen in diabetic rats. *Ind J Exp Bio* 29(11): 1081-1083.
43. Santhakumari P, Prakasam A, Pugalendi KV (2006) Antihyperglycemic activity of *Piper betle* leaf on streptozotocin-induced diabetic rats. *J Medi Food* 9(1): 108-112.

44. Radhika K, Kumaravel B, Thamizhiniyan V, Subramanian S (2013) Biochemical evaluation of antidiabetic activity of Piper betel leaves extracts in Aalloxan-induced diabetic rats. Asian J Res Chem 6(1): 76-82.
45. Bischoff HB (1994) Pharmacology of alpha-glucosidase inhibition. Euro J Clin Inves 24:3-10.
46. Bray GA, Greenway FL (1999) Current and potential drugs for treatment of obesity. Endocrine reviews 20(6):805-75.
47. Perumal PA, Saravanabhavan K (2018) Antidiabetic and antioxidant activities of ethanolic extract of Piper betle L. leaves in catfish, Clarias gariepinus. Asian J Pharma Clin Res 11(3):194-8.