# Nutrient Enhancing and Flesh Quality Improvement in Catfish (*Clarias gariepinus*) FedDietarySweetPotato(*Ipomoa butatas*)Leaves AqueousExtract

#### 1. ABSTRACT

Dietary sweet potato(*I. batatas*) leaves extract was assessed for nutrient enhancing ability and improvement of flesh quality in catfish (*C. gariepinus*). Thirty five (35) % crude proteinfeed was formulated using locally available ingredients. Four different diets were prepared from the formulated feed by adding varying quantities of sweet potato leaves extracts as follows: Oml/kg; 50ml/kg; 100ml/kg; and 150ml/kg and labeled as Do, D1, D2 and D3respectively. One hundred and twenty (120) sub-

adult*C.gariepinus* were usedfortheexperiment, they were divided into four groups intriplicates of 10. Feeding with the experimental diets (Do-D3) commenced after two weeks of acclimatization and they were fedfor eight (8) weeks, and water quality parameters such as temperature, dissolve oxygen and pH were determined daily, and measurement of length and weight was done fortnightly. After the feeding period fish were collected from each of the groups for proximate composition analysis and organoleptic assessment. The proximate composition of the diets were done to assess the effects of *I.batatas* on the quality of the diets. The results revealed the following:

- (i) the diets had no effects on the assessed water quality parameters; (ii) there were no significance difference in the proximate composition of the experimental diets; (iii) the *I.batatas* leaveextracts enhanced the lipids, protein and fibre contents on the flesh of *C.gariepinus*; (iv)the *I.batatas* improvestexture, taste, appearance and general acceptability of
- C. gariepinus flesh; (v) nutrient utilization parameters such as protein intake (PI), proteinefficiency ratio (PER), protein retention (PR), fat retention (FR) and net protein retention(NPU) increases significantly as the quantity of *I. batatas* extracts increases in the diet
- D3). It was concluded that sweet potato (I.batatas) leaves extract simproves nutrient sutilization and flesh quality in C. gariepinus by enhancing bioavailability, digestion and absorption of nutrients.

#### Keywords:

NutrientUtilization,OrganolepticAssessment,ProximateCompositionofE xperimentalfishflesh,Proximatecomposition ofexperimentaldiets

#### 2. INTRODUCTION

Fisheries and aquaculture plays significant role in the promotion of food sufficiency, and itcontributes above 15% to the protein consumed by humans especially in the underdeveloped countries of the world (1). One of the problems in aquaculture is the availability of goodquality fish feed and a healthy environment free of diseases. Good quality

feed will boost

the production of fish, enhance growth rates and reduced is ease presence (2,3). The quality of a superior of the production of the prod

fish feed is determined by the quality of nutrients in its ingredients, and how the fish utilizesthese nutrients determines the growth rate and taste of the fish. The growth and taste of the fishis further determined by the bioavailability of nutrients in the fish feed (4).

Oneoftheessentialingredientsinfishfeedproductionisprotein, because of its uniqueroleinthed evelo pment of fish. Protein plays an important role in the growth and health of fish (3). Fish meal is among the desirable protein ingredients because of it's high content of aminoacids, but the decrease in it's supply as a result of demand and cost is putting the sustainability of the aquaculture industry at risk (1). Since the cost of fish meal is increasing each passing day, identifying alternative feeds tuffs to fish meal will enhance productivity in a quaculture (5).

So many authors have reported the importance of plants in the improvement of growth andhealth in fish (6, 7, and 8). (9) reported that sweet potato leaves (*Ipomoa batatas*) contains varying percentages of ash, fat, protein, fibre, carbohydrate etc, and posses some important phytochemicals such as flavonoid, coumanins, sapoinine, tannins, anthraquinnies, alkaloids and phenols. (9) further stated that these components of the *I.batatas* has the capacity to boostgrowth and health in fish culture. The phytochemicals contained in *I. batatas* have the ability to enhances digestibility and absorption of nutrients in fish (10, 11).

African catfish (*Clarias gariepinus*) is one of the most cultured fish outside its environmentbecause of its ability to survive in high stocking density, resist disease and good flesh quality. This research investigated the dietary effects of sweet potato (*I.batatas*) leaves on the nutrient utilization and flesh quality of *Clarias gariepinus*.

#### 3. MATERIALSANDMETHODS

#### 3.1 Experimental Area

The experiment was carried out in the fish farm of the Department of Fisheries and AquaticEnvironment,Rivers StateUniversity, Nigeria.

#### 3.2 ExperimentalFish/Acclimatization

The fish (sub-adult *Clarias gariepinus*) was purchased from a reputable fish farm withinRivers State, and was taken to the experimental area between the hours of 6am – 7am in themorning. The fish was acclimatized for two (2) weeks, observed for disease presence and feeding to satiation was donetwice aday, while waterparameters were monitored.

#### 3.3 Preparation of Experimental Herb/Diets

Sweetpotato(*Ipomoabatatas*) leaves were harvested within Rivers State. It was was hed clean and processed using the methods of (12). The *I. batatas* leaves were pounded to paste and soaked in hot water (50°c) at 500g/L for twelve (12) hours. It was filtered and the filtrate was used immediately.

35% crude protein feed was formulated using locally available ingredients, and four differentiates were produced from the feed by adding varying quantities of the prepared *I. bantatas*extracts as follows: 0ml/kg, 50ml/kg 100ml/kg and 150ml/kg and labeled as Do, D1, D2 and D3 respectively.

#### 3.4 Experimental Designand Feeding Trials

A total of one hundred and twenty (120) sub-adult *Clarias gariepinus* were distributed intofour (4) groups in triplicates of ten (10) fish per replicate into twelve (12) aquariums (10fish/aquarium). The fish were acclimatized in the aquariums for two weeks and were fed tosatiation twice a day with a commercial diet. After the acclimatization period, the fish werefed with the experimental diets (Do - D3) according to their group for a period of eight (8)weeks,andcompletewaterexchangewas done ones daily.

#### 3.5 Proximate Analysis of the Experimental Diets and Experimental Fish

The proximate analysis of the experimental diets and fish were carried out in the DepartmentofFood Scienceand Technologyin theRivers StateUniversity, usingthemethods in

(13).

#### 3.6 Determination of Nutrient Utilization

The following parameters were evaluated to determine the nutrient utilization, using the methods in (14) and (15):

#### - FeedIntake(FI)

$$FI = \frac{Weight of feed consumed(g)}{Number of fish}$$

#### - NetProteinUtilization(NPU)

$$NPU = \frac{FishPr}{otein}_{x100} ProteinF$$

#### FoodConversionRatio(FCR)

$$FCR = \frac{Feed intake(g)}{Weightgain(g)}$$

#### - ProteinIntake(PT)

PI = PercentagecrudeproteinoffeedXFeedConsumed(g)

#### - ProteinEfficiencyRatio(PER)

$$PER = \frac{Weightgain}{(g)Proteinintake(g)}$$

#### - FatRetention(FR)

$$FR = \frac{FFCxFFW(g) - (IFCxIFW(g))}{FCDxFCRx[FFW(g) - IFW(g)]} X100$$

whereFR = Fatretention

FFC = Finalfatconcentration

FFW = Finalfishweight

IFC = Initial fish concentration

IFW = Initialfishweight

FCD = Fatcontentofdiet

FCR = Feedconversionratio

#### ProteinRetention(PR)

$$PR = \frac{FPC - FFW(g) - (IPCxIFW(g))}{PCDxFCRx[FFW(g) - IFW(g)]} X100$$

where PR = Proteinretention

FPC = Finalproteinconcentration

FFC = Finalfishweight.

IPC = Initial protein concentration

IFW = Initialfishweight

FCR = Feedconversion ratio

### 3.7 DeterminationofWaterQualityParameter

The temperature (Temp) and dissolve oxygen (Do) were monitored aily. While the pH was monitored twice a week. They were monitored as follows:

Temperature: Thetemperaturewasdeterminedusingmercuryglass thermometer

PH: ThePHwas determinedusingpHmeter

DO: Thedissolveoxygen wasdeterminedusingtheDometer

#### 3.8 OrganolepticAssessmentoftheExperimental FishFlesh

This was determined using the sense of touch, smell, taste and sight (16). A ten man panel ofjudges were constituted for the assignment. Five fish from the different diets were eviscerated and soak in brime solution for five (5) minutes; they were later dried in an electric fishsmoking oven. At the end of every taste exercise the panelists were given cabin biscuits andwaterto erasethetastebeforetastinganotherset.

## 3.9 StatisticalAnalysis

The data analysis was expressed as a mean + SE for each of the variables. The statistical difference (P<0.05) of the determined values were tested using one way ANOVA. Followed to a turkey multi-comparison test with spss 17.0 packages of tware (17).

#### 4. RESULTS

#### 4.1 PhysicochemicalParametersoftheExperimentalWaters

The results of the physic ochemical parameters is shown in table 1. The rewereno significant difference across the treatments, the values for the tested parameters were similar.

# 4.2 ProximateCompositionofExperimentalDietsandClariasgariepinusFedDietaryIpo mea batatasforEightWeeks

The proximate composition of the experimental diets formulated with different levels of the proximate composition of the experimental diets for mulated with different levels of the proximate composition of the experimental diets for mulated with different levels of the proximate composition of the experimental diets for mulated with different levels of the proximate composition of the experimental diets for mulated with different levels of the proximate composition of the experimental diets for mulated with different levels of the proximate composition of the experimental diets for mulated with different levels of the proximate composition of the experimental diets for mulated with different levels of the proximate composition of

IpomoabatatasarepresentedinTable2. The results obtained indicated that the values for

moisturecontentwaswithinthesamerange(11.30–11.43)betweendietsD<sub>1</sub>-

D<sub>2.</sub>However, alowervalue of  $10.82\pm1.29$  % was recorded in diet D<sub>3</sub>. The same trendwas equally observed in ash, where the same values (16.20-16.92) were recorded between diets D<sub>0</sub> - D<sub>2</sub> and a higher value of  $17.55\pm3.72$  % was recorded in diet across the diets D<sub>3</sub>. The value of crude fiber and lipidwere within the same range. The values for lipidcrude fibre and carbohydratewere higher in Do (6.69±0.29 and 15.08±3.19 respectively).

#### 4.3 ProximateComposition of the Fleshof the Experimental Fish (C. gariepinus)

Theproximatecompositionofthefleshof *Clariasgariepinus* feddietary *Ipomeabatatas* for eightweeks are presented in Table. 3. The results indicated that the values for moisture, crude protein and lipid were higher in the *I. batatas* fed fish (D1-D2) compared to the control (Do). While the values of carbohydrates in the experimental fish varied significantly (P<0.05) among the dietary treatments with no definite pattern. However, the values of ash and crude fibrewere within the same range of 2.36-2.90 and 0.29-0.42 respectively.

 $Table 1: Summary of the Physicochem cial Parameters of the Experimental Waters (Mean \pm SE)$ 

Parameters $D_0$ $D_1$ $D_2$ $D_3$ Dissolveoxygen (mg/L) $4.65\pm0.17$ $4.11\pm0.31$ $4.01\pm0.21$ $3.81\pm0.34$ Temperature(°C) $25.09\pm1.01$ $28.37\pm1.13$ $28.17\pm0.91$ $27.39\pm1.21$		Treatments					
(mg/L)	Parameters	$\mathbf{D_o}$	$\mathbf{D}_1$	$\mathbf{D}_2$	$\mathbf{D}_3$		
Temperature(°C) 25.09±1.01 28.37±1.13 28.17±0.91 27.39±1.21		4.65±0.17	4.11±0.31	4.01±0.21	$3.81 \pm 0.34$		
	Temperature(°C)	25.09±1.01	28.37±1.13	28.17±0.91	27.39±1.21		
pH $6.91\pm1.31$ $6.31\pm0.09$ $6.09\pm0.09$ $6.13\pm1.23$	рН	6.91±1.31	6.31±0.09	6.09±0.09	6.13±1.23		

 $Table 2: Proximate Composition of Composition of Experimental Diets (Mean \pm SD)$ 

Treatments	ProximateParameters(%)					
	Moisture	Moisture Ash CrudeProtein Lipid Crude		CrudeFibre	Carbohydrate	
-						
$\mathrm{D}_0$	$11.30\pm0.71^{b}$	16.86±1.49 <sup>a</sup>	35.15±0.23 <sup>a</sup>	$6.69\pm0.29^{b}$	15.08±3.19 <sup>b</sup>	$29.15 \pm 0.28^{b}$
$D_1$	11.43±0.69 <sup>b</sup>	16.20±0.30 <sup>a</sup>	35.24±0.09 <sup>a</sup>	4.57±0.50 <sup>a</sup>	12.72±2.32 <sup>a</sup>	26.72±0.92 <sup>a</sup>
$D_2$	11.36±0.35 <sup>b</sup>	16.92±0.45 <sup>a</sup>	35.34±0.15 <sup>a</sup>	$5.76\pm0.88^{a}$	12.74±7.78 <sup>a</sup>	25.93±0.56 <sup>a</sup>
$D_3$	10.82±1.29 <sup>a</sup>	17.55±3.72 <sup>b</sup>	35.17±0.01 <sup>a</sup>	4.10±0.06 <sup>a</sup>	11.10±3.48 <sup>a</sup>	26.16±0.24 <sup>a</sup>

 $Table 3: Proximate Composition of C{\it C. gariepinus} Flesh Fed Dietary {\it I. batatas} Leaves Extract (Mean \pm SD)$ 

Treatments	ProximateParameters					
	% Moisture	% Lipids	%Protein	%Carbohydrate	%Ash	% Fibre
BeforeExperiment	58.73±1.50 <sup>a</sup>	4.80±0.45 <sup>a</sup>	14.91±0.59 <sup>a</sup>	0.76±0.14 <sup>a</sup>	2.26±0.16 <sup>a</sup>	0.29±1.31 <sup>a</sup>
$D_0$	71.90±0.04°	$5.82{\pm}1.20^{a}$	$16.52\pm1.22^{b}$	1.02±0.56 <sup>b</sup>	$2.88\pm0.60^{a}$	$0.40\pm1.12^{a}$
$D_1$	71.90±0.08°	$6.81 \pm 1.12^{b}$	17.17±1.30°	0.86±1.91 <sup>a</sup>	2.93±0.43 <sup>a</sup>	$0.41\pm0.35^{a}$
$D_2$	71.61±1.36°	$6.66\pm0.97^{b}$	18.37±0.91 <sup>d</sup>	$0.91\pm0.79^{a}$	2.66±0.93 <sup>a</sup>	$0.42\pm0.77^{a}$
$D_3$	69.29±1.53 <sup>b</sup>	6.35±0.59 <sup>b</sup>	17.89±0.07°	$0.87{\pm}1.23^{a}$	$2.28{\pm}1.15^{a}$	$0.41\pm0.29^{a}$

# ${\bf 4.4} \qquad {\bf Organoleptic Assessment of \it Clarias gariepinus Fed Dietary \it Ipomeabatatas \it Leaf extract sfor Eight Weeks}$

Theorganolepticassessmentof Clarias gariepinus feddietary Ipomeabatatas leafextracts for eight weeks are presented in Table 4. The results revealed a significant (P<0.05) difference in the taste of the experimental fish between the control (D<sub>0</sub>) and other experimental diets of D<sub>1</sub>. D<sub>3</sub>. The aroma of C. gariepinus were within the same range of 7.33-7.67 in the fish fed D<sub>0</sub> – D<sub>2</sub>, while the fish fed D<sub>3</sub> had higher value (8.33  $\pm$  0.58). However, the fish fed diets D<sub>1</sub>-D<sub>3</sub> had higher values (8.33 $\pm$ 0.58). The fish fed diets D<sub>1</sub>-D<sub>3</sub> had higher values (6.00 $\pm$ 1.00–8.33 $\pm$ 1.53) for texture compared to the value in the control (D<sub>0</sub>)(5.00 $\pm$ 1.00). In terms of appearance, the fish fed with dietary treatments of Ipomea batatas of D<sub>1</sub>. D<sub>3</sub> recorded significantly (P<0.05) higher values of 7.33 $\pm$ 1.16, 8.95 $\pm$ 1.16 and 8.67 $\pm$ 0.07 respectively while the fish fed D<sub>0</sub> had 6.67  $\pm$ 1.53. In terms of mouthful and acceptability, C. gariepinus fish fed with dietary treatments of Ipomea batatas of D<sub>1</sub>-D<sub>3</sub> recorded significantly (P<0.05) higher values than the those fed with control diet D<sub>0</sub>, but

# $\textbf{4.5} \qquad \textbf{NutrientUtilizationof} \textit{Clariasgariepinus} \textbf{FedDietary} \textit{Ipomeabatatas} \textbf{LeafextractsforEightWeeks}$

dietD<sub>2</sub>recorded thehighest amongall thedietarytreatments.

Thesummaryofnutrientutilization of Clarias gariepinus feddietary Ipomea batatas for eightweeks are presented in Table 5. The results indicated that the values of protein in take (PI) in the experimental fish fed with different dietary treatments were within the same range, however higher values of  $34.84\pm0.01$ ,  $34.56\pm1.16$  and  $35.33\pm0.09$  were observed in  $D_1, D_2$  and  $D_3$  respectively. In food conversion ratio (FCR), lower values of  $1.21\pm0.07$  and  $1.20\pm0.04$  were observed in  $D_2$  and  $D_3$ . While higher values of  $1.97\pm0.05$  and  $1.50\pm0.09$  were observed in  $D_2$  and  $D_3$ 

 $Table. 4: Organoleptic Assessment of \textit{Clarias gariepinus} Fed Dietary \textit{Ipomeabatatas} Leaf extracts for Eight Weeks (Mean \pm SD)$ 

Treatments			OrganolepticParan	neters		
	<b>Taste</b> (10)	Aroma (10)	Texture (10)	Appearance (10)	MouthFull (10)	Acceptability (10)
$D_0$	6.67±2.08 <sup>a</sup>	7.67±1.53 <sup>a</sup>	5.00±1.00 <sup>a</sup>	6.67±1.53 <sup>a</sup>	7.07±2.08 <sup>a</sup>	34.33±7.09 <sup>a</sup>
$D_1$	$8.00\pm0.00^{b}$	7.33±0.37 <sup>a</sup>	7.33±1.33 <sup>b</sup>	7.33±1.16 <sup>a</sup>	$7.67\pm0.58^{a}$	38.00±0.00 <sup>a</sup>
$D_2$	8.67±1.53 <sup>b</sup>	$7.33\pm2.08^{a}$	8.33±1.53 <sup>b</sup>	8.95±1.16 <sup>b</sup>	$9.00\pm1.00^{b}$	41.00±4.36 <sup>b</sup>
$D_3$	$7.67\pm1.53^{b}$	8.33±0.58 <sup>b</sup>	$6.00\pm1.00^{a}$	8.67±0.07 <sup>b</sup>	$8.00\pm0.00^{b}$	39.33±2.31 <sup>a</sup>

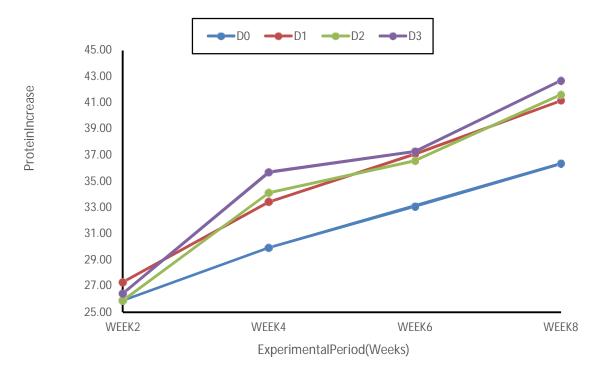
 $Table 5: Summary for Nutrient Utilization of {\it Clarias gariepinus} Fed Dietary {\it Ipomeabatatas}$ 

for Eight Weeks (Mean

	$\pm SE)$ NutrientUtilizationParameters								
	FW	WG	FI	PI	FCR	PER	NPU(%)	PR(%)	FR(%)
$D_0$	$231.01\pm3.50^{a}$	$54.00\pm2.75^{a}$	89.46±2.03 <sup>a</sup>	31.32±0.72 <sup>a</sup>	$1.97 \pm 0.05^{b}$	$1.85\pm0.36^{a}$	46.99±0.92 <sup>a</sup>	31.62±7.97 <sup>a</sup>	16.47±7.41 <sup>a</sup>
$D_1$	263.65±3.90 <sup>b</sup>	74.65±4.90 <sup>b</sup>	99.53±0.93 <sup>a</sup>	34.84±0.33 <sup>a</sup>	1.50±0.09 <sup>b</sup>	2.07±0.11 <sup>b</sup>	48.72±0.72 <sup>a</sup>	43.48±1.82 <sup>b</sup>	53.00±35.02 <sup>b</sup>
$D_2$	259.87±0.87 <sup>b</sup>	84.37±4.13°	98.72±0.02 <sup>a</sup>	34.56±0.01 <sup>a</sup>	1.21±0.07 <sup>a</sup>	$2.41\pm0.12^{b}$	50.98±2.43 <sup>b</sup>	59.85±11.56 <sup>c</sup>	68.47±27.20°
$D_3$	265.12±0.38 <sup>b</sup>	87.62±2.13°	102.34±1.22 <sup>b</sup>	35.53±0.09 <sup>b</sup>	1.20±0.04 <sup>a</sup>	$2.45\pm0.10^{b}$	$50.87 \pm 0.76^{b}$	55.93±3.48°	62.56±38.98 <sup>c</sup>

For protein efficiency ratio of the experimental fish fed with different dietary treatments were withinthe same range of 2.07-2.45 between diets D<sub>1</sub> to D<sub>3</sub>. However, a higher value of 1.85±0.36 were recorded at diet D<sub>0</sub>. In Apparent Net Protein Utilization (ANPU) in C. gariepinus fed with different levels of *Ipomea batatas* inclusion dietary treatments over eight weeks, the values of ANPU obtainedwere within the same range of 7.14-8.11 in diets D<sub>0</sub>, D<sub>2</sub>, andD<sub>3</sub>. However, a higher value of 13.14±2.43 were recorded at diet D<sub>2</sub>. The values of protein retention (PR) obtained in the experimental fish varied significantly (P<0.05) among the dietary treatments, with the highest value(59.85±11.56) observed in diet D<sub>2</sub>. And the lowest (31.62±7.97) observed in diet D<sub>0</sub>. Also, the values of FR obtained in the experimental fish varied significantly (P < 0.05)among the dietary treatments, with the highest value (68.47±27.20) observed in diet D<sub>2</sub>, while the lowest (16.47±7.41) was obser vedin diet  $D_{0...}$ 

Comparative values of protein increase (PI) in C.gariepinus fed with fed with different levels of Ipomeabatatas inclusion dietary treatments over eight weeks is shown in Figure 1. The values of PI increase das the experimental period increased. In all dietary treatments. The highest value of 42.70 obtained in diet  $D_3$  at week 8. While the lowest (25.90) was observed at diet  $D_0$  at week 2. Comparatively, the values of FCR in C.gariepinus fed with fed with different levels of Ipomea Ipome



 $\label{lem:comparative} Figure 1: Comparative values of Protein Increase in \emph{C.gariepinus} \\ fed with dietary \emph{Ipomeabatatas} leaf extracts for eight weeks$ 

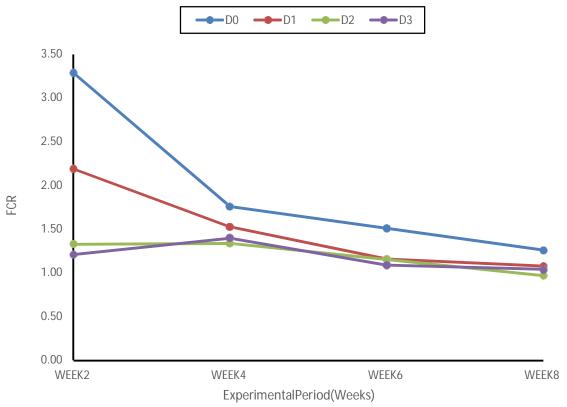
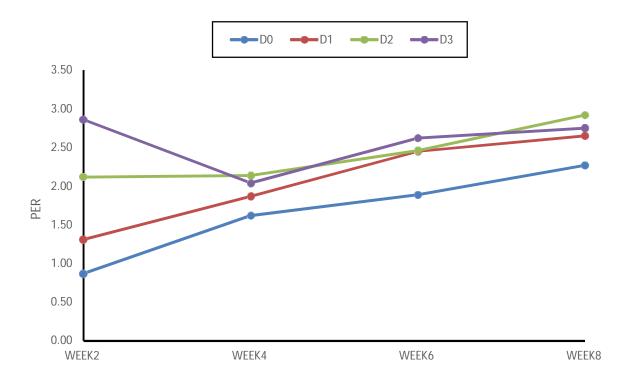


Figure 2: Comparative values of Feed Conversion Ratio in *C. gariepinus* fedwithdietary *Ipomeabatatas* leaf extracts for eightweeks



ExperimentalPeriod(Weeks)
Figure 3: Comparative of Protein Efficiency Ratio (PER) in
C. gariepinus fedwith dietary I pomeabatatas leaf extracts for eight weeks

 $The comparative values of feed intake (FI) in \emph{C. gariepinus} fed with different levels of \emph{Ipomea} \\ batatas in clusion dietary treatments over eight weeks is shown in Figure 4. The values of FI \\ increased as the experimental period increased, in all dietary treatments. The highest value of \\ 122.00 obtained in diet D_3 at week 8. While the lowest (75.3) was observed at diet D_0 at week 2. \\ However, dietary treatment D_0 consistently recorded the lowest value among all the dietary treatments in all experimental periods.$ 

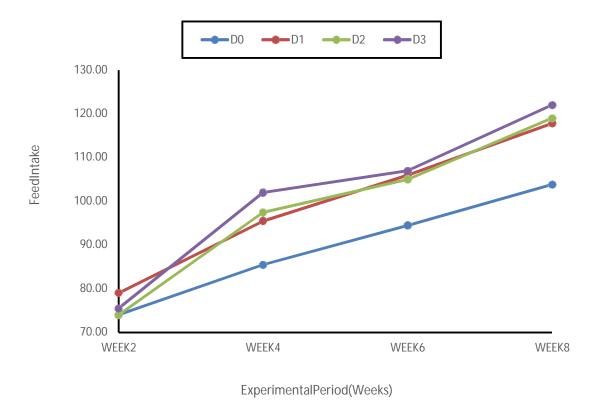


Figure.4:ComparativevaluesofFeedintakein *C. gariepinus* fedwith dietary *Ipomeabatatas* leafextracts for eight weeks

#### **DISCUSSION**

## 5.1 ProximateCompositionofExperimentalDiets

Theresults for the physiochemical parameters (table 1) were similar to that of (9), who stated that they were conducive for aquaculture practice. (18) reported that water qualities such astemperature, dissolve oxygen and pH determines to a large extend the growth and health of fishin a quacu lture. The growth, health and reproduction of commercial fish and other aquaticanimals are primarily dependent upon an adequate supply of nutrient, both in terms of quantity and quality, irrespective of the culture system in which they are grown. Therefore, supply ofinputs(feeds, fertilizersetc.) must be ensured so that the nutrients and energy requirements of the species under cultivation are met and the production goals of the system are achieved (19). Nowadays formulated and commercial fish feeds are widely used formore yield in aquaculture. The protein requirement of commercial fish is influenced by various factors such as commercialfish size, water temperature, feeding rate, availability and quality of natural foods and overalldigestible energy content of diet (20). In this study, the crude protein content of the experimentaldietsanalysedwerewithinthe acceptable range recommended for commercial fish(21).(22) reported that most of the commercial fish feeds for example catfish feeds contain 32% crude protein.(23) estimated the protein requirement for tropical catfish to be 35-40, 25-35 and 28-32% for fry, grow-out and broodstock respectively. However (24) observed that fish production increased through the utilization of high amounts of protein i.e., 35% and above intheirdietanddietaryproteinhavebeenreportedtoimprovethequalityoffishflesh(25).

Lipids are primarily included in formulated diet to maximize their protein sparing effect (26) bybeingasourceofenergy. The observed lipid values were in line with that of (23) who reported

that 10-20% of lipid in most freshwater fish diets gives optimal growth rates without producingan excessively fatty carcass. On the other hand, (27) reported that lipid level in catfish feedsshouldbe5to6%. Moreover, (28) and (29), also stated that dietary lipid levels of 5to6% are often used in tilapia diet. In this study, the lipid content of the experimental diets  $(D_1 - D_3)$  werelower, this may be due to the fact that *Ipomea batatas* leaves extract is low in fat content (30), it could also be that the phytochemicals in *Ipomoa batatas* leaves facilitated the reduction of the lipids in the diets.

All plant ingredients contain a certain amount of fibre, and fibre provides physical bulk to thefeeds. Adequate quantity offibre in feed permits better binding and moderates the passage offeed through the alimentary canal. However, (31) noted that it was not desirable to have a fibrecontent above 8-15% in diets for animals, as the increase in fibre content would consequently result in the decrease of the quality of nutrient in the diet. (32) also stated that high fibre contentin feeds tends to increase the energy content in the feeds, with the resultant effects of poorgrowth. The analysed crude fibre content of all the diets under study were within the safe dietary limit for fish.

#### 5.2 ProximateCompositionofExperimentalFishFlesh

The moisture and carbohydrate content of the experimental fish fed D1 – D3 after the feedingtrial were elevated above the control values. While the ash and fibre content of the fish werewithin same range across the diets. This result agrees with the findings of (33) in the flesh of *C.gariepinus* fed with cassava leaves. The result of the proximate composition of the experimentalfish shows higher protein and fat values in fish fed diets  $D_1$ -  $D_3$  compared to the fish fed Do andthevaluesbeforethecommencementoftheexperiment. Similar result was recorded in (14)

when sea bass (*Dicentrarchus labrak*) was fed dietary Thyme, but Rosemary and Fenugreek in the same experiment showed no significant difference in the protein content of the experiment fish, and (34) who reported improvement in the protein content and reduction in the fat contentwhen *Ipomoea batatas* leaf meal was administrated to *Tilapia Zilli*. (35) also reported increase inboth protein and fat content when African catfish was fed with 40% *Ipomoea batatas* leaf mealfor fourteen days. The deviation in the fat content of *Tilapia Zilli* sited above compare to thepresent result could be as a result of the life stage of the fish. The increase in the protein and fat content of the fish fed D1-D3 could be as a result of the bio-active compounds and mineralscontain in *Ipomoea batatas* leaves extracts (36) that enhanced the digestion and absorption of theprotein and fat in the diets. Minerals such as calcium, phosphorous, potassium etc contain in *Ipomoea batatas* (36) are known to improve fish flesh quality (37, 38). (39) reported the positive effects of plant phytochemicals such as astaxanthin and carotene on the flesh quality of Atlanticsalmon.

#### 5.3 OrganolepticAssessmentsoftheExperimentalFish

Fishes are great source of highly valuable protein that is premium in human nutrition (40), withits irregular water and fat content (41). The crude protein content of the fish is also affected bythe quantity of salt and water-soluble protein in the diets (42), and the presence of endogenous enzymeand bacteria influencing deterioration during the processing period (43).

Eating healthy is a prerequisite to a good and sustainable live, and people are more concern withwhat they eat (44). One factor that influences organoleptic assessment in fish is acceptability of available feed by the fish. (45) postulated that the overall acceptability and sensory characteristic of the fish is correlated to the quality of the water body, and (46)also  $reported difference in taste when {\it Sardinellaspp} and {\it M. pontasson} from different locations within the$ 

South-WestNigeriawereorganolepticallyaccessed.Despitethatseveralmethodshavebeenused to evaluate the flesh quality and freshness of fish, sensory evaluation remains the valuableand trusted means of obtaining the best result (47).(48) reported a positive correlation betweenbody composition and sensory quality, whereas (49) observed no significant differences in thesensory evaluation of fresh fish fed different diets. In this study, fish fed diets D1-D3 were significantly superior to those fed the control diets (Do) in the organoleptic assessment.

The superiority of the fish fed D1-D3 in the organoleptic assessment could be as a result of thebio-active compounds present in the diets, that enhanced bioavailiability of the nutrients in thediets. This assertion is supported by (4) who postulated that the quantity of nutrient delivered to the blood steam for use is more relevant than the quantity present in the feed. Some of thebioactive compounds found in *Ipomoea batatas* leaves includes phenolic compounds, flavonoids, carotenoids, dietaryfibres, dietaryproteinetc(50;51;9) and essential minerals and trace elem ents such as iron, calcium, zinc, copper among others (52; 51). *Ipomoea batatas* leaves have been proven to have high digestibility for proteins/amino acids (11) and digestibility enhances absorption of nutrients (10). (52) also reported that herbs stimulate the secretion of pancreatic enzymes which facilitates nutrients digestion and assimilation.

The noticeable changes in the catfish aroma, colour and taste are the main criteria that qualifycatfish at table size (53). It was demonstrated in this study that feeding catfish with differentlevels of *Ipomea batatas* leaves inclusion diet enhanced the typical characteristics of fish; taste, aroma, texture, and appearance in catfish flesh and did not yield any off odour or flavour, and this can be attributed to the higher lipid content in the fish fed D1-D3 (Table.3).

The observation was similar to previous reports which suggested that the lipid in fish flesh affects these need flavour and the general sensation of cooked fish in the mouth as well as a roma (53). (54)

also noticed the effect of inclusion of the supplemental plant-based protein in the diet of browntrout on taste, texture, and acceptability. On the contrary, (55) observed that no effects of dietarytreatments were found to affect taste, texture and aroma of the Indian major carps fed withdifferentplant baseddietarytreatments.

## 5.4 NutrientUtilizationoftheExperimentalDiets

Plants and plants products have been utilized as additive or supplements in fish feeds due to theirability in the maintenance of fish health andenhancing digestibility/absorption of nutrients infeed (56; 10). Plants and plants products are preferred to synthetic drugs as growth enhancers inaquaculture (57), and have been proved to be growth promoters, anti-bacteria, environmentallyfriendly and not immunospecific (58). Some of the bioactive compounds in plants and plantproducts that facilitates the above qualities includes; polyphenols, flavonoids, saponines,

tannins, essentialoils etc. (8,9), and *Ipomoeabatatas* leaves extracts contains these bioactive compounds (50; 51; 9).

Some of the valuable indices used to determine the effectiveness of how an experimental fishutilizes its diet are: the feed conversion ratio (FCR) which is the expression of how the fishconverts feed to flesh; the protein efficiency ratio (PER) which expresses theeffectiveness ofthefishtoutilizeproteininthedietforgrowth; the proteinintake (PI) which states the quantity of protein taken from the injected feed by the fish, and the feed intake (FI) which expresses the quantity of feedinjected by the fish, and the feed intake (FI) which expresses the quantity of feedinjected by the fish, and 32).

There were difference in the PI, FCR, and PER in the fish fed Do – D3, with the value increasing the concentration of *Ipomoea batatas* leaves extracts in the diets increases. The result of this research shows that the PI and FI increases as the period of feeding increases in fish

 $fed\ D1-D3 compared to the fish fed\ Do. This result is similar to the report of (6) when \textit{Clarias gariepinus}$ 

was fed dietary *Terminalia catappa*, *Chromolaena odorata* and *Psidium guajava*. The increase inthe feed intake (FI) and protein intake (PI) in the fish fed D1-D3 could be as a result of the the presence of the bioactive compounds in *Ipomoeabatatas* leaves extract, that enhanced the palatability of the dietsor having direct bacteric idal effects on the digestive system of the the the contract of the dietsor having direct bacteric idal effects on the digestive system of the dietsor having direct bacteric idal effects on the digestive system of the dietsor having direct bacteric idal effects on the digestive system of the dietsor having direct bacteric idal effects on the digestive system of the dietsor having direct bacteric idal effects on the digestive system of the digestive system

D3therebyenhancingproteindigestion(61).Polyphenols,flavonoids,calcium,phosphorus, and potassium are some bioactive compounds and minerals found in *Ipomoa batatas* and have the ability to enhance palatability and digestion of feed (50; 62). The feed conversionratio(FCR)reducedastheperiodoftheexperimentincreases and were lower in the fishfed D1-D3 compared to the fishfed D0 (Fig. 2). Feed utilization by the fish determines the FCR (63).

The reduction in the FCR is an indication that the fish made a good conversion of feed to flesh, and the reduced values in D1-D3 depicts that the biochemical compounds in *Ipomoea* batatasenhanceddigestion and absorption of the feed compared to fish fed Do (11, 10).

Theproteinefficiencyratio(PER)increasesastheperiodoftheexperimentincreasesinthefish fedD1-D3showingbetterperformance(Fig..3). Proteinefficiencyratioistheabilityofthe fish to use the protein absorbed from the ingredients for growth. The increase in the PER in fishfed D1-D3 depicts the fact that there were enhance digestion and absorption of the proteincomponentsofthedietscomparedtothefishfedDo.Thiscouldbeasaresultofthephytochemicals in *I. batatas* (9), which have been reported to enhance protein digestiability andabsorption in fish (11). Other authors have reported results similar to these findings: (64) whenstriped catfish was fed dietary ginger and (6) when African catfish was fed dietary *Terminaliacatappa*.

Aftertheeight(8) weeksfeeding period, the NPU, PR and FR increased with increase in *Ipomoeabatatas* inclusion in the diet, with significant increase in PR and FR (Table 5). The

increase of these parameters (NPU, PR, and FR) in the fish fed D1-D3 compared to Do suggest the fact that the *Ipomoea batatas* enhanced nutrient utilization, and it is as a result of thestimulating effect of *Ipomoea batatas* leaves extracts on the secretion of pancreatic enzymes that facilitates digestion and absorption of nutrients (52).

Though the PR and FR were higher in the fish fed D1 – D3 the values for PR were highercompared to FR as the *I. batatas* leaves extract increases in their diet ( $D_0 - D_3$ ). This depicts the fact that more fat was utilized for energy while more protein was utilized for growth, and as are sult led to increase in the quantity of protein deposited in the fish flesh (NPU) which is ameasure of digestibility. This position is supported by (14) and (65).

#### 6. Conclusion

The present study shows that application of *Ipomea batatas* leaf extract can be utilized in fishfeed for optimal performance of fish, as it enhances bioavailability, digestion and absorption of nutrients. It was demonstrated that feeding catfish with different levels of *Ipomea batatas* leafextract inclusion diet improved the typical characteristic of fish such as flesh color, flavour andaroma in catfish fillet and did not yield any off odour, which connotes the fact that what the fishconsume as feed affects the quality of the fish flesh. The result of the study shows that 100 – 150ml/kg dietary inclusion level of *Ipomea batatas* leaves extract produced fish of better fleshqualityandcompositionthanthecontrol.

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# **REFERENCES**

1.	Ayoola, A.A. (2010). Replacement of Fishmeal with Alternative Proteins our ces in Aquacultu re Diets. Athesis submitted to the radiate Faculty of North Carolina State University in Partial fulfilment of the requirements for the Degree of Master of Science
2.	Alatise, P. S., Ogundele, O, Eyo, A. A. & Oludunjoye, F. (2006). Evaluation of Different Soybean Base Diets on Growth and Nutrient Utilization of Heterebrancheslongifilis in Aquaria Thanks. Proceedings of the Fisheries society of Nigeria (FISON)Conference held at the University of Calabar, Calabar, Nigeria, 13-17 November, 13-17.
3.	Effiong, M. U., Akpan, A. W., Essien-Ibok, M. A. (2019). Effects of Dietary Proteinlevelsonproximate, Haematological and lenthocytecompositions of <i>Clarias gariepi</i> nus. <i>Journal of applied Science and Environmental Management</i> , 23(11):2065-2069.
4.	Parada, J& Aguilera, J.M (2007). Foodmicrostructure affects the Bioavailability of several nutrients. <i>Journal of Food Science</i> . 72(2); R21-R32.
5.	Idowu, E. O. & Afolayan, E. B. (2013). The Effects of Supplementing of FishmealwithMaggotsatVaryinglevelsintheDietofClariasgariepinus.InternationalArch iveof appliedScienceandTechnology,4(4):41-47.
6.	Lawal, M. O., Ademole, Z., Aderolu & Wahab, A. G. (2021). Effects of Terminaliacatopa, chiomolgenaodorata and psidium guaja valeaf Extracts on Growth. Bioch emical and Haematology of Clarias gariepinus. FUW Trends in Science of Technology, 6(2): 327 – 332
7.	Abu, O.M.G; Ukwe, I.O.K & Audu, S. (2023) Therapeutic effects of Zea Mays HusksextractsonthebehaviouralandHaematologicalalterationsof <i>Pseudomonasaerugino sa</i> infected <i>Clariasgariepinus.SumeriansJournalsofAgricultureandVeterinary.</i> 6(2): 12-18
8.	Ukwe,I.O.K.&Deekae,S.N.(2022).PhytochemicalAssessmentof <i>PerseaAmericana</i> Powered Leaves and its Potency in Protecting <i>Claris gariepinus</i> against <i>Klebsiellapneumonea.AsianJournalofFisheriesandAquaticResearch</i> ,16(6):1-9.
9.	Ukwe, I.O.K & Deekae, S.N (2024) Phytochemical and Proximate Analysis of sweetpotato(Ipomoabatatas)leavesaqueousextractanditsprophylaticeffectsonPseudomo nasaeruginosainfectedcatfish(Clariasgariepinus). Asian Journal of Aquatic Research. 26(6):76-87
10.	Koushik,R.&Mraz,J.(2021).Digestibility <a href="https://dor/1">https://dor/1</a> of protein feeds in Tilapia. <a href="https://dor/1">0.1314/RG</a> .2.2.25-455.36006.
11.	Leon,N. (2023). Evaluation of locally Available Feed Resources for Nile tilapia ( <i>Oreochromis noliticus</i> ) in Rwanda. PhD Thesis. Faculty of Veterinary Medicine and animal Science, Animal Nutrition and Management Uppsala Sweden. Swedish University yof Agricultural Science. <i>Acta Universitatis Agriculture Sueclae</i> .

12.	Ukwe, I.O.K. & Jamabo, N. A. (2020). Effect of dietary mango bark (Mangniferaindicia) extracton Clarias gariepinus (Burchell, 1822) infected with Pseudomo naaeruginosa. World Journal offishand Marine Science, 12(3):74-80.
13.	Oh,H.Y.,Lee,T.H.,Lee,D-Y.,Lee,C-H.,Joo,M-S.,Kim,H.S.,Kim,K-D.(2022). Dietary supplementation with ginger ( <i>Zingiber officinale</i> residue from jucie extractionimprovesjuvenileblackrockfish( <i>Sebastesschlegelii</i> )growthperformance,antio xidant enzyme activity and resistance to strepotococcus iniae- infection. <i>Animals</i> 12(5).https://doi.org/10.3390/ani/2050546.
14.	Yilmaz, S., Ergun., S., Celik, E.S. (2012). Effects of herbal supplements on growthperformance of sea bass ( <i>Dicentrarchus labrax</i> ): change in body composition and some blood paramters. <i>Journal of Bioscience and Biotechnology</i> , 1(3), 217-222.
15.	Ukwe,I.O.K,Amachree,D&Jamabo,N.A(2019).GrowthAssessmentandMicrobialFlorap resenceinAfricancatfish( <i>C.gariepinus</i> )larvaefedliveandcommercialfeeds. <i>International Journal of Science</i> .8(7): 1-6.
16.	Lubis, A.S, ZaKaria, I.J&Efrizal (2021), Organoleptic, physical and chemical tests of Formula ted Feedfor Panulirushomarus, enriched with spinach extract. <i>AACL Bioflux</i> : 14(2):866-873
17.	Wahua, T.A.T (1999). Applied statistics for scientific studies. A fricanlink books. Aba, Nigeria 365pp.
18.	Ukwe, I.O.K. & Abu, O. M. G. (2016). Physico-Chemical Parameters Of Water InHoldingTanksOf <i>ClariasGariepinus</i> InducedWithOvaprimaAndOvalinHormoness. <i>Int ernationalNationalJournalofInnovativeStudiesinAquaticBiology andFisheries</i> .2(4):12-19
19.	Gabriel, U.U., O.A; Akinrotimi, D.O. Bekibele, D.N. Onunkwu& P.E. Anyanwu(2007). Locally produced fish feed, potentials for aquaculture development in sub-saharan <i>African Journal of Agricultural Research</i> , 297, 287-295.
20.	Storebakken, T. & Refstie, S. (2000). "Vegetable Proteins for Carnivorous Fish". <i>Aqua2000InternationalConference</i> , <i>ResponsibleAquacultureintheNewMillenniu m</i> , May2-6,2000, Nice, France, p. 682.
21.	Shiau, S., Lin, S., Yu, S., Lin, A. & Kwok, C. (1990). "Defatted and Full – FatSoybeanMealasPartialReplacementsforFishMealinTilapia( <i>OreochromisniloticusX O.aureus</i> )DietsatLowProteinLevel". <i>Aquaculture</i> 86:401-407.
22.	Soliman, A. K. (2015). "Aspects of AscorbicAcid (Vitamin C) Nutrition in <i>O. niloticus</i> and <i>O. mossambicus</i> ". Ph.D. Thesis, Institute of Aquaculture, University of Stirling, Scotland.
23.	Agani, E., Nwanna, L. & Musa, B. (2004). Replacement of Fishmeal with Maggot meal indiets of Tilapia <i>Oreochemeomis niloticus</i> . <i>Worlda quaculture</i> , 35:52-54.

24.	Atteh, J. & Ologbenla, F. (2015). Replacement of fish meal with maggots in broilerdiets:effectsonperformanceandnutrientretention. <i>Nigerian Journal of Animal Production</i> , 20,44-49.
25.	Kuo,H;Hu,J:Liu,X"Zhao,L;Zhang,K;Pan,X;Wang,A;Miao,Y&LinL(2022). Dietary protein improves flesh quality enhancing antioxidant ability via theNF-EZ- related factor 2/Kekh – like ECH – associated protein 1 signalling pathway insoftshell turtle( <i>PelodiscusSinensis</i> )Frontiers inNutrition.9: 1030583.
26.	Hassan, M.R. (2001). Nutrition and feeding forsustainable aquaculturedevelopment in the third millennium. Technical proceedings of the conference on aquaculture in the third millennium J.R. Arthur eds, M.J. Phillips publishers, Bongkok, Thailand. Pp193-219.
27.	Wilson, R.P. (2000). Channel catfish, Ictalurus punctatus. In: Handbook of Nutrient Requirem entoffin fish Wilson, R.P. (Ed). CRCPress, Boca Raton, USA., pp:35-53.
28	Atack, T., Jauncey, K. & Matty, A. (2019). The utilization of some singlecell proteins by fingerling mirror carp ( <i>Cyprinus carpio</i> ). <i>Aquaculture</i> , 18,337-348.
29.	Luquet,P.(2000).TilapiaOreochromisspecies.In:Handbookofnutrientrequirementoffinfi sh.Wilson,R.P.(Ed).CRC press,BocaRaton,FL.,USA.,PP:169-180.
30.	Oyin O. (2006). Nutritive potential of sweet potato meal and root replacement value for maize in Diets of African Catfish ( <i>Clarias gariepinus</i> ) advanced fry. <i>Journal ofFeedTechnology</i> .20-22pp.
31.	Awoniyi, D.O., Aboua YG, Marnewick JL, Du Plesis S.S, & Brooks NL. (2011)Protective effects of rooibos ( <i>Aspalathus Linearis</i> ), green tea ( <i>Camellia sinensis</i> ) and commercial supplements on testicular tissue of oxidative stress-induced rats. <i>African Journal Biotechnol</i> 10:17317-1732
32.	Ukwe,I. O. K., Gabriel, U. U. & Deekae, S. W. (2020). Assessment of DietaryPowderedAvocadoPear( <i>PerseaAmerican</i> )leavesonGrowthPerformanceandSurv ival of African catfish. ( <i>C. gariepinus</i> ). <i>Global scientific Journals</i> , 8(8): 1612-1626.
33.	Oresegun A,. Alegbeleye W.O. (2001). Growth response and nutrient utilization oftilapia ( <i>Oreochromis niloticus</i> ) fed varying Dietary levels of cassava peels based onrations supplemented with dimethionine. Fish Nutrition and Fish Feed Technology inNigeria,pp: 38-44.
34.	Adewolu, M.A. (2008) Potentials of sweet potato (I. batatas) leaf meal as dietarying redients for Tilapia zillifingerlings. Parkinstan Journal of Nutrition. 7(3):444-449.

35.	Oludayo, O.C (2010). Growth Performance and blood profile of African catfish fedsweetpotatoleafmeal5 <sup>th</sup> InternationalSeminaronTropicalAnimalProductionCommunityEmpowermentandTropicalAnimalIndustry.Yogyakarta,Indonesia.
36.	Olamiposi, O. O. & Tolulope, G. O. (2018). Potentials of Sweet potatoes ( <i>IpomeaBatata</i> ) as a mineral and grown supplement in diets of hybride catfish (Heteroclaria) fingerlings. <i>Journal of Entomology and Zoology Studies</i> 6(4): 300–304.
37.	Wu,J.H.,Li,Z.P.,Wei,H.M.Zhang,J.,L,Y.C.,Zhuo,Z.S.,Zhon,Y.L.,Araghi, S. I. & Li, G. H. (2017). Dietary carotenoid availability antioxidant status of pacificwhiting (erluccius productus) fed diets with different carotenoids sources. <i>Journal offoodCompositionAnalysis</i> .7(1),122-130.
38.	Bell,J.P.,Jeyaloganathan,F.R.,Moretti,P.A.,Hale,C.A.&Schilling,M.E.(2013). Effect of dietary calcium, phosphorus and potassium on muscle water holdingcapacity and text of cooked pacific whiting (merluccius productus). Journal of foodChemistry. 101:439–446.
39.	Overland, M. A., Berge, T., Hemre, K. M., Sappola, H., M., Sorensen, M., Williamson, R. & Bell, S. C. (2008). Impact of dietary astaxanthin, beta – carotene and lutein onflestqualityofAtlanticSalmon(SalmoSola). <i>JournalofAgriculturalandfoodChemistry</i> . 56:5012–5020.
40.	Nargis, A. (2006). Seasonal variation in the chemical composition of body flesh of kiofish Anabaste studineus (Bloch) (Anabantidae: percifomes). Bangladesh Journal Science Industry Research, 41 (3-40, 219-226
41.	Pal. J, Shukla BN, Maurya AK. & Verma, H.O. (2018). A review on role of fish inhuman nutrition with special emphasis fatty acid. <i>International Journal of FisheriesandAquaticStudies</i> .2018; 6(2):427-430.
42.	Chomnawang, C., Nantachai, K., Yongsawatdigul, J., Thawornchinsombut, S., & Tungkwac harara, S. (2007). Chemical and Biochemical changes in hybrid catfish filletstoredat 4oCandits gelproperties, <i>FoodChemistry</i> , 103, 420-427.
43.	Hultman, L., Rustard, T. (2004). Iced storage of Atlantic salmon (Salmo salar) effectsonendogenousenzymesandtheirimpactonmuscleproteinsandtexture. <i>FoodChemis try</i> , 87,31-34.
44.	Oriakpono,O.,Frank-Peterside,N.,Ndome,C.(2011).MicrobiologicalAssessmentofStored <i>Tilapiaguineensis.Af ricanJournalofFoodScience</i> ,5(4),242-247.
45.	Farmer, L.J., J.M. Mcconnell & D.J. Kilpartrick. (2000). Sensory Characteristics of farmed and wild Atlantic Salmon. <i>Aquaculture</i> . 187: 105-125.
46.	Fawole,O.O.,Oyelese,O.A&Etim,E.U.(2018).OrganolepticandchemicalAssessment of Two frozen marine fishes obtained from markets in four AgriculturalZonesofOyoState,Nigeria. <i>IfeJournalof Science</i> ,20(2): 39-343.

47.	FatmaHassan,&MohamedAli(2011).Qualityevaluationofsomefreshandimported frozen seafood. <i>Advance Journal of Food Science and Technology</i> . 3(1), 83-88.
48.	Shiau, S., Lin, S., Yu, S., Lin, A. & Kwok, C. (1990). "Defatted and Full – FatSoybeanMealasPartialReplacementsforFishMealinTilapia( <i>OreochromisniloticusX O.aureus</i> )DietsatLowProteinLevel". <i>Aquaculture</i> 86:401-407.
49.	Ochang, S.N, O.A. Fagbenro, & O. Adebayo. (2007). Growth Performance, BodyComposition, Hematology, and Product Quality of <i>Clarias gariepinus</i> Fed Diets withpalmoil. <i>Pakistan Journal f Nutrition</i> , 1:452-459.
50.	Nguyen,H.C.,Chen,C.C.,Lin,K.H.,Chao,P.Y.Lim,H.H.&Haung,M.Y.(2021).BroactiveC ompounds,AntioxidantsandHealth BenefitsofSweetpotato leave. <i>Molecules</i> ,26(7);19820.https;//des.org/103390/molecules26071820
51.	Awol, A. (2014). Phytochemical Screening Proximate and Mineral Composition of Sweet Pot ato Leaves Grown In Tepi Provision, South-west of Ethiopia. <i>Science</i> , <i>Technology and Arts Research Journal</i> , 3(3):112. <a href="http://doi.org/10.4314/star.v3i3.19">http://doi.org/10.4314/star.v3i3.19</a>
52.	Frankic, T., Voljc, M., Salobir, J. & Rezar, V. (2009). Use of herbs and species and their extracts in an imal nutrition. ( <i>ActaUniversitatisAgricultureSueclae</i> ), 94,95-102.
53.	Muin, H., Fatah, N.N.A., Bahari, I.H. and Razak, S.A. (2014). Replacement of ricebran with Pleurotus florida stalks on growth performance of Oreochromis niloticusfingerlings. <i>Sains Malaysiana</i> , 43(5),675-681.
54.	Ramezani, H. (2009). Effects of different protein and energy levels on growth performance of Caspian Brown Trout, Salmotrutta capinus (Kessler, 1877). <i>Journal of Aquatic Science</i> . 4(4), 203-209.
55.	Khan,N.,Qureshi,N.&Nasir,M.,Vandenberg,G.,Mughal,M.,Maqbool,A.,Jabbar,M.&Zi kria,N.(2012).EffectofartificialfeedonsensoryattributesoffleshofIndianmajorcarps( <i>Labe orohita</i> , <i>Catlacatla</i> and <i>Cirrhinusmrigala</i> ) fedinmonocultureandpolyculturesystems. <i>Paki stanVeterinaryJournal</i> , 32,349 -353.
56.	Dawood,M.A.O.,ElBasuini,Yilmaz,S.,Abdel-Latif,H.M.R.,Alagawany,M.,Kari, Z. A., Abdul Razab M.K., Hamidd, N.K., Moonmanee, T. & Van Doan, H. (2022).Exploring the roles of dietary herbal essential oils in aquaculture: a review. <i>Animals</i> 12(823):1-19 <a href="https://doi.org/10.3390/ani/2070823">https://doi.org/10.3390/ani/2070823</a> .
57.	Ghafoor, K., Fahad, A., Mehmet, M.O., Isam, M.A., Elfadil, E.B. & Omer N.A. (2020). Evaluti on of the Atioxidant Activity of some Plant Extracts (Rosemary, Sage, and Savory, Summer) on Stability of Moringa Oil. <i>Journal of Food Processing and Preservation</i> 1. 45(3). https://doi.org/10.1111/jfpp.15203
58.	Ukwe,I.O.K.&Gabriel,U.U.(2019).HerbsandHerbalsupplements:keytoaproductive, healthy and Eco-Friendly Aquaculture. <i>Delta Agriculturist</i> , 11 (1/1): 55-67.

59.	De Verdal, H., Vandeputte, M. Nekkawy Wo, Chatan B., & Benzie, J. A. H., (2018).QuantifyingthegeneticparametersoffeedefficiencyinJurveniteNiletilapia. <i>Oreoch romisniliticus.BMCGenetics</i> , 19(1),105https//dox.org/1p.1186/S12863-018-0691-y.
60.	Qi, G., Ai, Q., Mai, K., Xu, W., Liufu, Z., Yun, B., & Zhou, H (2012). Effect ofDietary Taurine supplemention to a caseiu- based diet on growth performance andtaurinedistributionintwosizesofjuvenileturbot( <i>ScophthalmusMaximusL.</i> )Aquacultur e, 358-355,122-128.
61	Citarasu, T. (2010). Herbalbiomedicines: a new opportunity for a quaculture industry. Aquaculture International, 18:403-414.
62.	Zhang, f., Man, Y.B., Mo, W.Y., Wong, M.H. (2020). Application of spirulina inaquaculture:areviewonwastewatertreatmentandfishgrowth.ReviewsinAquaculture12(2): 582-599.
63.	Ekanem, A.P., Eyo, V.O. & Ndome, C.B. (2010). The Efeect of diet with differentinclusionlevelsofcassavaleafmeal(CLM)Manihotutilissimaonthegrowthperfor manceofheteroclaraisfingerlings.J.sci.muitidisciplinaryRes.,2,58-67.
64.	Ashry, A.M., Habiba, M.M., El-Zayat, A.m., Badreldeen, A.h., Younis, N.A., Ahmed, H. A., El-Dakroury, M.F., M Ali, M.A.M., & Dawood. M.A.O. (2023). Effects ofginger (Zingiber officinale) on the growth performance, digestive enzyme activity, antioxidative response, and antibacterial capacity of striped catfish (Pangasianodonhypophthalmus) reared inoutdoor conditions. <i>Aquaculture Reports</i> , 33,101 760. <a href="https://doi.org/10.1016/j.aqrep.2023.101760">https://doi.org/10.1016/j.aqrep.2023.101760</a> .
65.	Robb, D.H., S.C. Kestin, P.D. Warriss & P.D. Nute. (2002). Muscle lipid contentdetermines the eating quality of somked and cooked Atlantic salmon (Salma salar). <i>Aquaculture</i> , 2005:345-358.