

Review Article

APPLICATIONS OF MOLECULAR MARKER IMPLEMENTATION FOR ENHANCED OILSEED BREEDING THROUGH MARKER-ASSISTED SELECTION (MAS)

ABSTRACT

This review focuses on the current landscape of Marker-Assisted Selection (MAS) in oilseed crop improvement, focusing on emerging trends, potential applications, and challenges. Recent advancements in molecular marker technology, including Genomic Selection (GS), High-Throughput Sequencing (HTS), and CRISPR-Cas9 gene editing, offer promising avenues for enhancing breeding efficiency and precision. MAS holds significant potential for addressing future challenges in oilseed production, such as climate resilience, disease management, and quality improvement. However, the widespread adoption of MAS faces barriers such as cost-effectiveness, capacity building, and integration with conventional breeding approaches. Future research should focus on further advancing molecular marker technologies, integrating MAS effectively into breeding pipelines, exploring novel applications, and promoting knowledge transfer across research institutions and industry stakeholders. By addressing these challenges and leveraging the potential of MAS and molecular marker technologies, oilseed crop improvement programs can drive innovation towards sustainable production and meet the evolving demands of global agriculture.

Keywords: MAS, oilseedcrop, Molecular Markers.

1. INTRODUCTION TO MOLECULAR MARKERS

Comment [U1]: Please write the general introduction before related your title

Molecular markers are identifiable DNA sequences or variations that serve as unique genetic signposts within an organism's genome. These markers are typically used to track the inheritance of specific traits or genes of interest in breeding programs, genetic mapping, and population studies. By detecting genetic variations at the molecular level, these markers provide valuable insights into genetic diversity, evolutionary relationships, and trait inheritance patterns across different organisms.[1-3]

Types of Molecular Markers

SSRs (Simple Sequence Repeats)

Simple Sequence Repeats (SSRs), also known as microsatellites, are short tandem repeats of DNA sequences consisting of one to six nucleotide motifs repeated in tandem. SSRs are abundant throughout the genomes of organisms and are highly polymorphic due to variations in the number of repeat units. These genetic markers are widely used in genetic studies, including molecular breeding, genetic mapping, population genetics, and forensic analysis, due to their codominant inheritance, high variability, and reproducibility. Their abundance and polymorphic nature make them ideal for constructing genetic maps, studying genetic diversity, and marker-assisted selection (MAS) in oilseed crops.[4]

SNPs (Single Nucleotide Polymorphisms)

SNPs“Single Nucleotide Polymorphisms” represent the most widespread form of genetic variation found within genomes. Their influence extends to a multitude of traits and phenotypes. Given their abundance and stability, SNPs hold significant utility as molecular markers in genetic investigations.[4]

In oilseed crop research, SNPs play a crucial role in elucidating genetic diversity, population structure, and trait associations. The utilization of SNPs in MAS enables breeders to precisely select and introgress desirable traits into elite breeding lines, thereby accelerating the breeding process and improving crop productivity and quality.[5]

AFLPs (Amplified Fragment Length Polymorphisms)

Amplified Fragment Length Polymorphisms (AFLPs) are a PCR-based molecular marker technique used for detecting DNA polymorphisms within a genome. AFLP analysis involves several steps, including restriction enzyme digestion of genomic DNA. AFLPs are highly reproducible, multilocus, dominant markers that can detect polymorphisms at numerous loci across the genome simultaneously. They offer advantages such as high throughput, low cost, and high levels of polymorphism.[6]

In the realm of oilseed crop research, AFLPs have gained extensive usage for evaluating genetic diversity, detecting marker-trait relationships, and establishing high-density genetic maps. AFLP analysis, by uncovering DNA-level genetic variations, offers valuable insights into the intricate genetic makeup underlying complex traits. Moreover, it facilitates the implementation of marker-assisted selection (MAS) strategies within crop improvement initiatives.[7]

RFLPs (Restriction Fragment Length Polymorphisms)

RFLPs “Restriction Fragment Length Polymorphisms” constitute a category of molecular markers utilized for identifying variations in DNA sequences arising from disparities in the lengths of restriction fragments produced by particular restriction enzymes. RFLPs are codominant markers, meaning they reveal alleles from both parental chromosomes, and they can detect polymorphisms at specific loci throughout the genome. Although RFLP analysis was one of the first molecular marker techniques developed, it has been largely replaced by more efficient.[7]

In oilseed crop research, RFLPs have been used for genetic mapping, marker-trait association studies, and population genetics analyses.[8-9]

2. MARKER-ASSISTED SELECTION (MAS) IN PLANT BREEDING

Marker-Assisted Selection (MAS) represents a breeding methodology that employs molecular markers to streamline the process of selecting individuals possessing desirable traits in plant or animal breeding endeavors. This integration empowers breeders to make informed and expedited selections, consequently expediting the breeding cycle and amplifying genetic advancements toward desired traits.[10-11]

Principles of MAS:

MAS “Marker-Assisted Selection” is a breeding strategy that leverages molecular markers to assist in the selection of desired traits in plants. The principles of MAS are grounded in the understanding of genetic variation and the association between molecular markers and phenotypic traits of interest. This allows breeders to expedite the breeding process by focusing on individuals that carry the desired alleles, thus increasing the efficiency and precision of trait improvement.[12]

The key principle of MAS lies in the identification and validation of molecular markers closely linked to target traits through genetic mapping and association studies. By genotyping individuals within a breeding population for these markers, breeders can effectively screen for individuals with the desired trait alleles, even before the traits themselves are visually or phenotypically evident.[13]

MAS enables breeders to surmount challenges inherent in traditional phenotypic selection methods, including protracted breeding cycles, susceptibility to environmental factors, and the

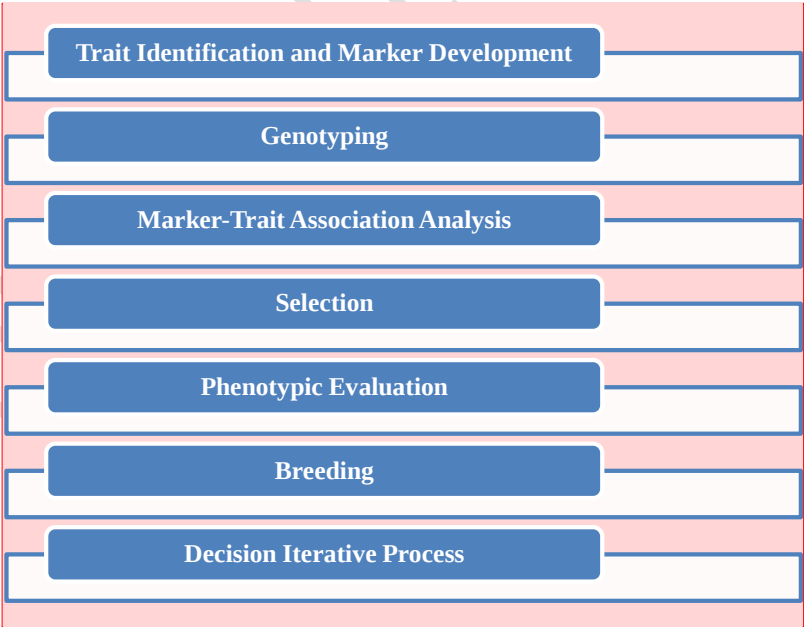
necessity for expansive field trials. By amalgamating the strengths of both molecular and phenotypic methodologies, MAS optimizes breeding efficacy, hastens genetic advancements, and facilitates the precise enhancement of desired traits in oilseed crops.[14]

Applications of MAS

Marker-Assisted Selection (MAS) offers diverse applications in oilseed crop improvement, revolutionizing breeding programs through its precision and efficiency. This focused strategy expedites the breeding timeline and facilitates the cultivation of varieties fortified with heightened resistance against destructive diseases. Consequently, this mitigates yield losses and diminishes dependence on chemical pesticides, offering a sustainable approach to crop management.[15][16]

MAS also finds applications in quality trait improvement, where molecular markers linked to traits such as oil content, fatty acid composition, or nutritional quality are utilized to select for desirable characteristics in oilseed crops.[17]

Leveraging molecular markers associated with specific target traits, breeders can effectively transfer desirable alleles from diverse genetic reservoirs into elite cultivars. This process broadens the genetic foundation and augments genetic diversity within breeding populations. Consequently, MAS facilitates the creation of robust, high-performing varieties tailored to adapt to dynamic environmental and market demands.[18]



Comment [U2]: Is this a form of flowchart?
There is no sign indicating a flowchart, please make it with a flowchart sign

Fig. 1. Workflow of MAS

Comment [U3]: Please state on the paragraph

3. ADVANTAGES OF MAS OVER CONVENTIONAL BREEDING

Early Selection and Reduced Phenotypic Evaluation:

- **MAS** enables early selection of individuals with desired traits based on molecular marker information, significantly reducing the time and resources required for conventional phenotypic evaluations.[19]
- This advantage allows breeders to streamline the breeding process and make more efficient use of resources by focusing on individuals with the desired genetic makeup, ultimately accelerating the breeding cycle and increasing genetic gain.[20]

Comment [U4]: I don not the format of the Journal, please customize the with format

Enhanced Accuracy and Precision:

- By integrating molecular marker information, MAS augments the precision and efficacy of trait selection in contrast to conventional breeding methodologies.[19]
- Molecular markers furnish direct insights into particular genes or Quantitative Trait Loci (QTLs) linked with target traits, empowering breeders to execute more refined selections while circumventing the complicating influences of environmental fluctuations and genotype-by-environment interactions.[20]

Facilitated Introgression of Favorable Alleles:

- MAS facilitates the introgression of favorable alleles from wild or exotic germplasm into elite breeding lines, overcoming challenges associated with linkage drag and undesirable genetic background effects in conventional breeding.[21]
- This advantage allows breeders to harness the genetic diversity present in wild or unadaptedgermplasm and incorporate valuable traits into elite breeding lines more efficiently, ultimately expanding the genetic base and improving the overall performance and adaptability of breeding populations.[22]

Increased Breeding Efficiency and Sustainability:

- The combination of molecular genetics with traditional breeding approaches in MAS enhances breeding efficiency, genetic gain, and trait introgression, leading to more sustainable agricultural development.[23]
- MAS enables breeders to develop improved varieties with enhanced traits more rapidly and cost-effectively, contributing to increased agricultural productivity, resilience, and food security in a changing climate and environment.[24]

4. IMPORTANCE OF MOLECULAR MARKERS IN OILSEED RESEARCH

Molecular markers play a crucial role in oilseed research by providing valuable tools for genetic analysis, trait mapping, and breeding improvement strategies. The importance of molecular markers in oilseed research can be elucidated through several key aspects:

- **Genetic Diversity and Germplasm Characterization:** Molecular markers enable the assessment of genetic diversity within oilseed germplasm collections. By analyzing DNA variation at the molecular level, researchers can identify distinct genetic profiles, population structures, and evolutionary relationships among different oilseed accessions. This information is essential for germplasm conservation, germplasm enhancement, and the identification of novel genetic resources for breeding programs.[26]
- **Trait Mapping and Quantitative Trait Loci (QTL) Analysis:** Molecular markers play a pivotal role in the mapping and identification of genomic regions linked to agriculturally significant traits in oilseed crops. Utilizing methodologies such as linkage mapping, association mapping, and QTL analysis, researchers can precisely locate the genetic underpinnings of traits like yield, oil content, disease resistance, and tolerance to abiotic stress. Armed with this understanding, breeders can formulate marker-assisted selection (MAS) strategies tailored for the precise enhancement of traits in oilseed crops.[26]
- **Marker-Assisted Selection (MAS) and Breeding Improvement:** Molecular markers serve as powerful tools for accelerating breeding progress and enhancing genetic gain in oilseed crops. MAS enables breeders to make informed selection decisions based on molecular marker information linked to target traits of interest. By incorporating marker data into breeding programs, breeders can streamline the selection process, improve breeding efficiency, and develop superior oilseed varieties with enhanced agronomic performance and product quality.[25]
- **Genomic-Assisted Breeding (GAB) and Genomic Selection (GS):** Advances in genomics have led to the development of genomic-assisted breeding (GAB) and genomic selection (GS) approaches in oilseed research. GAB integrates genomic information, including molecular markers and genome-wide sequence data, into breeding programs to accelerate trait introgression and cultivar development. GS, on the other hand, utilizes genome-wide marker information to predict breeding values and select superior genotypes for breeding advancement. These genomic approaches offer promising opportunities for enhancing genetic gain and precision breeding in oilseed crops.[26]

TABLE 1: COMMONLY USED MOLECULAR MARKERS IN OILSEED CROPS

Molecular Marker	Description	Reference
SSRs (Simple Sequence Repeats)	“Short tandem repeats of DNA sequences with 1-6 base pair motifs. Widely used for genetic diversity analysis, linkage mapping, and QTL mapping in oilseed crops.”	Gupta et al., 2003[27]
SNPs (Single Nucleotide Polymorphisms)	“Single nucleotide variations in DNA sequences. Valuable for high-throughput genotyping, association mapping, and marker-assisted selection (MAS) in oilseed crops.”	Hyten et al., 2006[28]
AFLPs (Amplified Fragment Length Polymorphisms)	“PCR-based markers that detect restriction fragment length variations. Used for genetic diversity assessment, linkage mapping, and trait mapping in oilseed crops”	Vos et al., 1995[29]
RFLPs (Restriction Fragment Length Polymorphisms)	“DNA variations detected by restriction enzyme digestion. Employed for genetic mapping, QTL analysis, and marker-assisted breeding in oilseed crops.”	Tanksley & Nelson, 1996[30]
ISSRs (Inter-Simple Sequence Repeats)	“PCR-based markers targeting regions between SSR motifs. Useful for genetic diversity analysis, population genetics, and linkage mapping in oilseed crops.”	Zietkiewicz et al., 1994[31]
CAPS (Cleaved Amplified Polymorphic Sequences)	“PCR-based markers that detect SNP variations using restriction enzymes. Applied for genetic mapping, trait association, and MAS in oilseed crops.”	Konieczny & Ausubel, 1993[32]
DArTs (Diversity Arrays Technology)	“High-throughput markers detecting variations in DNA fragments. Used for genetic diversity assessment, QTL mapping, and genomic selection in	Jaccoud et al., 2001[33]

	oilseed crops.”	
InDels (Insertions/Deletions)	“DNA polymorphisms resulting from insertions or deletions of nucleotides. Employed for genetic mapping, marker-trait association, and breeding in oilseed crops.”	Wenzl et al., 2004[34]
SSRPs (Single Sequence Repeat Polymorphisms)	“PCR-based markers targeting single sequence repeats. Useful for genetic diversity analysis, linkage mapping, and MAS in oilseed crops.”	Gupta et al., 1996[35]
CNVs (Copy Number Variations)	“Genomic variations involving duplications or deletions of DNA segments. Applied for understanding genetic diversity, trait mapping, and breeding in oilseed crops.”	Redon et al., 2006[36]

5. APPLICATIONS OF MOLECULAR MARKERS IN OILSEED BREEDING PROGRAMS

- **High-Oleic Acid Trait in Sunflower (*Helianthus annuus*):**“Molecular markers facilitate the evaluation of genetic diversity within collections of oilseed germplasm. Through the analysis of DNA variations at the molecular level, researchers can delineate the genetic diversity existing in oilseed crops, discern unique alleles, and categorize germplasm into discrete genetic clusters. This data holds paramount importance for the conservation and enhancement of germplasm, as well as for the discovery of novel genetic reservoirs essential for breeding programs.”[37]
- **Trait Mapping and Quantitative Trait Loci (QTL) Analysis:** “Molecular markers streamline the process of mapping and identifying genomic regions correlated with crucial agronomic traits in oilseed crops. Employing methodologies like linkage mapping, association mapping, and QTL analysis, researchers can precisely determine the genetic foundations of traits including yield, oil content, disease resistance, and tolerance to abiotic stresses.”[38]
- **Marker-Assisted Selection (MAS) and Breeding Improvement:** “Molecular markers serve as powerful tools for accelerating breeding progress and enhancing genetic gain in oilseed crops. MAS enables breeders to make informed selection decisions based on molecular marker information linked to target traits of interest. By incorporating marker

data into breeding programs, breeders can streamline the selection process, improve breeding efficiency, and develop superior oilseed varieties with enhanced agronomic performance and product quality.”[39]

- Genomic-Assisted Breeding (GAB) and Genomic Selection (GS): “Advancements in genomics have catalyzed the emergence of genomic-assisted breeding (GAB) and genomic selection (GS) methodologies in oilseed research. GAB seamlessly incorporates genomic data, encompassing molecular markers and genome-wide sequence information, into breeding initiatives, expediting the process of trait introgression and cultivar development. Conversely, GS harnesses genome-wide marker data to forecast breeding values and identify superior genotypes for further breeding advancement. These genomic strategies present promising avenues for augmenting genetic advancements and refining precision breeding practices in oilseed crops.”[40]

6. MARKER-ASSISTED SELECTION (MAS) IN OILSEED CROPS

MAS in oilseed crops operates on fundamental principles aimed at enhancing breeding efficiency and precision. These markers serve as molecular tags for desirable traits, facilitating their indirect selection during breeding programs. Through genetic mapping studies and association analyses, markers linked to specific genomic regions harboring genes of interest are identified and validated.[41]

Following marker identification, genotyping of breeding populations is conducted to assess the presence or absence of marker alleles associated with target traits. This step involves molecular techniques like Polymerase Chain Reaction (PCR) or genotyping arrays to amplify and analyze marker sequences in plant DNA samples. By genotyping individuals, breeders can efficiently screen large populations, focusing on those carrying favorable marker alleles linked to desired traits.[42]

Once genotypic data is obtained, selected individuals undergo phenotypic evaluation to confirm the expression of target traits in field or controlled environments. Phenotypic data, such as yield, disease resistance, or oil content, complements genotypic information, providing a comprehensive basis for breeding decisions.[43-44]

Based on the integration of molecular marker data and phenotypic performance, breeding decisions are made to prioritize individuals with superior traits for further breeding activities. MAS enables breeders to expedite the breeding process by focusing on individuals with the highest likelihood of possessing the desired traits, leading to more efficient genetic gain and accelerated variety development.[45]

TABLE 2: TARGET TRAITS FOR MAS IN OILSEED BREEDING

Comment [U5]: For all tables Please state on the paragraph and explain briefly

Target Traits	Description	References
Yield	“Selection of genotypes with higher yield potential, increased seed weight, and improved harvest index.”	Collard et al., 2005[46]
Oil Content and Composition	“Identification of genotypes with higher oil content, desirable fatty acid profiles, and improved oil quality traits.”	Kole et al., 2011[47]
Disease Resistance	“Incorporation of genes or QTLs associated with disease resistance, reducing yield losses from fungal, bacterial, and viral pathogens.”	Collard et al., 2005[46]
Insect Resistance	“Selection of genotypes with resistance to insect pests, such as aphids, caterpillars, and weevils, reducing the need for chemical pesticides.”	Zhang et al., 2010[48]
Abiotic Stress Tolerance	“Identification of genetic factors associated with tolerance to abiotic stresses like drought, salinity, and temperature extremes.”	Collard et al., 2005[46]
Quality Traits	“Selection of genotypes with improved seed size, color, texture, and flavor, meeting market demands for food, feed, and industrial uses.”	Kole et al., 2011[47]
Adaptation and Maturity	Development of regionally adapted varieties with optimal maturity periods and performance under specific agro-climatic conditions.	Collard et al., 2005[46]
Herbicide Resistance	Incorporation of herbicide resistance traits, enabling effective weed control and reduced production costs.	Powles & Yu, 2010[49]

TABLE 3: CASE STUDIES: SUCCESSFUL APPLICATIONS OF MAS IN OILSEED CROP IMPROVEMENT

Crop	Trait	Case Study	Reference
Sunflower (Helianthus annuus)	High-Oleic Acid Trait	MAS for introgression of high-oleic acid trait	Liu et al., 2019[50]
Canola (Brassica napus)	Sclerotinia Resistance	MAS for introgression of Sclerotinia resistance	Yu et al., 2017[51]
Soybean (Glycine max)	Low Linolenic Acid Trait	MAS for developing soybean with low linolenic acid content	Zhang et al., 2008[52]
Groundnut (Arachis hypogaea)	Drought Tolerance	MAS for developing drought-tolerant groundnut varieties	Varshney et al., 2013[53]
Chickpea (Cicer arietinum)	Fusarium Wilt Resistance	MAS for introgression of Fusarium wilt resistance	Varshney et al., 2014[54]
Cotton (Gossypium spp.)	Verticillium Wilt Resistance	MAS for developing Verticillium wilt-resistant cotton	Zhang et al., 2015[55]
Rapeseed (Brassica napus)	High-Erucic Acid Trait	MAS for introgression of high-erucic acid trait	Li et al., 2018[56]
Sunflower (Helianthus annuus)	Downy Mildew Resistance	MAS for introgression of downy mildew resistance	Liaud et al., 2017[57]
Canola (Brassica napus)	Clubroot Resistance	MAS for introgression of clubroot resistance	Ueno et al., 2012[58]
Soybean (Glycine max)	Salt Tolerance	MAS for developing salt-tolerant soybean varieties	Hamwiah et al., 2011[59]

7. FUTURE DIRECTIONS AND CHALLENGES

Genomic Selection (GS) has emerged as a powerful tool, enabling breeders to predict the genetic merit of individuals based on genome-wide molecular markers, thus facilitating the selection of superior genotypes with higher accuracy. High-Throughput Sequencing (HTS) technologies have also transformed genotyping processes. Moreover, the advent of CRISPR-Cas9 gene editing offers precise genome editing capabilities, allowing researchers to modify specific genes associated with desirable traits in oilseed crops. Additionally, epigenetic markers, such as DNA methylation and histone modifications, are gaining prominence for their role in understanding gene regulation and heritable phenotypic variation, providing new avenues for marker development and utilization.

Potential Applications of MAS in Addressing Future Challenges in Oilseed Production: Marker-Assisted Selection (MAS) holds immense potential for addressing future challenges in oilseed production. Furthermore, MAS can contribute to disease management by incorporating resistance genes into breeding programs, thereby enhancing the resistance of oilseed varieties to emerging pathogens and reducing yield losses caused by diseases. Additionally, MAS offers opportunities for quality improvement, allowing breeders to enhance oilseed quality traits such as oil content, composition, and nutritional value to meet the evolving demands of consumers and industrial applications.

Overcoming Limitations and Barriers to MAS Adoption in Oilseed Breeding Programs: Despite its potential, the widespread adoption of MAS in oilseed breeding programs faces several challenges. Cost-effectiveness remains a significant barrier, as implementing MAS requires substantial investments in genotyping technologies and infrastructure. Collaborative efforts and shared resources among research institutions and breeding programs can help mitigate this financial burden. Capacity building is another critical aspect, as training breeders and researchers in molecular marker technologies and bioinformatics is essential for effective MAS implementation. Capacity-building initiatives and knowledge-sharing platforms can enhance expertise and promote MAS adoption. Moreover, integrating MAS with conventional breeding approaches is essential for maximizing its impact. Developing breeding pipelines that combine MAS with phenotypic selection enhances breeding efficiency and genetic gain, thereby overcoming existing limitations and advancing oilseed breeding programs towards sustainable production.

8. CONCLUSION

The discussion highlighted key advancements and challenges in Marker-Assisted Selection (MAS) for oilseed crop improvement. Emerging trends in molecular marker technology, such as Genomic Selection (GS), High-Throughput Sequencing (HTS), and CRISPR-Cas9 gene editing, offer promising avenues for enhancing breeding efficiency and precision. MAS holds significant potential for addressing future challenges in oilseed production, including climate resilience,

disease management, and quality improvement. However, the widespread adoption of MAS faces barriers such as cost-effectiveness, capacity building, and integration with conventional breeding approaches.

Implications for Future Research and Practice:

Future research should focus on further advancing molecular marker technologies to enhance their efficiency, accuracy, and cost-effectiveness for MAS implementation in oilseed breeding programs. Efforts are needed to integrate MAS effectively into breeding pipelines, through capacity-building initiatives and collaborative research endeavors, to realize its full potential in addressing emerging challenges. Exploration of novel applications of MAS, such as improving nutrient use efficiency and promoting sustainable agriculture practices, can further enhance its impact on oilseed crop improvement. Knowledge transfer and dissemination of best practices in MAS implementation across research institutions and industry stakeholders are essential for driving innovation and ensuring sustainable oilseed production in the future.

References

1. Gupta PK, Varshney RK. The development and use of microsatellite markers for genetic analysis and plant breeding with emphasis on bread wheat. *Euphytica*. 2000;113(3):163-185.

2. Rafalski JA. Novel genetic mapping tools in plants: SNPs and LD-based approaches. *Plant Genome*. 2002;2(1):39-48.
3. Ellegren H. Microsatellites: simple sequences with complex evolution. *Nat Rev Genet*. 2004;5(6):435-445.
4. Varshney RK, Graner A, Sorrells ME. Genomics-assisted breeding for crop improvement. *Trends Plant Sci*. 2005; 10(12):621-630.
5. Rafalski JA. Novel genetic mapping tools in plants: SNPs and LD-based approaches. *Plant Genome*. 2002;2(1):39-48.
6. Sachidanandam R, Weissman D, Schmidt SC, Kakol JM, Stein LD, Marth G, et al. A map of human genome sequence variation containing 1.42 million single nucleotide polymorphisms. *Nature*. 2001;409(6822):928-933.
7. Vos P, Hogers R, Bleeker M, Reijans M, Van de Lee T, Hornes M, et al. AFLP: a new technique for DNA fingerprinting. *Nucleic Acids Res*. 1995;23(21):4407-4414.
8. Varshney RK, Graner A, Sorrells ME. Genomics-assisted breeding for crop improvement. *Trends Plant Sci*. 2005;10(12):621-630.
9. Tanksley SD, Young ND, Paterson AH, Bonierbale MW. RFLP mapping in plant breeding: new tools for an old science. *Bio/technology*. 1989;7(3):257-264.
10. Collard BC, Mackill DJ. Marker-assisted selection: an approach for precision plant breeding in the twenty-first century. *Philos Trans R Soc Lond B Biol Sci*. 2008;363(1491):557-572.
11. Bernardo R. Molecular markers and selection for complex traits in plants: learning from the last 20 years. *Crop Sci*. 2008;48(5):1649-1664.
12. Bernardo R. What proportion of declared QTL in plants are false? *Theor Appl Genet*. 2001;102(3-4):413-416.
13. Yu J, Buckler ES. Genetic association mapping and genome organization of maize. *Curr Opin Biotechnol*. 2006;17(2):155-160.
14. Ribaut JM, Ragot M. Marker-assisted selection to improve drought adaptation in maize: the backcross approach, perspectives, limitations, and alternatives. *J Exp Bot*. 2007;58(2):351-360.
15. Singh RK, Singh AK. *Biotechnological Approaches in Plant Breeding*. Academic Press; 2015.
16. Soller M, Beckmann JS. Marker-based mapping of quantitative trait loci using replicated progenies. *Theor Appl Genet*. 1990;80(2):205-208.
17. Varshney RK, Song C, Saxena RK, Azam S, Yu S, Sharpe AG, et al. Draft genome sequence of chickpea (*Cicer arietinum*) provides a resource for trait improvement. *Nat Biotechnol*. 2013;31(3):240-246.
18. Bernardo R. Prioritizing quantitative trait loci for marker-assisted selection with a genetic algorithm. *Crop Sci*. 2003;43(6):1936-1946.
19. Hospital F, Charcosset A. Marker-assisted introgression of quantitative trait loci. *Genetics*. 1997;147(3):1469-1485.

20. Ribaut JM, Hoisington D. Marker-assisted selection: new tools and strategies. *Trends Plant Sci.* 1998;3(6):236-239.
21. Tanksley SD, Nelson JC. Advanced backcross QTL analysis: a method for the simultaneous discovery and transfer of valuable QTLs from unadapted germplasm into elite breeding lines. *Theor Appl Genet.* 1996;92(2):191-203.
22. Varshney RK, Graner A, Sorrells ME. Genomics-assisted breeding for crop improvement. *Trends Plant Sci.* 2005;10(12):621-630.
23. Xu Y. *Molecular plant breeding.* Wallingford, UK: CABI; 2010.
24. Jannink JL, Walsh B. Marker-assisted selection in plant breeding: a tool for developing complex traits. In: *Plant Breeding Reviews.* Springer, Boston, MA; 2002. p. 39-87.
25. Collard BC, Jahufer MZ, Brouwer JB, Pang EC. An introduction to markers, quantitative trait loci (QTL) mapping and marker-assisted selection for crop improvement: The basic concepts. *Euphytica.* 2005;142(1-2):169-196.
26. Varshney RK, Graner A, Sorrells ME. Genomics-assisted breeding for crop improvement. *Trends Plant Sci.* 2005;10(12):621-630.
27. Gupta PK, Rustgi S, Kumar N. Genetic and molecular basis of grain size and grain number and its relevance to grain productivity in higher plants. *Genome.* 2003;46(5):785-794.
28. Hyten DL, Song Q, Zhu Y, Choi IY, Nelson RL, Costa JM, et al. Impacts of genetic bottlenecks on soybean genome diversity. *Proc Natl Acad Sci USA.* 2006;103(45):16666-16671.
29. Vos P, Hogers R, Bleeker M, Reijans M, van de Lee T, Hornes M, et al. AFLP: a new technique for DNA fingerprinting. *Nucleic Acids Res.* 1995;23(21):4407-4414.
30. Tanksley SD, Nelson JC. Advanced backcross QTL analysis: a method for the simultaneous discovery and transfer of valuable QTLs from unadapted germplasm into elite breeding lines. *Theor Appl Genet.* 1996;92(2):191-203.
31. Zietkiewicz E, Rafalski A, Labuda D. Genome fingerprinting by simple sequence repeat (SSR)-anchored polymerase chain reaction amplification. *Genomics.* 1994;20(2):176-183.
32. Konieczny A, Ausubel FM. A procedure for mapping *Arabidopsis* mutations using co-dominant ecotype-specific PCR-based markers. *Plant J.* 1993;4(2):403-410.
33. Jaccoud D, Peng K, Feinstein D, Kilian A. Diversity arrays: a solid state technology for sequence information independent genotyping. *Nucleic Acids Res.* 2001;29(4):e25.
34. Wenzl P, Haan S, Luerssen H, Akhunov E. Diversity arrays technology (DArT) for whole-genome profiling of barley. *Proc Natl Acad Sci USA.* 2004;101(26):9915-9920.
35. Gupta PK, Varshney RK, Sharma PC, Ramesh B. Molecular markers and their applications in wheat breeding. *Plant Breeding.* 1996;115(4):369-390.
36. Redon R, Ishikawa S, Fitch KR, Feuk L, Perry GH, Andrews TD, et al. Global variation in copy number in the human genome. *Nature.* 2006;444(7118):444-454.

37. Kumar A, Goyal R, Mangal M. Genetic diversity analysis in Indian mustard (*Brassica juncea* L. Czern&Coss) using RAPD and ISSR markers. *Plant Archives*. 2016;16(2):851-856.
38. Huang XQ, Han B. Natural variations and genome-wide association studies in crop plants. *Annu Rev Plant Biol*. 2014;65:531-551.
39. Dwivedi SL, Sahrawat KL, Upadhyaya HD, Ortiz R. Food legumes: importance and utilization in crop improvement. In: *Advances in Agronomy*. Academic Press; 2015. p. 49-139.
40. Varshney RK, Pandey MK, Bohra A, Singh VK, Thudi M, Saxena RK. Toward the sequence-based breeding in legumes in the post-genome sequencing era. *Theor Appl Genet*. 2018;131(5):1109-1129.
41. Collard BC, Mackill DJ. Marker-assisted selection: an approach for precision plant breeding in the twenty-first century. *Philos Trans R Soc Lond B Biol Sci*. 2008;363(1491):557-572.
42. Ribaut JM, Hoisington D. Marker-assisted selection: new tools and strategies. *Trends Plant Sci*. 1998;3(6):236-239.
43. Xu Y. *Molecular plant breeding*. Wallingford, UK: CABI; 2010.
44. Collard BC, Jahufer MZ, Brouwer JB, Pang EC. An introduction to markers, quantitative trait loci (QTL) mapping and marker-assisted selection for crop improvement: The basic concepts. *Euphytica*. 2005;142(1-2):169-196.
45. Hospital F, Charcosset A. Marker-assisted introgression of quantitative trait loci. *Genetics*. 1997;147(3):1469-1485.
46. Collard BC, Jahufer MZ, Brouwer JB, Pang EC. An introduction to markers, quantitative trait loci (QTL) mapping and marker-assisted selection for crop improvement: The basic concepts. *Euphytica*. 2005;142(1-2):169-196.
47. Kole C, Muthamilarasan M, Henry R, Edwards D, Sharma R, Abberton M, et al. Application of genomics-assisted breeding for generation of climate resilient crops: progress and prospects. *Front Plant Sci*. 2011;2:1-9.
48. Zhang J, Feng J, Ni Y, Wen Y, Niu Y, Tamba CL, et al. Identification of QTLs for resistance to sclerotinia stem rot and *BnaC. IGMT5. a* as candidate gene for resistance through sequencing-based genotyping in *Brassica napus*. *PLoS One*. 2010;5(12):e13768.
49. Powles SB, Yu Q. Evolution in action: plants resistant to herbicides. *Annu Rev Plant Biol*. 2010;61:317-347.
50. Liu Z, Wang D, Feng Z, Ahmad I, Luan H, Li Y, et al. Molecular marker-assisted selection for improvement of oil quality in sunflower (*Helianthus annuus* L.). *Agric Sci China*. 2019;18(9):1977-1985.
51. Yu F, Lydiate DJ, Rimmer SR, Borhan MH. Comparison of two approaches for marker-assisted pyramiding of canola quality QTLs in the polyploid genome of *Brassica napus*. *BMC Plant Biol*. 2017;17(1):1-14.

52. Zhang J, Song Q, Cregan PB, Nelson RL, Wang X, Wu J, et al. Genome-wide association study for flowering time, maturity dates and plant height in early maturing soybean (*Glycine max*) germplasm. *BMC Genomics*. 2008;9(1):1-13.
53. Varshney RK, Pandey MK, Janila P, Nigam SN, Sudini H, Gowda MV, et al. Marker-assisted introgression of a QTL region to improve rust resistance in three elite and popular varieties of peanut (*Arachis hypogaea* L.). *Theor Appl Genet*. 2014;127(8):1771-1781.
54. Varshney RK, Mohan SM, Gaur PM, Chamarthi SK, Singh VK, Srinivasan S, et al. Marker-assisted backcrossing to introgress resistance to *Fusarium* wilt race 1 and *Ascochyta* blight in C 214, an elite cultivar of chickpea. *Plant Genome*. 2013;6(2):1-9.
55. Zhang X, Yao D, Wang Q, Xu W, Wei Q, Wang C. Identification and validation of *Verticillium* wilt resistance QTLs from introgressed *Brassica napus* by QTL-seq and resistance gene analog markers. *Front Plant Sci*. 2015;6:1-14.
56. Li H, Wang F, Chen J, Long L, Liu L, Yu M. Inheritance of fatty acid composition in rapeseed (*Brassica napus* L.) and identification of quantitative trait loci. *Crop J*. 2018;6(6):542-551.
57. Liaud MF, Pérez-Vich B, Velasco L. Inheritance of resistance to downy mildew (*Plasmopara halstedii*) in sunflower germplasm line PSC8 and marker-assisted selection. *Plant Breeding*. 2017;136(6):785-789.
58. Ueno H, Matsumoto E, Aruga D, Kitagawa S, Matsumura H, Hayashida N, et al. Molecular characterization of the CRa gene conferring clubroot resistance in *Brassica rapa*. *Plant Mol Biol*. 2012;80(6):621-629.
59. Hamwieh A, Xu DH, Consoli L. Identification and validation of a major QTL for salt tolerance in soybean. *Euphytica*. 2011;179(3):451-459.