

Original Research Article

Isolation and characterization of heavy metal-tolerant bacterial isolates from industrial effluents in Uttar Pradesh, India

ABSTRACT

Industrial activities over the past century have significantly increased human exposure to pollutants such as heavy metals. Industrial emissions of heavy metals, which are carcinogenic, mutagenic and toxic, contaminate natural water supplies and the agricultural environment. Due to the high concentration of heavy metals in industrial effluents, the bacteria present there naturally develop resistance to heavy metals. The aim of this study is to isolate and characterize bacteria resistant to heavy metal, lead (Pb), copper (Cu), chromium (Cr), cadmium (Cd), and nickel (Ni) from sites contaminated by industrial effluents in Uttar Pradesh. Two isolates were identified up to species level based on their physiological, biochemical and molecular characterization as *Comamonastestosteroni* (S4C1) and *Bacillus cereus* (S5C3). Both isolates are highly resistant to lead (Pb), copper (Cu), chromium (Cr), cadmium (Cd), and nickel (Ni) and they show different MICs against the above heavy metals at different levels. A growth experiment showed that the presence of heavy metals concentration had no discernible impact on the growth rate among the isolates. Gel analysis showed interesting patterns of protein expression were observed in the presence of various heavy metals. MALDI-TOF analysis found that specific proteins (S layer protein, F0F1ATP synthase subunit b, Flagellin, 50S ribosomal protein L4, Molecular chaperone) were overexpressed in the presence of heavy metals. As a result, identifying the heavy metal resistance bacteria and their proteins study could be useful as a preliminary investigation for the development of prospective bioremediation agents of potentially hazardous waste treatment technology.

Keywords: Environment, Wastewater, Industrial effluents, Pollution, Heavy metals *Comamonastestosteroni*, *Bacillus cereus*.

1. INTRODUCTION

One of the main causes of environmental pollution is heavy metals. The amounts of these harmful chemical compounds in the environment vary greatly because they are released as a result of the effluent that is discharged into the environment by a huge number of industrial activities, including metal processing, mining, electroplating, leather tanning, and pigment manufacture [1,2]. Excessive heavy metal concentrations in wastewater are not broken down by the traditional wastewater treatment process, which negatively affects aquatic life

[3,4]. While many heavy metals, including copper, iron, and zinc, are necessary trace elements for cells at low concentrations, they can have hazardous effects at higher concentrations found in contaminated settings [5,6]. According to Chovanová et al., [7] cadmium is not only harmful to humans, animals, plants, and microorganisms, but it is also non-essential. Lead is a highly poisonous and dangerous pollutant that can be found in soil, water, and the air [8-11]. The most common forms of chromium in the environment are hexavalent (Cr(VI)) and trivalent (Cr(III)), both of which are harmful at high concentrations [12]. Besides exceedingly toxic and harmful, heavy metals are present in soluble form in the aquatic environment and disrupt ecological activities [13-15]. In addition, they can create complicated unspecific chemicals inside of cells, which has harmful effects. Many microorganisms, including bacteria, can evolve resistance and become heavy metal resistant in response to harmful quantities of metal ions [16]. Bioaccumulation [17], bio-sorption [18], bio-mineralization and precipitation [19], enzymatic oxidation or reduction to a less toxic form, and efflux of heavy metal systems are some of the various intra- and extracellular mechanisms found in bacteria [20]. Heavy metal stress is known to change a native bacterial community's makeup so that resistant and tolerant bacteria become widely predominate [21]. The removal of metal ions by microbes has attracted a lot of attention due to its potential use in protecting the environment and recovering harmful or important heavy metals, [22]. The present research aims to isolate and characterize heavy metal resistant bacteria from industrial effluents.

2. MATERIAL AND METHODS

Materials: To prepare metal stock solutions: lead, copper, chromium, cadmium, and nickel salts were used to prepare stock solution. Applicable solutions were prepared daily by diluting the stock solution. Stocks were used to adjust the pH of the solutions. All chemical materials and culture media were procured from Sigma-Aldrich.

Study area and sample collection: In this study, effluent samples were collected in sterilized screw cap bottles from the industrial discharge area, Fertilizer production unit, Pryagraj and Iffco fertilizer Bareilly, U.P., India. The samples were transported with ice boxes to the laboratory for further analysis.

Isolation of heavy metal resistant bacteria: Isolation of the bacteria from the collected samples were carried out using the serial dilution and spread plate method, where the samples were serially diluted in 0.85% NaCl solution and then spread in sterilized nutrient agar media and incubated at 37 °C, for 24 hours. The isolates were further screened for the heavy metals in Luria-Bertani (LB) agar media supplemented with 100 ppm (~100 mg/L) of PbNO₃, CuSO₄, K₂Cr₂O₇, CdCl₂.H₂O and NiCl₂.6H₂O. The inhibition in growth of bacteria indicates positive results [23, 24].

Minimum Inhibitory Concentrations (MICs) study: Heavy metal resistance were assessed through MIC protocol described previously by Filali et al. (2000). Isolated bacterial strains were grown on nutrient broth (NB) medium containing the different concentrations of heavy metals. The stock solution of concentration 1000 ppm (1 mg/ml) of different heavy metal was prepared and sterilized by autoclaving at 121 °C for 15 min. The cultures were allowed to grow at a given concentration and incubated at 37 °C for 48 hours in shaker incubator. The absorbance of bacterial growth was studied at 600 nm. The MIC was defined as the concentration at which a particular bacterial strain could not grow [30,31].

Studies of optimum pH and temperature on growth conditions: The growth of bacterial isolates in nutrient broth medium at various pH values (ranging from 3 to 12), temperatures

(ranging from 10 °C to 65 °C) was studied. In order to quantify the growth, the optical density of the log phase growing conditions was measured at 600 nm [32-34].

Heavy metal biodegradability assay: Sterilized different culture medium was prepared and supplemented with different heavy metals. The bacterial cultures were inoculated and then media was incubated at 37 °C for 48 hours. The samples were collected from the control and then after 48 hours of treatment and then processed for Atomic Absorption Spectroscopy (AAS) analysis [35].

Identification of heavy metal resistant bacteria: The identification of the bacterial culture which showed the best results in the presence of PbNO₃, CuSO₄, K₂Cr₂O₇, CdCl₂H₂O and NiCl₂6H₂O was carried out through biochemical and molecular characterization. Gram staining, catalase test, glucose fermentation test, Methyl Red and Voges-Proskauer (MRVP) test were carried out for biochemical characterization [25,26].

Molecular characterization of heavy metal tolerant bacterial isolates: The molecular identification of the culture was carried out through 16S rDNA sequencing. Primer of 16S rDNA gene was generated from forward and reverse sequence data using aligner software. The 16S rDNA gene sequence was used to carry out BLAST with the database of NCBI GenBank database. Based on maximum identity score, the first ten sequences were selected and aligned using multiple alignment software program ClustalW. Distance matrix was generated, and the phylogenetic tree was constructed using MEGA7 [27-29].

Protein characterization: The protein profile was studied after extraction of the protein from bacterial culture using PBS buffer and then quantification was carried out using Bradford assay. Further the characterization and identification of proteins was carried out using SDS page and 2D Gel electrophoresis followed by MALDI-TOF-MS [36].

Statistical Analysis: Three replicates of each experiment were carried out. The means \pm standard deviation are used to present the data. Tukey tests were used after one-way Analysis of Variance (ANOVA) for statistical analysis, and a difference of $p < 0.05$ considered significant.

3. RESULTS AND DISCUSSION

Isolation and preliminary screening of heavy metal tolerant bacterial isolates: Total 58 bacterial strains were isolated from the selected 9 samples according to their different morphological parameters. Based on primary screening conducted on LB agar medium with a heavy metal concentration of 100 ppm (~100 mg/L). Total 32 strains were found positive in different all heavy metals after 24-72 hours of incubation at 37°C which showed that the collected effluent sample have metal resistant diversity of bacteria. The colonies count was performed of bacteria that were able to grow well on medium supplemented with Lead (8), Copper (4), Chromium (9), Nickel (6) and cadmium (6) similar study published by Joshi & Modi [37]; Zahoor & Rehman [38] found various heavy metal resistant microbes from industrial effluents. Among all these bacterial isolates, S4C1 and S5C3 were selected for further study as it showed maximum results with all heavy metal in screening as shown in Figure 1.

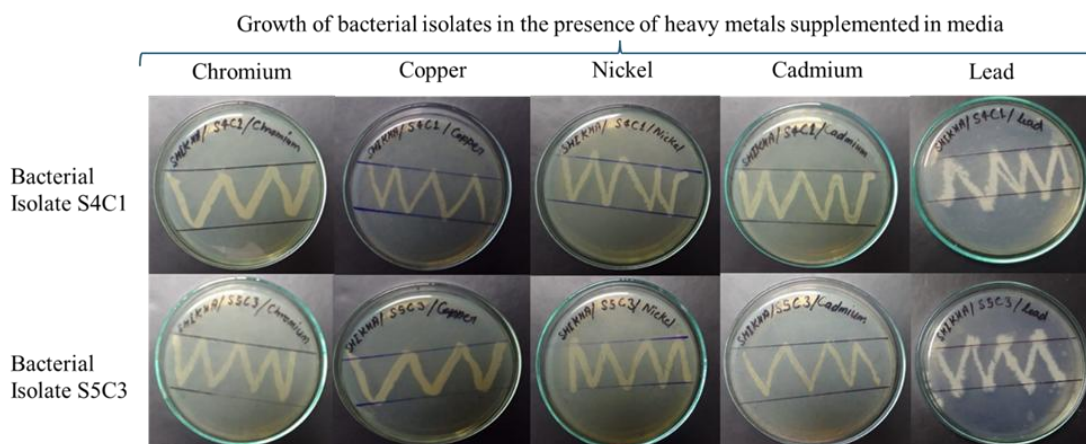


Figure 1: Growth of bacterial isolates with heavy metal (Chromium, Copper, Nickel, cadmium and Lead) tolerance properties in the presence of 100 ppm (~100 mg/L) concentrations on LB agar media

Minimum Inhibitory Concentrations: The MIC of heavy metal resistance in bacterial isolates S4C1 and S5C3 was found to be between 300 $\mu\text{g/ml}$ and 420 $\mu\text{g/ml}$ (Figure 2). S4C1 and S5C3 isolates showed a strong tendency to withstand and grow in environments with heavy metal stress, Bhardwaj et al. 2018 [39]; Hassen et al. 1998 [40] found similar outcome in their findings. Therefore, Potential bacterial isolates S4C1 and S5C3 were selected for further detailed study.

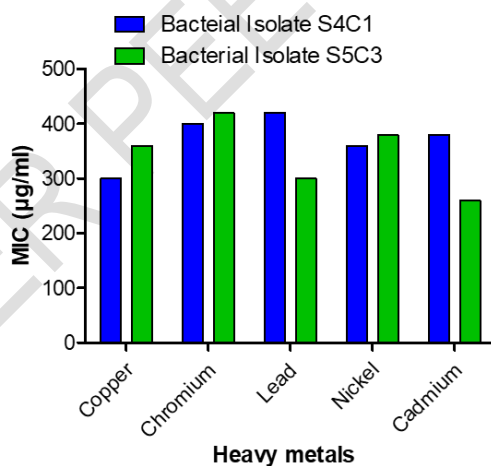


Figure 2: Graphical representation of the minimum inhibitory concentrations values ($\mu\text{g/ml}$) of heavy metal-resistant bacterial isolates.

Table 1. Minimal inhibitory concentrations ($\mu\text{g/ml}$) of bacterial isolates S4C1 and S5C3

| Metal compound | MIC ($\mu\text{g/ml}$) |
|----------------|--------------------------|
|----------------|--------------------------|

| | Bacterial Isolate S4C1 | Bacterial Isolate S5C3 |
|----------|------------------------|------------------------|
| Copper | 300 | 360 |
| Chromium | 400 | 420 |
| Lead | 420 | 300 |
| Nickel | 360 | 380 |
| Cadmium | 380 | 260 |

Effect of pH and temperature on growth of heavy metal tolerant bacterial isolates: The pH and temperature parameters in the medium were optimized for bacterial isolates S4C1 and S5C3 growth conditions. Nutrient broth (NB) medium was used for the optimization study. In the flasks with 200 ml of sterilized NB medium, a bacterial strain was cultivated at various pH (3-12) and temperature (10 °C - 65 °C) conditions. According to the results of the optimization study, the bacterial isolates S4C1 & S5C3 grew best at a pH of 7 similarly Kalaimurugan et al., [41] founds the optimum growth at pH 7-9 and a temperature of 35 °C, similarly Moghannem [42] and Sahin & Ozturk [43] found best result at 35°C as shown in figure 3 and figure 4.

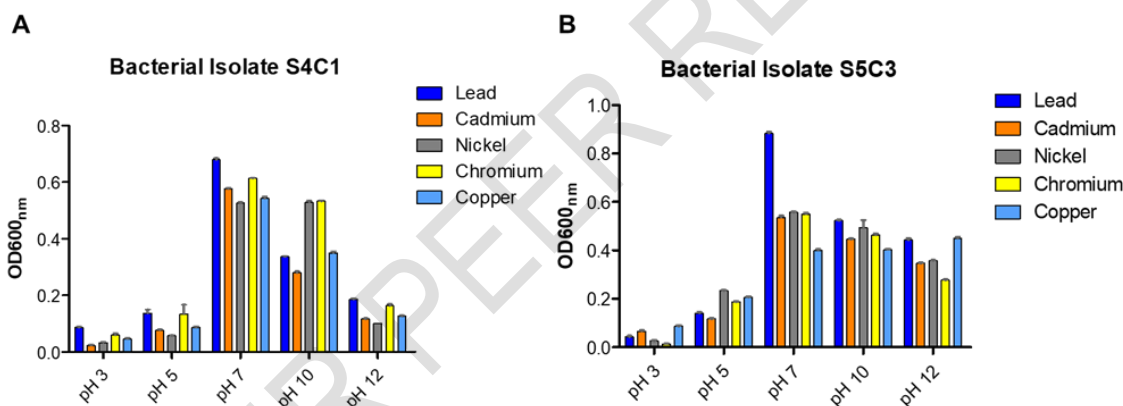


Figure 3: Graphical representation of heavy metal resistance bacteria isolates (A) S4C1 and (B) S5C3 growth at different pH.

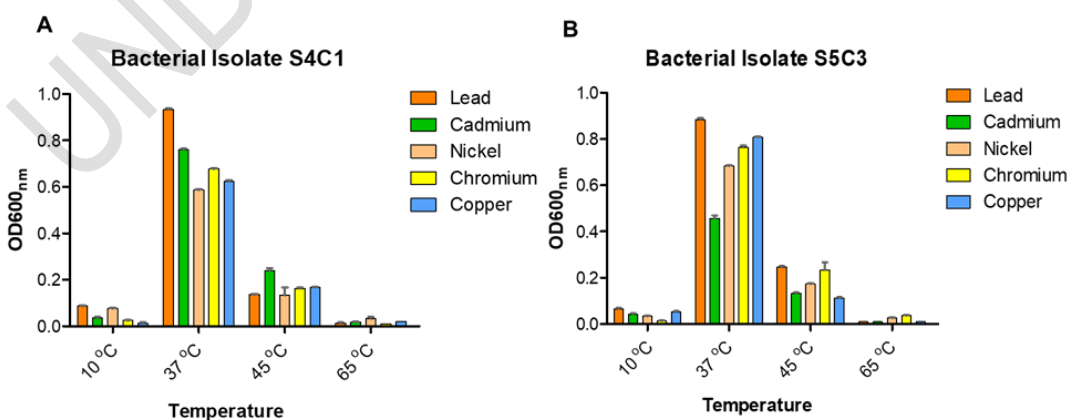


Figure 4: Graphical representation of heavy metal resistance bacteria isolates (A) S4C1 and (B) S5C3 growth at different temperatures.

Heavy metal biodegradability study: Following the optimization of the growth settings, the biodegradability of the relevant heavy metal was examined by Atomic Absorption Spectroscopy (AAS) for the selected bacterial isolates [44-46]. In order to determine the concentration of the heavy metal, a sample of the microbial growth in Nutrient Broth (NB) medium supplemented with heavy metals at pH 7.0 and 35 °C for 48 hours was taken. The results were then reviewed, and the results shown in Figure 5.

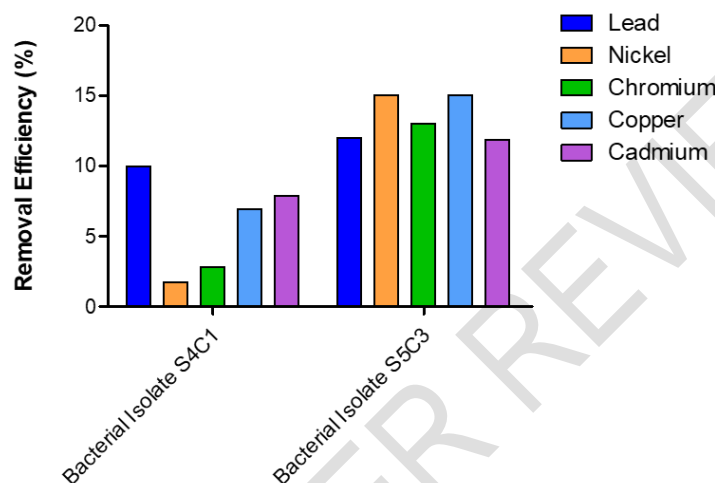


Figure 5: Graphical representation of percentage removal efficiency of heavy metals through Atomic Absorption Spectroscopy (AAS).

Morphological and biochemical characterization of heavy metal-tolerant bacterial isolates: The morphological characteristics of bacterial isolates S4C1 and S5C3 were studied under the microscope using Gram staining technique. Gram staining is a method used to differentiate two types of bacteria (gram-positive and gram-negative) based on their different cell wall components. Table 2 displays the results of morphological and biochemical characteristics.

Table 2: Morphological and biochemical analysis of bacterial isolates.

| Tests | Bacterial Isolate S4C1 (<i>Comamonastestosteroni</i>) | Bacterial Isolate S5C3 (<i>Bacillus cereus</i>) |
|---------------------------|--|--|
| Gram staining | Gram negative | Gram positive |
| Endospore staining | Negative | Positive |
| Catalase test | Positive | Positive |
| Glucose fermentation test | Negative | Positive |
| Mannitol test | Positive | Positive |
| Indole test | Negative | Negative |
| MR test | Positive | Positive |
| VP test | Negative | Negative |

Molecular characterization of heavy metal tolerant bacterial isolates: Bacterial isolates were identified by 16S rRNA gene sequencing to validate morphological and biochemical characterization. Isolated genomic DNA was amplified by 16S primers (Figure 6) and 16S rRNA gene sequencing was performed. Bacterial isolates S4C1 and S5C3 showed a high level (99–100%) similarity with known sequences in the NCBI database when compared using the Basic Local Alignment Search Tool (BLAST). This similarity was based on multiple sequence alignment by CLUSTALW along the branch length of the 16S rRNA sequence to create a phylogenetic tree (Figure 7). The MEGA 7.0 programme was used to create a Maximum Likelihood (ML) phylogenetic tree to determine the relationship between our isolates and other reference strains. Based on the first five identities, with nucleotide and amino acid similarities, respectively, the multiple heavy metal resistant bacteria isolates were identified as S4C1 is *Comamonas testosteroni* and S5C3 is *Bacillus cereus*. *Comamonas testosteroni* is aerobic gram-negative environmental bacteria that is able to break down estrogens and other sterols like ergosterol. *Bacillus cereus* is a Gram-positive bacterium commonly found in soil, food, and sea sponges. These also have the nature of forming spores, which can cause food borne diseases.

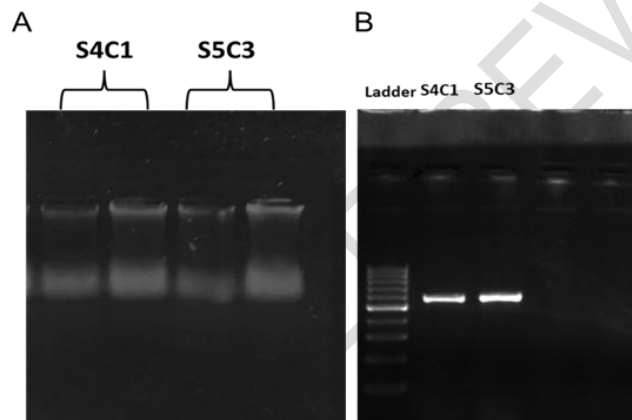


Figure 6: A. Isolated genomic DNA Bands on 0.7% Agarose gel, B. Amplified PCR product of 16sRNA.

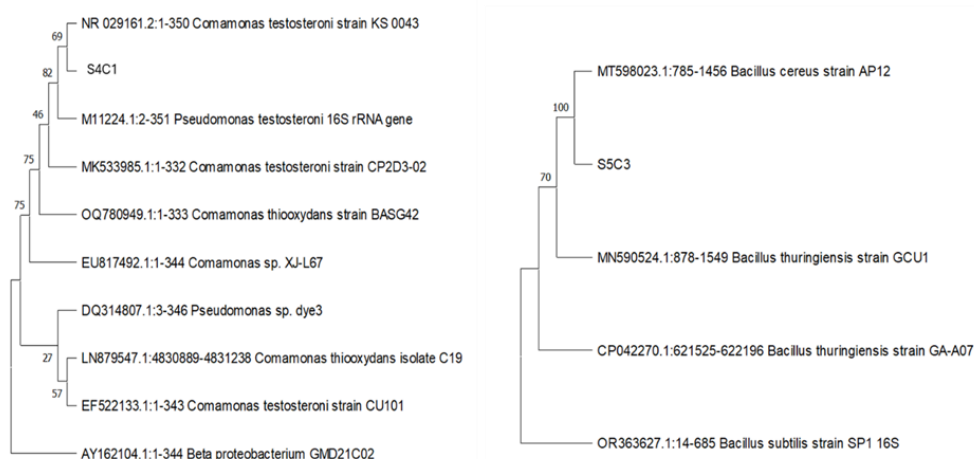


Figure 7: Neighbor joining tree showing the phylogenetic relationship, based on 16S rRNA

gene sequences, between the reference bacteria and the identified heavy metal-resistant bacterial isolates S4C1 and S5C3.

Identification of stress proteins in the presence of heavy metals: Bacteria were grown in NB media in the presence of heavy metals lead (Pb), copper (Cu), chromium (Cr), cadmium (Cd), and nickel (Ni). The results revealed the presence of heavy metal specific stress proteins in the treated groups (Table 3). The following results were obtained when analyzing the protein profiles via SDS-PAGE (Figure 9) and Identification by MALDI-TOFMS (matrix-assisted laser desorption/ionization time-of-flight mass spectrometry) as shown in Figure 10.

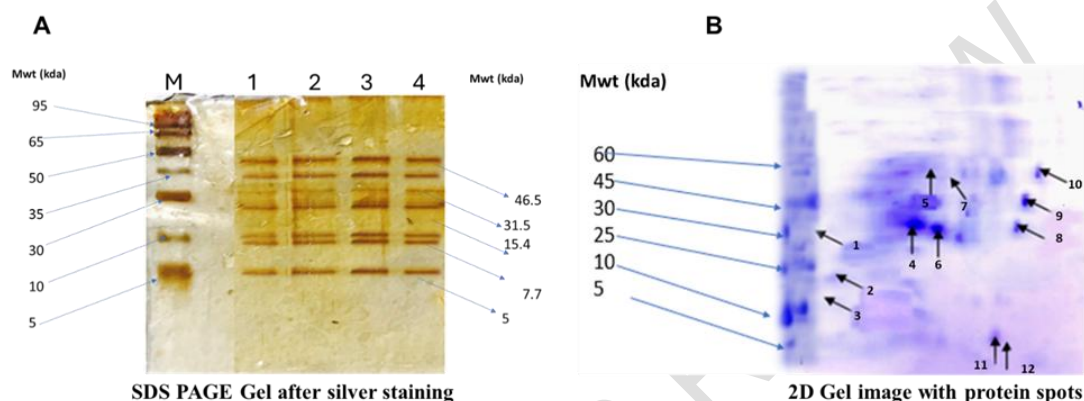


Figure 9: Protein profiles of bacterial isolate S4C1 (1, 2) and S5C3 (3, 4) via (A) SDS page electrophoresis and (B) 2D Gel electrophoresis

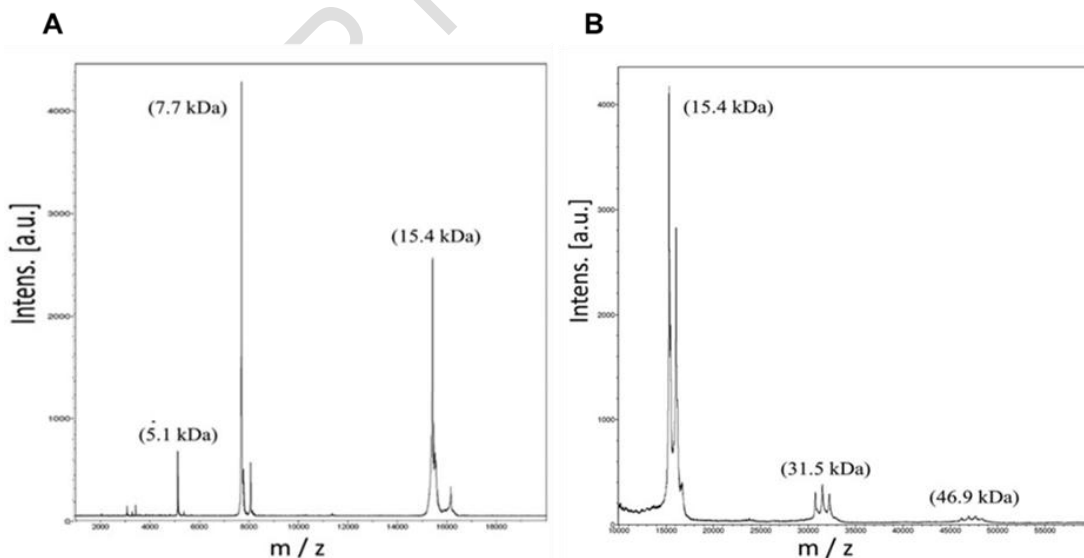


Figure 10: Representative MALDI-TOF MS chromatogram of (A) Bacterial isolate S4C1, (B) Bacterial isolate S5C3

Table 3: Characteristics of the differentially expressed proteins.

| Spot no. | Mol. Wt. (kDa) | pI | Probable protein name |
|----------|----------------|------|----------------------------|
| 1 | 5.1 | 5.44 | S layer protein |
| 2 | 7.7 | 4.93 | F0F1ATP synthase subunit b |
| 4 | 15.4 | 5.64 | Flagellin |
| 5 | 31.5 | 9.95 | 50S ribosomal protein L4 |
| 6 | 46.9 | 6 | Molecular chaperone |

The ion trapping function of the S-layer proteins allows the cells to withstand alterations in a dynamic environment. It was discovered that the S-layer proteins had uranium binding properties, thereby shielding the organism from the metal's harmful effects. It is well known that the S-layer proteins include several amino acids with carboxylic side chains, which likely facilitate metal binding. When the S-layer proteins were removed from *Bacillus thuringiensis*, the organism's ability to biosorb copper decreased. According to research by Gerbino et al. [47], the S-layer proteins in *Lactobacillus kefir* operate as a protective layer when exposed to lead. The isolated organism's increased production of S-layer proteins in response to metal exposure likely aids it in sequestering metals and inhibiting their entrance [48].

Five different types of subunits make up the F1 ATPase, with the b subunit being one of them and being present in three copies. Three catalytic sites are present in the F1 ATPase's b subunit, which is essential for the production of ATP. Four species that were able to withstand radiation exposure showed increased expression of the gene encoding the b subunit. The necessity for greater energy metabolism, which could lead to the creation of extra proteins to combat stress, is most likely implied by the increased expression of the b subunit under stressful conditions [49].

The 50S ribosomal L4 protein was overexpressed in this study when compared to the control group after exposure to metals. The 50S ribosomal L4 protein is a rRNA-binding protein from the L4 family that interacts with the 23S rRNA of the ribosome's 50S subunit several times. The processes of transcription and translation are likely aided by these proteins. The speed of bacterial growth may slow down under stressful circumstances. Starved microorganisms have slower growth rates and produce less rRNA, tRNA, and proteins. When under stress, the organism likely produces the ribosomal protein to speed up protein synthesis and provide a more hospitable environment for bacteria to proliferate. In times of duress, the L4 protein's protective properties may have served specialized functional purposes in the isolated organism to reduce metal stress [50].

Chromium and cadmium stress do not cause the expression of molecular chaperones (GroEL), but copper stress does. Certain metals can particularly trigger certain proteins. According to research, molecular chaperones are known to assist in the folding of freshly synthesized proteins and the refolding of proteins that have been changed under stress, protecting cells from harm. Chaperonins enclose polypeptide chains and aid folding under environmental stress circumstances until the hydrophobic portions are buried. Under conditions of copper stress, the isolated organism expressed the stress protein GroEL, likely to facilitate protein folding. The molecular chaperones are also produced extracellularly in some bacterial secretion systems to help fold the unfolded proteins that are transported across the membrane [51].

Chromium and copper resistance appear to be influenced by flagellin expression. Flagellin is secreted to assist the organism in moving to the best colonisation sites, avoiding toxins, and

boosting nutrition uptake. Flagellin overexpression in *B. cereus* CMG2K4 has been linked to nickel and cadmium tolerance in *Pseudomonas* species. According to the current study, metals are likely to affect flagellin expression in a way that promotes motility away from harmful environments. The *flhA* protein, a part of the flagellar type 3 secretion system, transports flagellin proteins, suggesting that flagellin may also have a role outside of the bacterial cell [52].

4. CONCLUSION

Given the abundance of heavy metals in industrial effluents, the bacteria living there would likely be resistant to heavy metals. The present investigation was initiated with the intention to isolate and identify heavy metal, Pb, Cu, Cr, Cd and Ni resistant bacteria from industrial waste contaminated sites in Uttar Pradesh. *Comamonastestosteronei* and *Bacillus cereus* were isolated and identified as heavy metal resistance isolates. Both isolates display high levels of resistance to Pb, Cu, Cr, Cd and Ni. The pattern of protein expression when various heavy metals were present was intriguing. It was found that certain proteins were overexpressed in the case of heavy metals. To fully comprehend the protein underlying the production of these stress proteins and their contribution to heavy metal tolerance, more study is required. The overall study demonstrates that bacterial isolates have high levels of metal tolerance; as a result, these isolates can be employed in the bioremediation of effluents from heavy metal-handling enterprises.

CONSENT

Not applicable.

REFERENCES

1. Ali, M. M., Hossain, D., Al-Imran, A., Khan, M. S., Begum, M., & Osman, M. H. (2021). Environmental pollution with heavy metals: A public health concern. *Heavy Metals-Their Environmental Impacts and Mitigation*, 771-783.
2. Raghunath, B. V., Punnaigaiarasi, A., Rajarajan, G., Irshad, A., Elango, A., & Mahesh Kumar, G. (2016). Impact of dairy effluent on environment—a review. *Integrated Waste Management in India: Status and Future Prospects for Environmental Sustainability*, 239-249.
3. Elbehiry, F., Alshaal, T., Elhawati, N., & Elbasiouny, H. (2021). Environmental-Friendly and Cost-Effective Agricultural Wastes for Heavy Metals and Toxicants Removal from Wastewater. In *Cost-efficient Wastewater Treatment Technologies: Natural Systems* (pp. 107-127). Cham: Springer International Publishing.
4. Velusamy, S., Roy, A., Sundaram, S., & Kumar Mallick, T. (2021). A review on heavy metal ions and containing dyes removal through graphene oxide-based adsorption strategies for textile wastewater treatment. *The Chemical Record*, 21(7), 1570-1610.

5. Bánfalvi, G. (2011). Heavy metals, trace elements and their cellular effects. Cellular effects of heavy metals, 3-28.
6. Pandey, G., & Madhuri, S. (2014). Heavy metals causing toxicity in animals and fishes. Research Journal of Animal, Veterinary and Fishery Sciences, 2(2), 17-23.
7. Chovanová, K., Sládeková, D., Kmet, V., Proksova, M., Harichová, J., Puskarova, A., ... & Ferienc, P. (2004). Identification and characterization of eight cadmium resistant bacterial isolates from a cadmium-contaminated sewage sludge. Biologia, 59(6), 817-827.
8. Raj, K., & Das, A. P. (2023). Lead pollution: Impact on environment and human health and approach for a sustainable solution. Environmental Chemistry and Ecotoxicology.
9. Duruibe, Ogwuegbu, & Egwurugwu. (2007). Heavy metal pollution and human biotoxic effects. International Journal of physical sciences, 2(5), 112-118.
10. Mishra, S., Bharagava, R. N., More, N., Yadav, A., Zainith, S., Mani, S., & Chowdhary, P. (2019). Heavy metal contamination: an alarming threat to environment and human health. Environmental biotechnology: For sustainable future, 103-125.
11. Khanna, P. (2011). Assessment of heavy metal contamination in different vegetables grown in and around urban areas. Research journal of environmental toxicology, 5(3), 162.
12. Chung, D. Y., Kim, H. I., Chung, Y. H., Lee, M. J., Yoo, S. J., Bokare, A. D., ... & Sung, Y. E. (2014). Inhibition of CO poisoning on Pt catalyst coupled with the reduction of toxic hexavalent chromium in a dual-functional fuel cell. Scientific reports, 4(1), 7450.
13. Jaishankar, M., Tseten, T., Anbalagan, N., Mathew, B. B., & Beeregowda, K. N. (2014). Toxicity, mechanism and health effects of some heavy metals. Interdisciplinary toxicology, 7(2), 60.
14. Khan, F. S. A., Mubarak, N. M., Tan, Y. H., Khalid, M., Karri, R. R., Walvekar, R., ... & Mazari, S. A. (2021). A comprehensive review on magnetic carbon nanotubes and carbon nanotube-based buckypaper for removal of heavy metals and dyes. Journal of Hazardous Materials, 413, 125375.
15. Rahman, Z., & Singh, V. P. (2019). The relative impact of toxic heavy metals (THMs)(arsenic (As), cadmium (Cd), chromium (Cr)(VI), mercury (Hg), and lead (Pb)) on the total environment: an overview. Environmental monitoring and assessment, 191, 1-21.
16. Kasan, H. C., & Baecker, A. A. (1989). An assessment of toxic metal biosorption by activated sludge from the treatment of coal-gasification effluent of a petrochemical plant. Water Research, 23(7), 795-800.
17. del Carmen Gómez-Regalado, M., Martín, J., Santos, J. L., Aparicio, I., Alonso, E., & Zafra-Gómez, A. (2023). Bioaccumulation/bioconcentration of pharmaceutical active compounds in aquatic organisms: Assessment and factors database. Science of The Total Environment, 861, 160638.
18. Ramesh, B., Saravanan, A., Kumar, P. S., Yaashikaa, P. R., Thamarai, P., Shaji, A., & Rangasamy, G. (2023). A review on algae biosorption for the removal of hazardous

pollutants from wastewater: Limiting factors, prospects and recommendations. *Environmental Pollution*, 121572.

19. Teng, Z., Hou, F., Bai, M., Li, J., Wang, J., Wu, J., ... & Guo, H. (2022). Bio-mineralization of virus-like particles by metal–organic framework nanoparticles enhances the thermostability and immune responses of the vaccines. *Journal of Materials Chemistry B*, 10(15), 2853-2864.

20. Fashola, M. O., Ngole-Jeme, V. M., & Babalola, O. O. (2016). Heavy metal pollution from gold mines: environmental effects and bacterial strategies for resistance. *International journal of environmental research and public health*, 13(11), 1047.

21. Feng, G., Xie, T., Wang, X., Bai, J., Tang, L., Zhao, H., ... & Zhao, Y. (2018). Metagenomic analysis of microbial community and function involved in cd-contaminated soil. *BMC microbiology*, 18, 1-13.

22. Zaki, S., & Farag, S. (2010). Isolation and molecular characterization of some copper biosorped strains. *International Journal of Environmental Science & Technology*, 7, 553-560.

23. Masi, C., Gemechu, G., & Tafesse, M. (2021). Isolation, screening, characterization, and identification of alkaline protease-producing bacteria from leather industry effluent. *Annals of Microbiology*, 71, 1-11.

24. Rahman, M. S., Islam, M. R., Mondol, O. K., Rahman, M. S., Sabrin, F., & Zohora, U. S. (2018). Screening of protease producing bacteria from tannery wastes of leather processing industries at Hazaribag, Bangladesh. *Jahangirnagar University Journal of Biological Sciences*, 7(1), 23-34.

25. Bergey, D. H. (1994). *Bergey's manual of determinative bacteriology*. Lippincott Williams & Wilkins.

26. Reyes, A. T. (2018). Morpho-biochemical aided identification of bacterial isolates from Philippine native pig. *Adv. Pharmacol. Clin. Trials*, 3(5), 000148.

27. Devulder, G., Perriere, G., Baty, F., & Flandrois, J. P. (2003). BIBI, a bioinformatics bacterial identification tool. *Journal of clinical microbiology*, 41(4), 1785-1787.

28. Pandit, R. J., Patel, B., Kunjadia, P. D., & Nagee, A. (2013). Isolation, characterization and molecular identification of heavy metal resistant bacteria from industrial effluents, Amala-khadi-Ankleshwar, Gujarat. *International Journal of Environmental Sciences*, 3(5), 1689.

29. Tang, Y. W., Stratton, C. W., & Han, X. Y. (2006). Bacterial identification based on 16S ribosomal RNA gene sequence analysis. *Advanced techniques in diagnostic microbiology*, 323-332.

30. Filali, B. K., Taoufik, J., Zeroual, Y. F. A. Z., Dzairi, F. Z., Talbi, M., & Blaghen, M. (2000). Waste water bacterial isolates resistant to heavy metals and antibiotics. *Current microbiology*, 41, 151-156.

31. Rajbanshi, A. (2008). Study on Heavy Metal Resistant Bacteria in Guheswori Sewage Treatment Plant. *Our nature*, 6(1).

32. Muccee, F., & Ejaz, S. (2020). Characterization of Multi-Potential Toluene Metabolizing Bacteria Isolated from Tannery Effluents. *Microbiology*, 89, 626-636.
33. Çoban, E. P., & Biyik, H. (2011). Evaluation of different pH and temperatures for bacterial cellulose production in HS (Hestrin-Scharmm) medium and beet molasses medium. *African Journal of microbiology research*, 5(9), 1037-1045.
34. Yu, B., Roy Choudhury, M., Yang, X., Benoit, S. L., Womack, E., Van Mouwerik Lyles, K., ... & Wang, B. (2022). Restoring and enhancing the potency of existing antibiotics against drug-resistant gram-negative bacteria through the development of potent small-molecule adjuvants. *ACS Infectious Diseases*, 8(8), 1491-1508.
35. Olaniran, A. O., Balgobind, A., & Pillay, B. (2013). Bioavailability of heavy metals in soil: impact on microbial biodegradation of organic compounds and possible improvement strategies. *International journal of molecular sciences*, 14(5), 10197-10228.
36. Link, A. J. (Ed.). (2008). 2-D proteome analysis protocols (Vol. 112). Springer Science & Business Media.
37. Joshi, B. H., & Modi, K. G. (2013). Screening and characterization of heavy metal resistant bacteria for its prospects in bioremediation of contaminated soil. *Journal of Environmental Research and Development*, 7(4A), 1531.
38. Zahoor, A., & Rehman, A. (2009). Isolation of Cr (VI) reducing bacteria from industrial effluents and their potential use in bioremediation of chromium containing wastewater. *Journal of Environmental Sciences*, 21(6), 814-820.
39. Bhardwaj, R., Gupta, A., & Garg, J. K. (2018). Impact of heavy metals on inhibitory concentration of *Escherichia coli*—a case study of river Yamuna system, Delhi, India. *Environmental monitoring and assessment*, 190(11), 674.
40. Hassen, A., Saidi, N., Cherif, M., & Boudabous, A. (1998). Effects of heavy metals on *Pseudomonas aeruginosa* and *Bacillus thuringiensis*. *Bioresource Technology*, 65(1-2), 73-82.
41. Kalaimurugan, D., Balamuralikrishnan, B., Durairaj, K., Vasudhevan, P., Shivakumar, M. S., Kaul, T., ... & Venkatesan, S. (2020). Isolation and characterization of heavy-metal-resistant bacteria and their applications in environmental bioremediation. *International Journal of Environmental Science and Technology*, 17, 1455-1462.
42. Moghannem, S. A., Refaat, B. M., El-Sherbiny, G. M., El-Sayed, M. H., Elsehemy, I. A., & Kalaba, M. H. (2015). Characterization of heavy metal and antibiotic-resistant bacteria isolated from polluted localities in Egypt. *Egyptian Pharmaceutical Journal*, 14(3), 158-165.
43. Şahin, Y., & Öztürk, A. (2005). Biosorption of chromium (VI) ions from aqueous solution by the bacterium *Bacillus thuringiensis*. *Process Biochemistry*, 40(5), 1895-1901.
44. Amor, L., Kennes, C., & Veiga, M. C. (2001). Kinetics of inhibition in the biodegradation of monoaromatic hydrocarbons in presence of heavy metals. *Bioresource technology*, 78(2), 181-185.
45. Marzuki, I., Daris, L., Nisaa, K., & Emelda, A. (2020, October). The power of biodegradation and bio-adsorption of bacteria symbiont sponges sea on waste contaminated

of polycyclic aromatic hydrocarbons and heavy metals. In IOP Conference Series: Earth and Environmental Science (Vol. 584, No. 1, p. 012013). IOP Publishing.

46. Olaniran, A. O., Balgobind, A., & Pillay, B. (2013). Bioavailability of heavy metals in soil: impact on microbial biodegradation of organic compounds and possible improvement strategies. *International journal of molecular sciences*, 14(5), 10197-10228.

47. Gerbino, E., Carasi, P., Mobili, P., Serradell, M. A., & Gómez-Zavaglia, A. (2015). Role of S-layer proteins in bacteria. *World Journal of Microbiology and Biotechnology*, 31, 1877-1887.

48. Overton, K. W., Park, D. M., Yung, M. C., Dohnalkova, A. C., Smit, J., & Jiao, Y. (2016). Two outer membrane proteins contribute to *Caulobacter crescentus* cellular fitness by preventing intracellular S-layer protein accumulation. *Applied and Environmental Microbiology*, 82(23), 6961-6972.

49. Menz, R. I., Walker, J. E., & Leslie, A. G. (2001). Structure of bovine mitochondrial F1-ATPase with nucleotide bound to all three catalytic sites: implications for the mechanism of rotary catalysis. *Cell*, 106(3), 331-341.

50. Fessler, M., Gummesson, B., Charbon, G., Svenningsen, S. L., & Sørensen, M. A. (2020). Short-term kinetics of rRNA degradation in *Escherichia coli* upon starvation for carbon, amino acid or phosphate. *Molecular Microbiology*, 113(5), 951-963.

51. Costa, T. R., Felisberto-Rodrigues, C., Meir, A., Prevost, M. S., Redzej, A., Trokter, M., & Waksman, G. (2015). Secretion systems in Gram-negative bacteria: structural and mechanistic insights. *Nature Reviews Microbiology*, 13(6), 343-359.

52. Matilda, C. S., Mannully, S. T., Viditha, R. P., & Shanthi, C. (2019). Protein profiling of metal-resistant *Bacillus cereus* VITSH1. *Journal of applied microbiology*, 127(1), 121-133.