Original Research Article

Effects of melatonin priming on seed germination of wheat under salt stress

ABSTRACT: Melatonin (N-acetyl-5-methoxytryptamine) has emerged as a new growth regulator in plants due to its positive role in alleviation of a biotic stresses. The experiment was conducted at the Wheat Cultivation Department Laboratory, Gezira Scheme, Sudan. investigate the effects of melatonin priming in alleviating the negative effect of salt stress on seed germination in two cultivars of winter wheat (Triticum aestivum L. cv Elnilein and Imam). Six concentrations were used: control, salt (300 mM NaCl) and four concentrations of melatonin (M) 10, 100, 500 and 1000 µM with salt (300 mM NaCl) add to each concentration: CT, ST, M1+ST, M2+ST, M3+ST, M4+ST. The treatments were arranged in a Randomized Complete Block Design with three replications. The parameters studied included: seed germination rate (%), germination index, mean germination time (d), lengths (cm) and dry weight (mg) of coleoptile and radicle, starch, soluble sugar, and sucrose contents, amylase activities, activities of the antioxidant enzymes including superoxide dismutase (SOD), peroxidase (POD), ascorbate peroxidase (APX) and catalase (CAT), reactive oxygen species (ROS) including malondialdehyde (MDA), Hydrogen peroxide (H₂O₂) contents and superoxide (O_2^-) production rate, abscisic acid (ABA). The parameters of seedling establishment were: Shoot height, root length, fresh and dry weight of shoot and root. The treatment M3+ST (500 µM Melatonin + 300 mM NaCl) had the highest seed germination rate, germination index, lengths and dry weights of coleoptile and radicle,

soluble sugar, sucrose contents, amylase and antioxidant enzymes activities, and ABA content, seedling shoot and root length, and fresh and dry weight of shoot and root in Imam and Elnilein cultivars, respectively. The study revealed that the melatonin priming effectively alleviates salt stress during seed germination and seedling establishment in winter wheat.

Key words: Salt stress, Melatonin, Germination, Antioxidant system, Winter wheat (*Triticum aestivum* L.)

1. Introduction

Amongst the various a biotic stresses that plants convene, heat and salt stress are mainly imperative in agriculture since both stresses can lead to considerable reduction in crop yields and are predictable to raise in incidence in the near future because of climate change [1]. Presently about 6% of total land area or 20% agricultural total lands are influenced by salt in Sudan [2]. Seed germination and seedling establishment are the two critical stages for the crops establishment [3], and these were to harmfully affect by salt stress [4]. Salinity causes reduced emergence and impaired seedling growth presumably due to salt-induced osmotic effects, specific ion toxicity, nutrient imbalance, oxidative damage, and alterations in endogenous levels of hormones [5]. In this admiration [6] confirmed that increases in salt content reduces the germination percentage and delays germination time.

Salinity can influence the seeds germination by the toxic effect of sodium and chloride ions on viability of embryo [7] and [8]. The toxic effects include disruption to the structure of enzymes and other macromolecules, damage to cell organelles and the plasma membrane, the disruption of respiration, photosynthesis and protein synthesis [9], [8] and [10]. accumulation of mono di-Aldehyde (MDA) in different crops [11]. MDA

content is a signal of membrane injure at the cellular levels in salt stress. Hence, the concentration of MDA can serve as an essential oxidative stress sign [12]. Oxidative stress is the term utilized to define the imbalance between production of ROS and scavenging or detoxification [13]. Plants have developed the antioxidant defense system to reduce the harmful impact of ROS like antioxidant enzymes include superoxide dismutase (SOD), peroxidase (POD) catalase (CAT), glutathione reductase (GR) and ascorbate peroxidase (APX) but salt stimulate ROS production exceeds capacity of antioxidant in the cells, oxidative stress arises [14] and [15].

Melatonin (*N*-acetyl-5-methoxytryptamine) has comprehensively be known as an important hormone of animal occupied in many biological processes [16]. However, its role in animals the same like a hormone has been well-characterized, it's information in plants kingdom is still bitty (Arnao and Hernández-Ruiz 2014). Melatonin is considered as a growth regulator of plant, and has role has as that of growth regulation of the shoots, root and explants, germination of seed, rhizogenesis and remain senescence in leaf [17]. The melatonin mechanisms mediated tolerance of stress occupy the antioxidant support biosynthesis, the start of connected enzymes and the directly ROS scavenging subsequent the plants exposure to harm environments [18] and [19]. Previous study has reported that while applied exogenously melatonin alleviates oxidative injury through ROS scavenging and stimulating antioxidant system that are in different environmental stresses [20] and [21].

In view of the above information, this study focused on determining the effect of melatonin pretreatment on seed germination, physiological, and biochemical changes happening in winter wheat geminating seeds exposed to salt stress.

2. Materials and methods

2.1 Experimental design

The study was performed in the wheat cultivation department Laboratory, Gezira scheme, Gezira state, Sudan Uniform seeds of two cultivars of winter wheat (Triticum aestivum L.) cv Elnilein and Imam) were chosen and surface- sterilized with 2.5% sodium hypochlorite for 10 min and washed seven times with sterile distilled water [22]. Two treatments were placed with soaking the seeds in distilled water as control and 10, 100, 500, and 1000 µM of melatonin solution for 20 h, respectively. The concentrations of melatonin used here were established to be optimal in the preliminary experiments. The soaked 50 seeds of each cultivar were arranged on a filter paper in petri dishes with a diameter of 9 cm, and 300 mM of NaCl was then added in to each dish. The dishes were incubated at 22± 0.5 °C for 7 days in Chamber. At the two-leaf stage (ca. 20 days after germination), wheat plants were transplanted to plastic containers (45 cm in length, 35 cm in width and 18 cm in height). The density was 30 plants in each container. The Hoagland nutrient solution was used for hydroponic culture. The growth temperatures were $25~^{0}\text{C}/20~^{0}\text{C}$ (day/night), six treatments were used: Control (CT), 300 mM of salt NaCl (ST) , 10 μM of melatonin+300 mM NaCl (M1+ST) , 100 μM of melatonin+300 mM NaCl (M2+ST), 500 µM of melatonin+300 mM NaCl (M3+ST), 1000 µM of melatonin+300 m M NaCl (M4+ST). The experiment was a Randomized Complete Block Design with three replications.50 seed were used in each treatment for the germination test for each cultivar and were sampled at days 2, 3, 4, 5 and 7 after treatment for biochemical and physiological measurements, and another batch was sampled at day 7 after treatment for determinations of dry weight, seed germination rate, and ABA content. At the four-leaf stage,

wheat seedlings were harvested for measurements of seedling shoot height, root length, fresh and dry weights of shoot and root.

2.2 Germination rate (GR), mean germination time (MGT) germination index (GI), dry weight of germinating seeds

GR is the proportion of germinated seeds in each Petri-dish during 7 days after germination. MGT was determined fallowing the method of (Ellis and Roberts 1978), i.e. $MGT = \sum (f \ x \ i) / \sum f$, where, f is the number of recently germinated seeds on Germinating Day i. Subsequently, long MGT indicates slow germination. Germinated seeds percentage on each day was measured. The method of [23] was used to calculate the GI as $GI = \sum (G_i / T_i)$, where Gi is the germination percentage per day, and Ti is day of germination test. High GI demonstrates high quality of seed and performance [23.

Radicles, coleoptile and the seed residues of germinating seeds of two cultivars were selected and dried at 105 0 C for 2 h, after that kept at 80 0 C to obtain stable weight.

2.3 Contents of starch and soluble sugars

Germinating seed oven- dried powder (1 g) was mixed by 10 ml 0.33 mM of HCl` and then heated at 100 0 C for 10 min to remove starch. Half ml of ZnSO4 (30 %, w/v) was added to the extraction to remove proteins. After that, 0.5 ml K3 [Fe (CN)₆] (30 %, w/v) was added and mixed, and then the mixture volume was attuned to 20 ml with distilled water. After completely shaking it was filtrated, then the concentration of starch was determined like the amount of glucose produced at 20–25 0 C using the polarimetric analysis through an automatic recording polarimeter (WZZ-2B, Shanghai, China) according to [24]. Moreover, to extract total soluble sugars, 0.1 g oven- dried germinating seed powder sample was homogenized in (80 %) ethanol at 80 0 C for 10 min and centrifugation at

600g. The supernatant was collected for content of soluble sugar and sucrose measurement using the method of anthrone [25].

2.4 Amylase extraction and activity assess

Activity of amylase and extraction were estimated after [26]. The concentration of soluble protein in germinating seeds was estimated by following the method of [27] by using bovine serum albumin (BSA) like standard. Activities of both α - and β -amylase were articulated as unit mg⁻¹ protein, and one unit is equivalent to release of one mg maltose from starch per minute by the amylases.

2.5 Activities of antioxidant enzymes and O_2 - production rate, H_2O_2 , MDA concentrations

The MDA and H₂O₂ content measurements and antioxidant enzymes activities in germinating seed were measured suggested by [28]. In brief, 0.5 g samples of germinating seed were sliced and homogenized in a mortar and pestle with 5 ml of ice-cold extraction buffer (50 mM potassium phosphate buffer, pH 7.0, 0.4 % polyvinylpoly pyrrolidone (PVP)). The homogenate was centrifuged at 10,000xg for 30 min and the supernatant was used like crude extract. All procedures were done at 4 °C. The MDA content was calculated after [29]. In brief, 20 % trichloroacetic acid containing 0.5 % 2-thiobarturic acid was added to 0.2 ml of the suspension and heated at 95°C for 25 min after that cooled down in a cool water bath and centrifuged at 3,000xg for 5 min. Concentration of MDA was calculated at 532 nm and corrected by subtracting the absorbance at 600 nm. The concentrations of H₂O₂ were carried out by the method given by [30]. Briefly, 1 ml suspension was added to 0.1 ml 20 % titanium tetrachloride (TiCl4) and mixed with shaking. 0.2 ml of 25 % ammonia was added to the mixture. The reaction solution was after that centrifuged at 5,930xg for 5 min, and the precipitate was washed three

times with cold acetone to eradicate the color. Lastly, the precipitate was dissolved in 3 ml 1 M H₂SO₄, and filtered prior to measurement of the absorbance at 415 nm. The O ₂ production rate was measured by using the method of [22]. The activities of SOD, and CAT were calculated according to [28]. For SOD activity measurement, 3 ml reaction mixture contained 130 mM methionine, 750 mM NBT, 100 mM EDTA and 50 ml of enzyme extract in 50 mM phosphate buffer (pH 7.8). The reaction was started with 20 mM riboflavin through exposing the cuvette to a 15 W circular white light tube for 10 min. One unit of SOD activity was identified as the amount of enzyme per mg of protein sample causing 50 % inhibition of the rate of NBT reduction at 560 nm [31]. CAT activity was estimated as described by [28]. The activity of POD was determined according to the changes in absorbance at 470 nm due to guaiacol oxidation. Ascorbate peroxidase (APX) was measured using the method of [32].

2.6 Determination of endogenous hormone concentration

The content of abscisic acid (ABA) was analyzed by an indirect ELISA technique. Briefly, the samples of 0.1 g dry seeds were homogenized in liquid nitrogen and removed in cold 80% (v/v) methanol by butylated hydroxytoluene (1 mM) overnight at 4 °C. The extracts were then collected after centrifugation at 10,000 x g (4 °C) for 20 min, and after that the extracts were passed throughout a C₁₈ Sep-Pak catridge (Waters, Milford, MA) and dried in N₂. Then the residues were melted in PBS (0.01 M, pH 7.4). To estimate the content of ABA using the method described by [33].

2.7 Statistical analysis

All data were subjected to the one-way analysis of variance (ANOVA) to determine the significant differences between the treatments using the software of SPSS (Ver. 10.0 SPSS, Chicago, IL, USA).

3. Results

3.1 Seed germination parameters

Seed germination rate was significantly reduced under salt stress. During 7 days of germination, seed germination rate (GR) and germination index (GI) were considerably enhanced whereas mean germination time (MGT) was reduced by melatonin priming treatment when compared with salt stressed-seed in both cultivars, with values being higher in Imam than that in the Elnilein (Table 1). Also, lengths and dry weights of coleoptile and radicle were clearly increased in both cultivars while seed residue dry weight was significantly (P < 0.05) reduced with application of melatonin in both cultivars, as compared with salt stress alone .Therefore, this increase was more prominent in Imam than that in the Elnilein (Table 1). However, seed pretreatment with melatonin it seemed that enhanced seed germination in wheat under salt stress. However, the control treatments performed better in all the germination parameters studied indicating that there is no problem of both the wheat cultivars. Further more if the seeds were subjected to salt stress under natural environments, then the necessity of melatonin pretreatment would be much helpful.

Table 1 Effect of melatonin priming on seed germination rate (GR), germination index (GI), mean germination time (MGT), length of radicle and coleoptile, and dry weight of radicle, coleoptile and whole seed after 7 days germination under salt stress in wheat.

					Length (cm)		Dry weight (mg seed ⁻¹)		
Cultivar	Treatments	GR(%)	GI	MGT(d)	Radicle	Coleoptile	Radicle	Coleoptile	Seed residue
Elnilein	CT	95.36±0.28a	92.78±0.05a	1.54±0.03d	9.38±0.05a	8.27±0.03a	13.18±0.02a	10.26±0.05a	7.13±0.02f
	ST	64.39±0.26e	21.49±0.36f	4.62±0.22a	$0.9\pm0.01f$	$0.82\pm0.01f$	1.39±0.05f	1.12±0.01f	32.36±0.04a
	M1+ST	76.28±0.23d	26.13±0.27e	3.59±0.13b	1.59±0.02e	1.21±0.02e	$2.84\pm0.02e$	2.03±0.01e	25.71±0.15b
	M2+ST	88.47±0.41b	51.47±0.23c	1.87±0.13d	5.23±0.02c	4.78±0.14c	6.35±0.04c	5.01±0.04c	12.45±0.16d
	M3+ST	94.23±0.20a	74.19±0.18b	1.73±0.01d	8.04±0.01b	7.14±0.04b	12.03±0.02b	8.63±0.28b	8.49±0.09e
	M4+ST	81.63±0.46c	46.68±0.33d	2.42±0.04c	3.12±0.01d	3.29±0.03d	4.15±0.01d	3.23±0.01d	$20.18\pm0.04c$
T									
Imam	CT	97.83±0.40a	95.32±0.17a	$1.47 \pm 0.02c$	10.89±0.09a	9.43±0.23a	15.72±0.06a	11.79±0.09a	$9.27 \pm 0.05 f$
	ST	$67.82 \pm 0.37 f$	$25.46 \pm 0.30 f$	$3.48\pm0.08a$	$0.95\pm0.01f$	0.91±0.01f	1.98±0.01f	$1.35\pm0.01f$	$36.55 \pm 0.27a$
	M1+ST	78.53±0.31e	29.38±0.49e	2.92±0.06b	$1.85 \pm 0.02e$	1.98±0.05e	3.94±0.02e	$3.28\pm0.03e$	29.18±0.36b
	M2+ST	91.72±0.35c	69.37±0.90c	1.69±0.02c	7.33±0.03c	5.83±0.05c	9.47±0.18c	7.37±0.19c	13.95±0.03d
	M3+ST	95.57±0.27b	78.67±0.65b	$1.58\pm0.02c$	8.63±0.22b	8.65±0.27b	13.69±0.14b	9.72±0.29b	10.87±0.31e
	M4+ST	82.68±0.67d	55.03±0.63d	2.12±0.07b	5.63±0.08d	4.61±0.04d	7.44±0.04d	5.04±0.01d	23.59±0.04c

Different small letters in the same column refer to significant difference between treatments at P < 0.05 level

3.2 Starch, soluble sugar, and sucrose contents

During seed germination progress starch content decreased, but total soluble sugar and sucrose contents were increased in germinated seeds of wheat in both cultivars (Fig. 1). Nevertheless, in both cultivars, starch content of seed was lower under melatonin priming compared to salt stress during 7 days of germination (Fig. 1). In comparison, the contents of total soluble sugar and sucrose were higher in the control and melatonin than that under salt stress in both cultivars. However, total soluble sugar and sucrose contents were significantly (P < 0.05) higher in Elnilein than those in Imam and oppositely in case of starch content (Fig. 1). These clearly indicated that melatonin priming improved the resistance of wheat seed to salt stress.

3.3 Activities of α - and β - amylase

In control the α -amylase activity in both wheat cultivars were improved in seeds germinating than that in the level of salt stress during 7 days of

germination (Fig. 2). Nevertheless, melatonin priming significantly (P < 0.05) enhanced α -amylase activity, when contrasted with salt stress alone. Furthermore, in both cultivars β -amylase activity decreased under salt stress. On other hand, melatonin pretreatment significantly (P < 0.05) increased the activity of β -amylase compared to salt stress alone (Fig. 2). Conversely, under control and melatonin mutually activities of α - and β -in germinated seeds in both cultivars were higher than those of under salt stress levels, and they increased more in Elnilein than that in Imam.

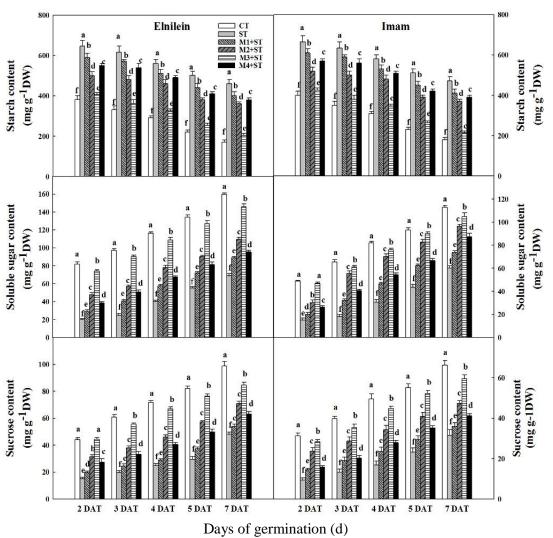


Fig. 1 Effect of melatonin priming on changes in contents of starch, soluble sugar, and sucrose in wheat seeds during 7 days germination under salt stress. Data are means $\pm SD$ (n=3). Different small letters at each day indicate significant difference between treatments at P < 0.05.

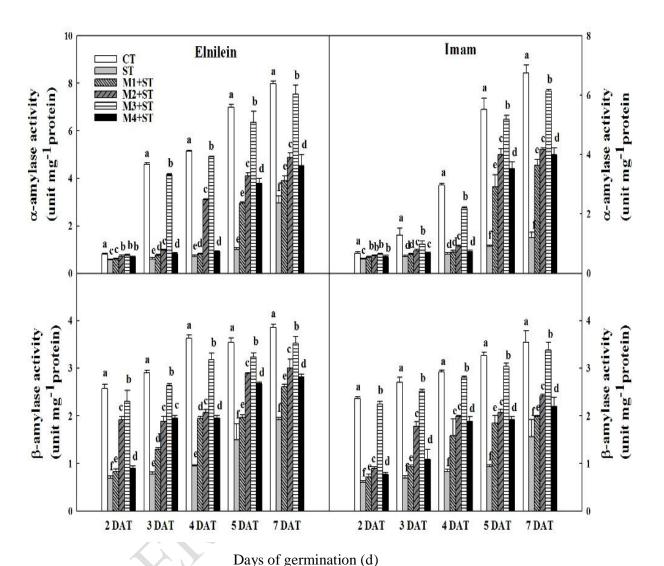


Fig. 2 Effect of melatonin priming on changes in activities of α - and β - amylase in wheat seeds during 7 day's germination under salt stress. Data are means $\pm SD$ (n=3). Different small letters at each day indicate significant difference between treatments at P< 0.05.

3.4 O_2 production rate, concentrations of H_2O_2 and MDA, and activities of antioxidant enzymes

Compared to the control, salt stress significantly (P < 0.05) increased the concentration of MDA and H_2O_2 in wheat germinating seed in both cultivars (Fig.3). In contrast, exogenous application of melatonin significantly (P < 0.05) decreased the concentration of MDA and H_2O_2 in both cultivars compared to salt stress alone. In both cultivars the production rate of O_2 demonstrated the same pattern as similar to H_2O_2 and MDA accumulation during the germinating seeds (Fig. 3). Reversely, salt stress alone significantly (P < 0.05) reduced the activities of SOD, POD, CAT and APX compared to control in both cultivars (Fig.4).In contrast, pretreatment of melatonin remarkably enhanced (P < 0.05) SOD, CAT, POD, and APX activities in both cultivars and they increased more in Elnilein than that in Imam. These results confirmed that priming of melatonin was responsible for the improvement of adaptive responses of wheat seed germination against salt stress.

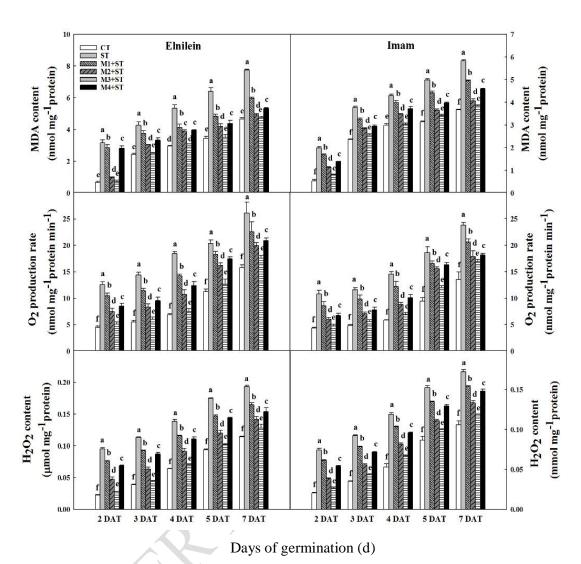


Fig.3 Effect of melatonin priming on changes in contents of malondialdehyde (MDA), hydrogen peroxide (H_2O_2), and $O2^-$ production rate in wheat seeds during 7 days germination under salt stress. Data are means $\pm SD$ (n=3). Different small letters at each day indicate significant difference between treatments at P < 0.05

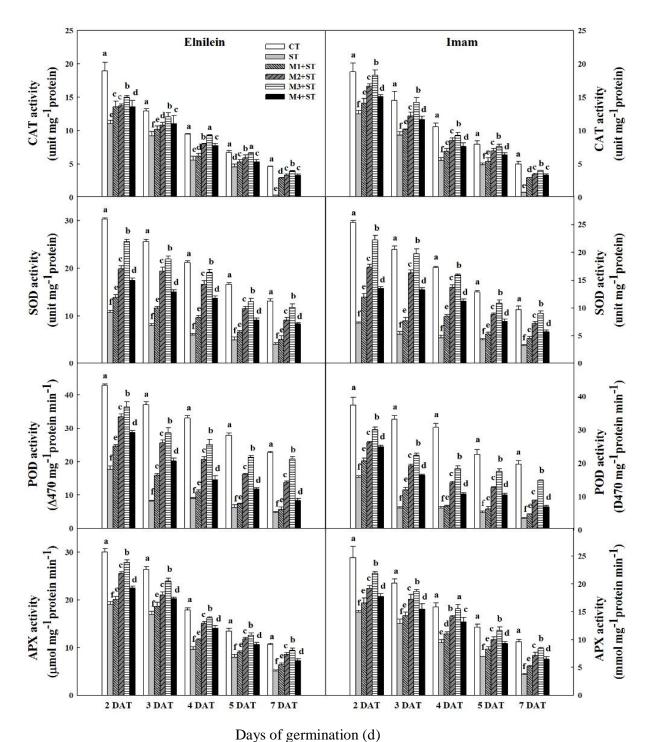


Fig. 4 Effect of melatonin priming on changes in activities of superoxide dismutase (SOD), catalase (CAT), peroxidase (POD), and ascorbate peroxidase (APX) in wheat seeds during 7 days germination under salt stress. Data are means \pm SD (n=3). Different small letters at each day indicate significant difference between treatments at P < 0.05

3.5 ABA concentration

After set of treatments the concentration of ABA in germinating seeds of wheat was much higher under salt stress when compared to control in both cultivars (Fig. 5). On the other hand, under salinity stress melatonin priming significantly (P < 0.05) enhanced germinated seeds ABA content in both cultivars (Fig. 5). In general, melatonin priming had higher levels of ABA in germinated seeds as compared to both control and salt stress alone. The protective effect of melatonin on ABA concentration was greater in Imam than in Elnilein (P < 0.05).

3.6 Seedling growth parameters

After germination, wheat plants were allowed to growing at the same temperature conditions till the 4-leaf stage, for further investigation (Table 2). Salt stress induced by 300 mM NaCl significantly inhibited the growth of wheat seedlings, This attribute was demonstrated by comparing the decrease in the shoot height, root length, fresh and dry weight of shoot and root, in both cultivars. However, melatonins priming significantly (P < 0.05) alleviate this response. The positive effects of melatonin priming had more profound in Imam than that in the Elnilein.

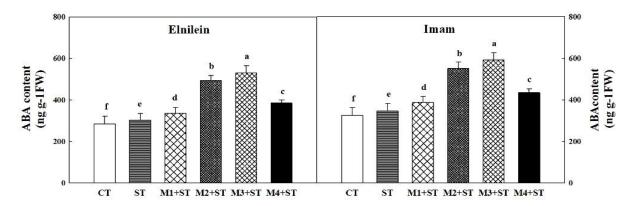


Fig. 5 Effect of melatonin priming on changes in content of ABA in wheat seeds during 7 days germination under salt stress. Data are means \pm SD (n=3). Different small letters indicate significant difference between treatments at P < 0.05

Table 2 Effect of melatonin priming on seedling shoot height and root length, fresh and dry weight of shoot and root of four- leaf-stage seedling grown under salt stress in wheat.

				Fresh weight (mg plant ⁻¹)		Dry weight (mg plant ⁻¹)	
Variety	Treatments	Shoot height (cm)	Root length (cm)	Shoot	Root	Shoot	Root
Elnilein							_
	CT	23.56±1.70a	$26.82 \pm 1.84a$	$23.97 \pm 0.54a$	$16.85 \pm 0.33a$	$6.97\pm0.44a$	$3.96\pm0.37a$
	ST	11.98±0.66f	$13.27 \pm 0.68 f$	$9.81 \pm 0.41 f$	$6.19\pm0.41f$	$2.38\pm0.42f$	$1.03\pm0.42d$
	M1+ST	15.11.±1.45e	17.93±1.54e	$13.21\pm0.39e$	8.88±0.32e	3.92±0.40e	$1.97 \pm 0.42d$
	M2+ST	19.67±0.50c	20.19±0.28c	$17.66 \pm 0.36c$	13.44±0.63c	4.98±0.57c	$2.89\pm0.53b$
	M3+ST	22.44±0.36b	$23.74 \pm 0.84b$	20.13±0.31b	15.67±0.23b	5.11±0.31b	$3.58\pm0.13b$
	M4+ST	$17.22 \pm 0.36d$	$18.87 \pm 0.84d$	$15.23\pm0.31d$	10.16±0.45d	4.09±0.38d	$2.01\pm0.35c$
Imam							
	CT	25.61±1.70a	28.87±1.81a	26.96±0.49a	19.79±0.28a	$8.96\pm0.49a$	$5.89\pm0.48a$
	ST	13.18±1.66f	15.59 ± 0.81 f	12.14±0.19f	7.92±0.45f	$3.56\pm0.49f$	$1.98\pm0.46e$
	M1+ST	16.96±0.78e	$18.24 \pm 1.52e$	16.15±0.56e	$10.53 \pm 0.16e$	$4.32\pm0.57e$	$2.67\pm0.46e$
	M2+ST	$20.73 \pm 0.50c$	$22.43\pm0.42c$	22.22±0.19c	15.91±0.62c	$6.12\pm0.43c$	$4.78\pm0.63c$
	M3+ST	23.47±0.36b	25.91±0.82b	24.42±0.12b	17.19±0.49b	$7.43\pm0.18b$	$5.17\pm0.19b$
	M4+ST	18.23±0.36d	20.34±0.82d	19.21±0.27d	$13.39\pm0.39d$	$5.83\pm0.47d$	$3.32\pm0.39d$

Different small letters in the same column refer to significant difference between treatments at P < 0.05 level

4. Discussion.

Seed germination and plant growth inhibition in saline surroundings were caused by the reduce in the supply of available energy of biochemical essential for synthesis reactions, most important to an inhibition growth of root and coleoptiles [34]. It has been recommended that melatonin acting a significant function in responses of plant to environmental stimulus, including biotic and a biotic stresses [19] and [35]. In this study, salt stress clearly reduced seed germination, growth of root and coleoptiles in both wheat cultivars (Table. 1). But the melatonin priming improved these parameters even under salt stress. These results are in agreement with the results obtained, in barley, bean, and maize [36]. Because melatonin pretreatment was effectual to defend plant cell from oxidative harm [37]. In agreement with this, our results revealed that

melatonin pretreatment significantly enhance germination rate and morphogenesis of radicle and coleoptile under salt stress. This was directly related with the rapid seed starch humiliation during with speedy increase in contents of soluble sugars and sucrose in germinating seeds under salt stress (Fig.1).

Throughout seed germination, different hydrolytic enzymes are created and implicated in reserve mobilization [38]. In harmony with this, our results discovered that salt stress caused extremely inhibition on amylase activity while melatonin use considerably alleviated this inhibition improved the activities of α -amylase and β -amylase in both cultivars (Fig. 2). Hence, melatonin priming assists the starch change into sugars (Fig. 1), which enhancement wheat seed germination under salt stress.

Salt stress stimulates more ROS production, which create injure to macromolecules like, proteins, lipids and nucleic acids [39] and [40], mentioned that salt stress increased the concentration of H₂O₂ and MDA in the tomato mitochondria. This might be caused by a salt-induced raise in the O_2 production rate [41]. In agreement with the previous researches mentioned by [36]. Our study confirmed that salt stress quickly increased the concentration of MDA, H₂O₂, and O₂, in wheat germinating seed. But, there was a significant reduces in these factors upon the exogenous of melatonin as contrasted by salt-stressed wheat germinating seeds in both cultivars (Fig. 3). A similarly stress role of anti-oxidative of melatonin priming was also seeing in salt-stressed barley plants [42]. It is strongly documented that the antioxidant enzymes like SOD, CAT, POD, and APX cooperate an important function in ROS scavenging in salt stressed plants [43]. Previous researches showed that exogenous melatonin defends contents against oxidative stress in reed under heat stress [44] and in wheat under drought. This associated with increased the activities of SOD, CAT, POD and APX. In agreement with these results, we

experiential that activities of these enzymes in the mitochondria of wheat seed were significantly improved by melatonin pretreatment application in the current study (Fig. 4), and which may have provided to the alleviated oxidative stress in the mitochondria of wheat germinating seeds and thus enhanced germination rate under salt stress.

Abscisic acid (ABA)) is essential in the rule of a-amylase through germination [45] and [46] mentioned that ABA plays a role in alleviating oxidative damage under a biotic stress. Abscisic acid (ABA) has been proposed to act as a mediator in plant responses to a range of stresses, including drought and salt stress [47] Among different phytohormones, abscisic acid (ABA), is the central controller of a biotic stress resistance in plants and organizes group of functions [48], allowing plants to cope with various stresses. ABA content considerably increases in plants under stress conditions like drought, extreme temperature, and high salinity, stimulating stress-tolerance effects that help plants adapt and survive under these stressful conditions [49], and is necessary in the plants for stress tolerance [50] and [51], stated that, the level of ABA in soybean seed germination increased significantly under salt stress. It is thus feasible that ABA improves salt tolerance in soybean. Our results also demonstrated that the salt stress increased the ABA content of both cultivars when compared to the control germinating seed. Furthermore, melatonin pretreatment significantly increased seed ABA contents as compared to the salt stress and control during germination under salt stress (Fig.5). Therefore, we can deduce that increasing ABA concentration due to seeds priming with melatonin may be one of the mechanisms that enabled wheat seeds to tolerate salt stress.[52] mentioned that, during the onset and development of salt stress all seedlings growth parameter are affected and reduced when compared with control plants. In this study, we found that values for almost all of the parameters of seedling growth measured were significantly (P < 0.05) decreased in plants under salt stress as contrasted with the control. Nevertheless, melatonin priming application considerably alleviated those inhibitory causes (Table 2). Various studies have established that melatonin can perform as a potential modulator of plant growth and improvement in a dose-dependent way [53]. This could be well exemplified by the increased heights and weights of both shoot and root of seedlings grown under salt stress. This of course was firstly ascribed to the significance of improved seed germination by melatonin pretreatment.

Conclusions

The study clearly demonstrated that the deleterious effects of salt stress on seed germination and early seedling growth of wheat cultivars were considerably alleviated by melatonin priming. On one hand, the exogenous application of melatonin improved soluble sugar, sucrose contents, α - and β activities under salt stress. Also enhanced ABA content in germinated seeds. Conversely, melatonin could effectively scavenge ROS and improved antioxidant enzymes activities. These findings provide a novel use of melatonin in wheat during seed germination and seedling establishment under salt stress. We expect that the positive effect of melatonin in mitigation salt stress present new opportunities for its employ in agriculture. Highest values for most parameters measured were obtained at 500 μ M of melatonin level and it is used for our next experiment of wheat seedling performance under salt stress.

References

- [1] Wassmann, R., S. Jagadish, et al. (2009). "Climate change affecting rice production: the physiological and agronomic basis for possible adaptation strategies." Advances in Agronomy **101**: 59-122.
- [2]Ibrahim, H., M. Eldeen, et al. (2016). "Comparison of Germination and Seedling Characteristics of Wheat Varieties from China and Sudan under Salt Stress." Agronomy Journal **108**(1): 85-92.
- [3] Hubbard, M., J. Germida, et al. (2012). "Fungal endophytes improve wheat seed germination under heat and drought stress." <u>Botany</u> **90**(2): 137-149.
- [4]Hampson, C. R. and G. Simpson (1990). "Effects of temperature, salt, and osmotic potential on early growth of wheat (*Triticum aestivum* L.). I. Germination." Canadian Journal of Botany **68**(3): 524-528.
- [5]Fahad, S., S. Hussain, et al. (2015). "Phytohormones and plant responses to salinity stress: a review." Plant Growth Regulation **75**(2): 391-404.
- [6]Rouhi, H. R., M. A. Aboutalebian, et al. (2011). "Effects of hydro and osmopriming on drought stress tolerance during germination in four grass species." <u>International journal of Agrisience</u> **1**(2): 107-114.
- [7]Jahromi, F., R. Aroca, et al. (2008). "Influence of salinity on the in vitro development of Glomus intraradices and on the in vivo physiological and molecular responses of mycorrhizal lettuce plants." Microbial Ecology **55**(1): 45-53.
- [8]Daszkowska-Golec, A. (2011). "Arabidopsis seed germination under abiotic stress as a concert of action of phytohormones." Omics: a journal of integrative biology **15**(11): 763-774.

- [9]Panda, S. and M. Khan (2009). "Growth, Oxidative Damage and Antioxidant Responses in Greengram (*Vigna radiata* L.) under Short-term Salinity Stress and its Recovery." <u>Journal of Agronomy and Crop Science</u> **195**(6): 442-454.
- [10]Matraszek, R., B. Hawrylak-Nowak, et al. (2015). "Protein hydrolysate as a component of salinized soil in the cultivation of Ageratum houstonianum Mill.(Asteraceae)." Acta Agrobotanica **68**(3): 247-253.
- [11]Khan, M. A. and D. J. Weber (2006). <u>Ecophysiology of high salinity</u> tolerant plants, Springer Science & Business Media.
- [12]Gong, H., K. Chen, et al. (2008). "Effects of silicon on defense of wheat against oxidative stress under drought at different developmental stages." Biologia plantarum **52**(3): 592-596.
- [13]Gamir, J., M. Cerezo, et al. (2014). "The plasticity of priming phenomenon activates not only common metabolomic fingerprint but also specific responses against P. cucumerina." <u>Plant signaling</u> & behavior 9(5): 227-240.
- [14]Erdal, S. and A. Demirtas (2010). "Effects of cement flue dust from a cement factory on stress parameters and diversity of aquatic plants." Toxicology and industrial health 26(6): 339-343.
- [15]Genisel, M., H. Turk, et al. (2013). "Exogenous progesterone application protects chickpea seedlings against chilling-induced oxidative stress." Acta physiologiae plantarum **35**(1): 241-251.
- [16]Calvo, J. R., C. Gonzalez-Yanes, et al. (2013). "The role of melatonin in the cells of the innate immunity: a review." <u>Journal of pineal research</u> **55**(2): 103-120.

- [17]Arnao, M. B. and J. Hernández-Ruiz (2014). "Melatonin: plant growth regulator and/or biostimulator during stress?" <u>Trends in plant science</u> **19**(12): 789-797.
- [18]Zhang, N., B. Zhao, et al. (2013). "Melatonin promotes water-stress tolerance, lateral root formation, and seed germination in cucumber (*Cucumis sativus* L.)." Journal of pineal research **54**(1): 15-23.
- [19]Bajwa, V. S., M. R. Shukla, et al. (2014). "Role of melatonin in alleviating cold stress in Arabidopsis thaliana." <u>Journal of pineal research</u> **56**(3): 238-245.
- [20]Zhao, Y., L. W. Qi, et al. (2011). "Melatonin improves the survival of cryopreserved callus of Rhodiola crenulata." <u>Journal of pineal</u> research **50**(1): 83-88.
- [21]Szafrańska, K., S. Glińska, et al. (2012). "Changes in the nature of phenolic deposits after re-warming as a result of melatonin presowing treatment of *Vigna radiata* seeds." <u>Journal of plant physiology</u> **169**(1): 34-40
- [22]Zheng, C., D. Jiang, et al. (2009). "Effects of salt and waterlogging stresses and their combination on leaf photosynthesis, chloroplast ATP synthesis, and antioxidant capacity in wheat." Plant Science 176(4): 575-582.
- [23]Wang, Y., L. Yu, et al. (2004). "Vigor tests used to rank seed lot quality and predict field emergence in four forage species." <u>Crop Science</u> **44**(2): 535-541.
- [24]Xie, Z., D. Jiang, et al. (2003). "Relationships of endogenous plant hormones to accumulation of grain protein and starch in winter wheat under different post-anthesis soil water statusses." <u>Plant Growth Regulation</u> **41**(2): 117-127.

- [25]Fales, F. W. (1951). "The assimilation and degradation of carbohydrates by yeast cells." <u>Journal of Biological Chemistry</u> **193**(1): 113-124.
- [26]Kishorekumar, A., C. A. Jaleel, et al. (2007). "Comparative effects of different triazole compounds on growth, photosynthetic pigments and carbohydrate metabolism of Solenostemon rotundifolius." Colloids and Surfaces B: Biointerfaces **60**(2): 207-212.
- [27]Bradford, M. M. (1976). "A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding." <u>Analytical biochemistry</u> **72**(1-2): 248-254.
- [28]Tan, W., J. Liu, et al. (2008). "Alterations in photosynthesis and antioxidant enzyme activity in winter wheat subjected to post-anthesis water-logging." Photosynthetica **46**(1): 21-27.
- [29]Du, Z. and W. J. Bramlage (1992). "Modified thiobarbituric acid assay for measuring lipid oxidation in sugar-rich plant tissue extracts." <u>Journal of Agricultural and Food Chemistry</u> **40**(9): 1566-1570.
- [30]Moloi, M. J. and A. J. van der Westhuizen (2006). "The reactive oxygen species are involved in resistance responses of wheat to the Russian wheat aphid." <u>Journal of plant physiology</u> **163**(11): 1118-1125.
- [31]Yang, S.-H., L.-J. Wang, et al. (2007). "Ultraviolet-B irradiation-induced freezing tolerance in relation to antioxidant system in winter wheat (*Triticum aestivum* L.) leaves." Environmental and Experimental Botany **60**(3): 300-307.
- [32]Nakano, Y. and K. Asada (1981). "Hydrogen peroxide is scavenged by ascorbate-specific peroxidase in spinach chloroplasts." <u>Plant and cell physiology</u> **22**(5): 867-880.

- [33]You-Ming, Y., X. Chu-Nian, et al. (2001). "Effects of plant growth regulators on secondary wall thickening of cotton fibres." <u>Plant Growth Regulation</u> **35**(3): 233-237.
- [34] Genisel, M., S. Erdal, et al. (2015). "The mitigating effect of cysteine on growth inhibition in salt-stressed barley seeds is related to its own reducing capacity rather than its effects on antioxidant system." Plant Growth Regulation 75(1): 187-197.
- [35]Lee, H. Y., Y. Byeon, et al. (2014). "Melatonin as a signal molecule triggering defense responses against pathogen attack in Arabidopsis and tobacco." <u>Journal of pineal research</u> **57**(3): 262-268.
- [36]Erdal, S. (2012). "Alleviation of salt stress in wheat seedlings by mammalian sex hormones." <u>Journal of the Science of Food and</u> Agriculture **92**(7): 1411-1416.
- [37]Zhang, N., Q. Sun, et al. (2015). "Roles of melatonin in abiotic stress resistance in plants." <u>Journal of Experimental Botany</u> **66**(3): 647-656.
- [38]Zhang, H., W.-B. Shen, et al. (2005). "A rapid response of β-amylase to nitric oxide but not gibberellin in wheat seeds during the early stage of germination." <u>Planta</u> **220**(5): 708-716.
- [39]Cakmak, T., R. Dumlupinar, et al. (2010). "Chilling resistance of *Phaseolus vulgaris* and *Brassica oleracea* under a high-intensity electric field." Zeitschrift für Naturforschung C **65**(5-6): 380-386.
- [40]Petrussa, E., V. Casolo, et al. (2008). "Activity of a channel in Arum spadix mitochondria during thermogenesis." <u>Journal of plant physiology</u> **165**(13): 1360-1369.

- [41]Møller, I. M. (2001). "Plant mitochondria and oxidative stress: electron transport, NADPH turnover, and metabolism of reactive oxygen species." <u>Annual review of plant biology</u> **52**(1): 561-591.
- [42]Li, Q.-Y., H.-B. Niu, et al. (2008). "Protective role of exogenous nitric oxide against oxidative-stress induced by salt stress in barley (*Hordeum vulgare*)." Colloids and Surfaces B: Biointerfaces **65**(2): 220-225.
- [43]Ashraf, M. (2009). "Biotechnological approach of improving plant salt tolerance using antioxidants as markers." <u>Biotechnology</u> advances **27**(1): 84-93.
- [44]Song, L., W. Ding, et al. (2006). "Nitric oxide protects against oxidative stress under heat stress in the calluses from two ecotypes of reed." Plant Science **171**(4): 449-458.
- [45]Bethke, P. C., R. Schuurink, et al. (1997). "Hormonal signalling in cereal aleurone." <u>Journal of Experimental Botany</u> **48**(7): 1337-1356.
- [46]Larkindale, J. and M. R. Knight (2002). "Protection against heat stress-induced oxidative damage in Arabidopsis involves calcium, abscisic acid, ethylene, and salicylic acid." <u>Plant physiology</u> **128**(2): 682-695.
- [47]Ashraf, M. and M. Foolad (2005). "Pre-sowing seed treatment—A shotgun approach to improve germination, plant growth, and crop yield under saline and non-saline conditions." <u>Advances in Agronomy</u> 88: 223-271.
- [48]Wani, S. and V. Kumar (2015). "Plant stress tolerance: engineering ABA: a Potent phytohormone." <u>Transcriptomics</u> **3**: 113.

- [49]Ng, L. M., K. Melcher, et al. (2014). "Abscisic acid perception and signaling: structural mechanisms and applications." <u>Acta Pharmacologica Sinica</u> **35**(5): 567-584.
- [50]Hamayun, M., S. A. Khan, et al. (2010). "Effect of salt stress on growth attributes and endogenous growth hormones of soybean cultivar Hwangkeumkong." Pak. J. Bot 42(5): 3103-3112.
- [51]Xu, X.-y., R. Fan, et al. (2011). "Proteomic analysis of seed germination under salt stress in soybeans." <u>Journal of Zhejiang</u> <u>University-Science B</u> **12**(7): 507-517.
- [52]Parida, A. K. and A. B. Das (2005). "Salt tolerance and salinity effects on plants: a review." <u>Ecotoxicology and environmentsafety</u> **60**(3): 324-349.
- [53]Hernández-Ruiz, J. and M. B. Arnao (2008). "Melatonin stimulates the expansion of etiolated lupin cotyledons." <u>Plant growth regulation</u> **55**(1): 29-34.