

Correlation Between Transgingival Probing and CBCT Evaluation for Determination of Gingival Biotype

ABSTRACT

Aims: This study aimed to determine the correlation between transgingival probing and CBCT evaluation, for the determination of gingival biotype

Study design: Cross-sectional study

Place and Duration of Study: Department of Periodontics, Fatima Memorial Hospital, Lahore and 17th August 2016 to 16th February 2016

Methodology: In this cross-sectional study, a total of 40 patients indicated to undergo implant placement for posterior maxillary teeth or any mandibular teeth, 18 to 50 years were included. Patients with the presence of restoration in the anterior maxilla, pregnant or lactating women, root canal treatment in the anterior maxilla, and h/o apical surgery were excluded. A single radiographer took all CBCT from the SIRONA machine of all subjects. Linear measurements for buccal wall & gingival biotype were measured.

Results: The mean age was 35.13 ± 7.75 years. Out of 40 patients, 22 (55.0%) were females and 18 (45.0%) were males with a female to male ratio of 1.2:1. Radiographic measurements on CBCT were 1.49 ± 0.34 mm for right central and 1.49 ± 0.34 mm for left central. Correlation between transgingival probing and CBCT evaluation, for determination of gingival biotype with Spearman's correlation coefficient of 0.985 and p-value = 0.0001 which is statistically significant.

Conclusion: This study concluded that there is a significant positive correlation between transgingival probing and CBCT measurements of gingival biotypes.

Keywords: Gingival Biotype, Transgingival Probing, Cone Beam Computed Tomography

1. INTRODUCTION

Gingival biotype is the term used to describe the thickness of the gingiva in the faciopalatal dimension [1]. Commonly, it is categorized as thin scalloped, thick flat, and thick scalloped [2]. Gingival biotype is considered "thin" if it is equal to or lesser than 1.5 mm and it is considered "thick" if it is equal to or greater than 2 mm [3]. Gingival dimensions, like width and thickness, show great intra and inter-individual variation, which are associated with tooth type and shape, and are certainly also genetically determined [4]. Recently, distinct gingival phenotypes have been identified on a subject level, and their existence was later confirmed in an independent, periodontally healthy population of young adults by using cluster analysis [5]. Individuals with a thin phenotype had slightly more recession than subjects with wide and thick gingival tissues [5].

Among the factors that may affect the prognosis of dental treatments, gingival biotype is a critical cause of concern. It may affect the outcomes of periodontal therapy, root coverage

procedures, and implant placement. Different tissue biotypes respond differently to inflammation and to surgical and restorative treatment; therefore, it is crucial to identify tissue biotypes prior to treatment planning [6].

Gingival thickness can be assessed by various invasive and non-invasive methods which include the direct method, transgingival probing method, ultrasound-guided methods, and, more recently, cone-beam computed tomography (CBCT). Periodontal probing-assessed gingival biotype is a simple, relatively objective, and suitable method for clinical examination. Goasind et al used a digital voltmeter and described 2 types of gingival biotypes commonly found in the natural dentition that is thick and thin [7].

Becker et.al proposed three different periodontal morphotypes: flat, scalloped, and pronounced scalloped gingiva. Measuring from the height of the bone interproximally to the height midfacially, findings were as follows: flat= 2.1 mm, scalloped= 2.8 mm, pronounced scalloped= 4.1 mm [8].

This study was being conducted to see a correlation between transgingival probing and CBCT as no local study was found in the literature search. The gingival biotype in the local Pakistani population is different from Caucasian and Chinese Asians. Mean values formulated by the above-mentioned authors may not serve as references for the Pakistani population. Each population should be treated according to specific characteristics of its own. It is thus important to establish the gingival biotype in the local population to provide predictable restorative and surgical treatment results. This study aimed to determine the correlation between transgingival probing and CBCT evaluation, for the determination of gingival biotype.

2. MATERIAL AND METHODS

2.1 Study design and Sample size

This descriptive, cross-sectional study was carried out between 17th August 2016 to 16th February 2016. at the Department of Periodontics, Fatima Memorial Hospital, Lahore. Fatima Memorial Hospital is a tertiary care hospital affiliated with the University of Health Sciences. This study has been carried out in accordance with the Declaration of Helsinki. For this study, the participants were recruited using a non-probability, consecutive sampling method. The process of data collection was started after being granted ethical approval. For calculation of sample size, OpenEpi software was used. The sample size of 61 cases was calculated with 5% type-I error, and 10% type-II error, and taking the expected correlation coefficient between clinical method (TP) and radiographic method (CBCT) for diagnosis of gingival biotype i.e., $r=0.401$. Since practically it's not possible for us to collect the data of 61 implant cases in a 6-months duration so we had taken the sample of 40 cases.

2.2 Inclusion and Exclusion Criteria

Participation in this study was based on pre-determined inclusion and exclusion criteria. The participants were included in this study on the basis of the following criteria:

- Age 18 to 50 years.
- Periodontally healthy individuals.
- The patient indicated to undergo implant placement for posterior maxillary teeth or any mandibular teeth as dictated by his/her treatment plan.
- No history of chemotherapy and radiotherapy.
- No history of diabetes, or any medications such as bisphosphonates, or drugs/conditions causing gingival enlargement.

The participants were excluded from this study on the basis of the following factors:

- Pregnant or lactating mothers
- Pathological migration of teeth, malalignment of teeth
- Presence of soft tissue recession

- Smokers
- Presence of restoration in the anterior maxilla
- Root canal treatment in the anterior maxilla
- Any history of apical surgery
- Any history of orthodontic treatment

2.3 Data Collection

The participants who fulfilled the selection criteria from the dental outpatient department of Periodontics Fatima Memorial Hospital were selected. Approval from the institutional review board (IRB) of Fatima memorial hospital was taken. A consent form was signed by every patient. The demographic profile of all the patients was recorded, history of past dental condition was explored and a thorough dental checkup was carried out. A single radiographer took all CBCT from the SIRONA machine of all subjects. As per operational definitions, linear measurements for buccal wall & gingival biotype were measured. All the information was recorded in a specifically designed Performa (Annexure-II)

2.4 Statistical Analysis

After the collection of the data, it was analyzed using SPSS version 20. Quantitative variables like age and radiographic measurements on CBCT were presented in the form of mean and standard deviation. Qualitative data like gender, visual inspection of the clinical method, and on radiographic (i.e., $\geq 1.5\text{mm}$) were presented in the form of frequency and percentages. Spearman's correlation coefficient was calculated to determine the relationship between the clinical method (Transgingival Probing) and radiographic method (CBCT) for the diagnosis of gingival biotype. P-value ≤ 0.05 was considered as significant. Effect modifiers like age and gender were controlled by stratification. Post-stratification Spearman's correlation coefficient was calculated to see the effect of these on the outcome and a p-value ≤ 0.05 was taken as significant.

3. RESULTS

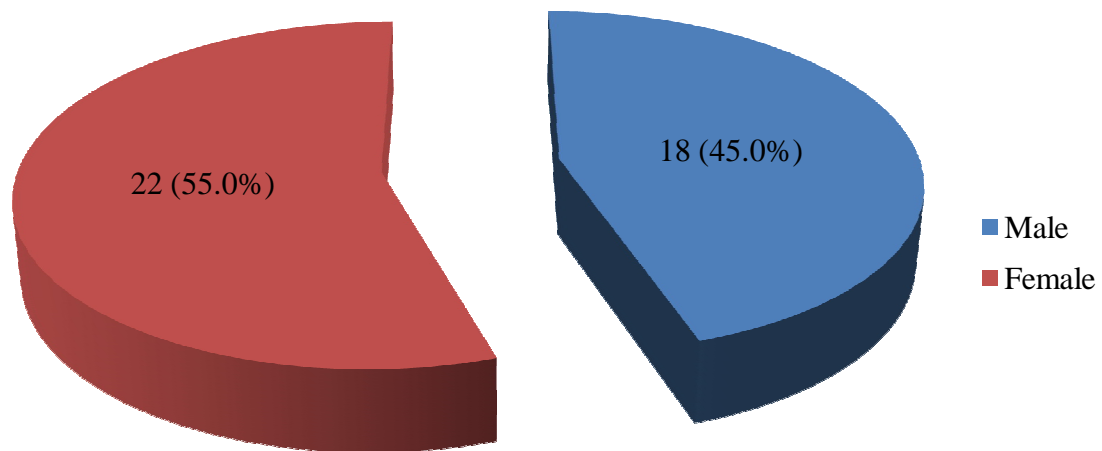
The age range in this study was from 18 to 50 years with a mean age of 35.13 ± 7.75 years. The majority of the patients 28 (70.0%) were between 18 to 40 years of age as shown in Table II

Table 1. Age distribution for both groups (n=40).

Age (in years)	No. of Patients	Percentage
18-30	14	35.0
31-40	14	35.0
41-50	12	30.0

Out of 40 patients, 22 (55.0%) were females and 18 (45.0%) were males with a female to male ratio of 1.2:1 (Figure 1).

Figure 1: Distribution of patients according to gender



Radiographic measurements on CBCT were 1.49 ± 0.34 mm for right central and 1.49 ± 0.34 mm for left central (Table 2). Gingival biotype for right central and left central on visual inspection on clinical method and on radiographic are shown in Table 3 & 4 respectively.

Table 2. Radiographic measurements on CBCT (n=40).

	Minimum	Maximum	Mean	SD
CBCT (Right Central)	0.97	2.13	1.49	0.34
CBCT (Left Central)	0.98	2.10	1.49	0.32

Table 3. Gingival biotype for right central on visual inspection on clinical method and on radiographic (n=40).

	Thin	Thick
Gingival biotype on visual inspection	19 (47.50%)	21 (52.50%)
Gingival biotype on visual inspection on radiographic	23 (57.50%)	17 (42.50%)

Table 4. Gingival biotype for left central on visual inspection on clinical method and on radiographic (n=40).

	Thin	Thick
Gingival biotype on visual inspection	19 (47.50%)	21 (52.50%)
Gingival biotype on visual inspection on radiographic	23 (57.50%)	17 (42.50%)

The correlation between transgingival probing and CBCT evaluation, for the determination of gingival biotype, is shown in Table 5 with Spearman's correlation coefficient of 0.985 and p-value = 0.0001 which is statistically significant.

Table 5. Spearman's correlation coefficient was calculated to determine the relationship between clinical method (Transgingival Probing) and radiographic method (CBCT) for the diagnosis of gingival biotype

Correlations				
			TP	CBCT
Spearman's rho	Transgingival Probing	Correlation Coefficient	1.000	0.985**
		Sig. (1-tailed)	.	0.000
		N	40	40
	CBCT	Correlation Coefficient	0.985**	1.000
		Sig. (1-tailed)	0.000	.
		N	40	40

** . Correlation is significant at the 0.01 level (1-tailed).

Stratification of age and Spearman's correlation coefficient to determine the relationship between clinical method (Transgingival Probing) and radiographic method (CBCT) for the diagnosis of gingival biotype is shown in Table 6. Stratification of gender and Spearman's correlation coefficient to determine the relationship between clinical method (Transgingival Probing) and radiographic method (CBCT) for the diagnosis of gingival biotype is shown in Figure 2.

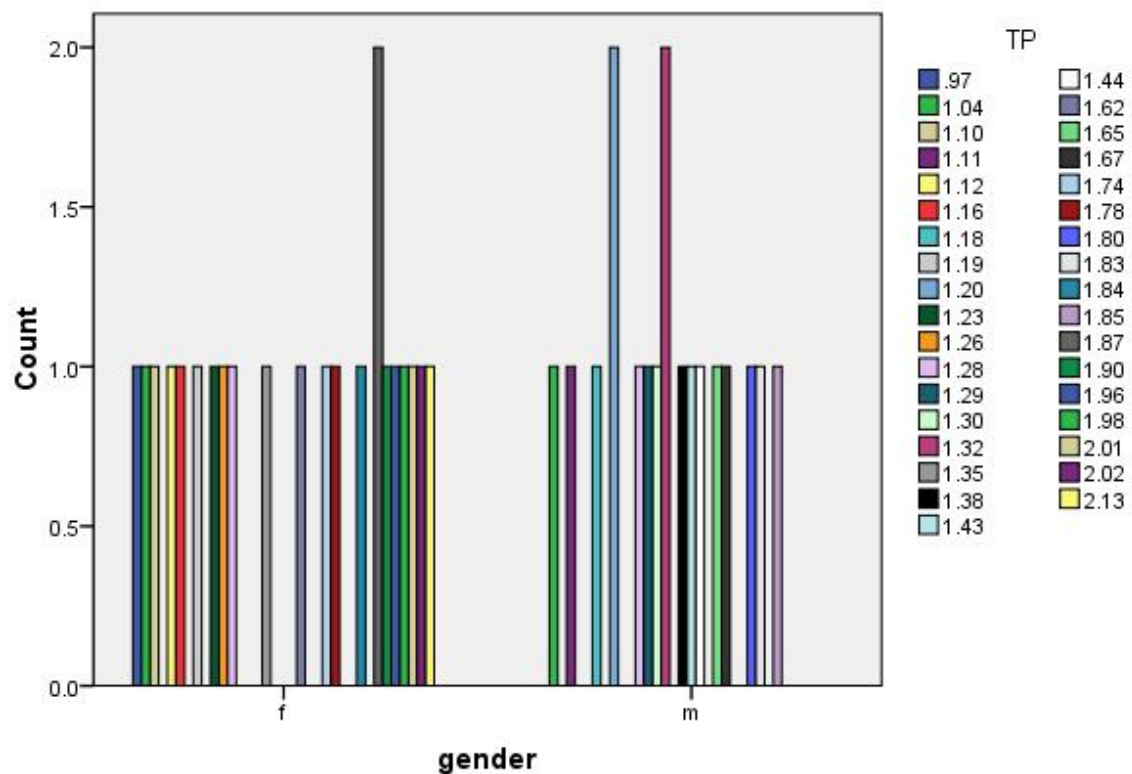
Table 6. Stratification of age and Spearman's correlation coefficient to determine the relationship between clinical method (Transgingival Probing) and radiographic method (CBCT) for the diagnosis of gingival biotype

Correlations			
	Age	TP	CBCT

Spearman's rho	age	Correlation Coefficient	1.000	.077	.073
		Sig. (1-tailed)	.	0.319	0.327
		N	40	40	40
	TP	Correlation Coefficient	0.077	1.000	0.985**
		Sig. (1-tailed)	0.319	.	0.000
		N	40	40	40
CBCT		Correlation Coefficient	0.073	0.985**	1.000
		Sig. (1-tailed)	0.327	0.000	.
		N	40	40	40

** . Correlation is significant at the 0.01 level (1-tailed).

Figure 2. Stratification of gender and Spearman's correlation coefficient to determine the relationship between clinical method (Transgingival Probing) and radiographic method (CBCT) for the diagnosis of gingival biotype



4. DISCUSSION

Gingival biotype can be evaluated either by direct visual assessment, by using a periodontal probe, or by direct measurements using endodontic spreaders, endodontic files, and calipers. If the terms “thick” and “thin” are focused upon, only the buccopalatal measurement of gingival thickness is worth evaluating for clinical and research purposes. Various invasive and non-invasive methods were proposed to measure tissue thickness. These include direct measurement, probe transparency (TRAN) method, ultrasonic devices, and cone-beam computed tomography (CBCT) scan [9][10][11].

The use of ultrasonic devices to determine thickness is a non-invasive method that has been proved to be reproducible [12],¹³¹ drawbacks include difficulties in maintaining the directionality of the transducer [13] and unavailability of the device [14] and high costs. These factors may be responsible for the fact that the device has not become part of the standard armamentarium of the clinician. A simpler method has been proposed to discriminate thin from thick gingiva based on the transparency of the periodontal probe through the gingival margin [15].

Recently cone-beam computed tomography scan (CBCT) is being used as an advanced diagnostic aid in measuring the thickness of hard as well as soft tissues [11]. Fu et al. stated that CBCT provides accurate measurements of both bone and labial soft tissue thickness. He concluded that CBCT measurements might be a more objective method to define the thickness of both soft and hard tissues than direct measurements [16]. Although several studies have previously investigated the thickness of palatal mucosa by transgingival probing, only a few reported the thickness of facial gingiva using the soft tissue CBCT method, the present study was undertaken to evaluate the association between soft tissue thickness of mandibular anteriors and underlying bone using transgingival probing and soft tissue CBCT.

The age range in my study was from 18 to 50 years with a mean age of 35.13 ± 7.75 years. The majority of the patients 28 (70.0%) were between 18 to 40 years of age. Out of 40 patients, 22 (55.0%) were females and 18 (45.0%) were males with a female to male ratio of 1.2:1. Correlation between transgingival probing and CBCT evaluation, for determination of gingival biotype with Spearman's correlation coefficient of 0.985 and p-value = 0.0001 which is statistically significant. Beijing Da Bao et al observed a significant positive correlation between transgingival probing and CBCT measurements of gingival biotypes with an R-value of 0.401.10.

Until now, there is no precise definition of how a thick biotype can be compared to a thin one. One of the reasons may be seen in the fact that the thickness of the gingiva has been assessed at different vertical levels. Earlier, invasive methods were used to determine the gingival thickness; direct measurement [17] was used but had various limitations i.e., invasive approach, lack of reproducibility, accuracy, improper angulation, and pressure. To overcome these limitations, non-invasive methods were devised; ultrasonic devices [18] and cone-beam computed tomography [19] but these methods are technique sensitive and quite expensive. Manual assessment using a caliper after tooth extraction [20], a syringe with an endodontic depth marker or cone beam radiographs without reference objects have limitations of their accuracy. The most recent technique devised is a modified radiographic technique [21] described by Alpiste-Illueca [22], which determined that different morphometric parameters such as crown width/crown length ratio and gingival width could represent surrogate parameters to anticipate the gingival thickness at the cemento-enamel junction.

Kan et al. [20] presented a simple method of periodontal type determination, which utilizes translucency of the free gingiva during the probing of gingival grooves in teeth. Visual

inspection of the transparency of the periodontal probe through the sulcus has become the most frequently used method for the discrimination of thin and thick biotypes. The gingival biotype is considered thin if the outline of the probe is shown through the gingival margin from the sulcus. The gingival tissue's ability to cover any underlying material's color is necessary for achieving esthetic results, especially in cases of implant and restorative dentistry, for this purpose subgingival alloys are widely used. Using a metal periodontal probe in the sulcus to evaluate gingival tissue thickness is the simplest way to determine the thin gingival biotype, the tip of the probe is visible through the gingiva [23]. This method is minimally invasive, and periodontal probing procedures are performed routinely during periodontal and implant treatments.

CBCT is used to visualize and measure the thickness of both hard and soft tissues. Various authors reported that CBCT measurements of both bone and labial soft tissue thickness are accurate and concluded that CBCT measurements might be a more objective method to determine the thickness of both soft and hard tissues than direct measurements. In contrast to transgingival probing and the ultrasonic device, the CBCT method provides an image of the tooth, gingiva, and other periodontal structures. Moreover, measurements can be repeatedly taken at different times with the same image obtained by ST-CBCT (soft tissue CBCT) which is not feasible by other methods [24].

Stein et al [25] performed a comparative study of 60 subjects and reported a positive correlation between buccal bone thickness and gingival thickness. However, the comparison in their study was not carried out at an identical level. Instead, the gingival thickness was evaluated at the supracrestal level, while bone thickness was measured under the alveolar crest. In contrast, in an in vivo study of 90 maxillary teeth, La Rocca et al [26] observed no significant correlation between the results of CBCT scans and transgingival probing, although the comparison in their study was also not performed at an identical level. Considering these conflicting results, and despite the limited sample size of our study, we observed a significant positive correlation between transgingival probing and CBCT measurements of gingival biotypes.

4. CONCLUSION

This study concluded that there is a significant positive correlation between transgingival probing and CBCT measurements of gingival biotypes. So, we recommend that CBCT is a beneficial method for measuring both hard and soft tissue thickness and gingival biotype should be established in every periodontal disease patient in order to provide predictable restorative and surgical treatment results.

CONSENT (WHERE EVER APPLICABLE)

All authors declare that 'written informed and verbal consent was obtained from the patients.

ETHICAL APPROVAL (WHERE EVER APPLICABLE)

All authors hereby declare that all experiments have been examined and approved by the appropriate ethics committee and have therefore been performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki."

REFERENCES

1. Manjunath RGS, Rana A, Sarkar A. Gingival Biotype Assessment in a Healthy Periodontium: Transgingival Probing Method. *J Clin Diagn Res*. 2015 May;9(5):ZC66-9.
2. Zweers J, Thomas RZ, Slot DE, Weisgold AS, Van der Weijden FGA. Characteristics of periodontal biotype, its dimensions, associations and prevalence: a systematic review. *J Clin Periodontol* [Internet]. 2014 Oct;41(10):958–71. Available from: <https://onlinelibrary.wiley.com/doi/10.1111/jcpe.12275>
3. Claffey N, Shanley D. Relationship of gingival thickness and bleeding to loss of probing attachment in shallow sites following nonsurgical periodontal therapy. *J Clin Periodontol* [Internet]. 1986 Aug;13(7):654–7. Available from: <https://onlinelibrary.wiley.com/doi/10.1111/j.1600-051X.1986.tb00861.x>
4. Schroeder HE, Schroeder H, hc Oksche A, Vollrath L. The Periodontium With Technical Assistance of Margrit Amstad-Jossi Rosmarie Kroni Walter Scherle. 1986;233–46.
5. Müller HP, Heinecke A, Schaller N, Eger T. Masticatory mucosa in subjects with different periodontal phenotypes. *J Clin Periodontol*. 2000 Sep;27(9):621–6.
6. Esfahrood ZR, Kadkhodazadeh M, Ardakani MRT. Gingival biotype: A review. *Gen Dent*. 2013;61(4):14–7.
7. Abraham S, Deepak KT, Ranjith A, Preeja C, Archana V. Gingival biotype and its clinical significance – A review. *King Saud Univ J Dent Sci*. 2013 Jan 1;5.
8. Cook DR, Mealey BL, Verrett RG, Mills MP, Noujeim ME, Lasho DJ, et al. Relationship between clinical periodontal biotype and labial plate thickness: an in vivo study. *Int J Periodontics Restorative Dent* [Internet]. 31(4):345–54. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/21837300>
9. Terakura T. [Non-invasive measurement of the thickness of oral soft tissues]. *Nihon Hotetsu Shika Gakkai Zasshi*. 1986 Dec;30(6):1402–11.
10. Lawson RB, Jones ML. An evaluation of a noninvasive method of assessing alveolar bone levels in an experimental model of cleft lip and palate. *Cleft palate-craniofacial J Off Publ Am Cleft Palate-Craniofacial Assoc*. 1998 Jan;35(1):1–8.
11. Barriviera M, Duarte WR, Januário AL, Faber J, Bezerra ACB. A new method to assess and measure palatal masticatory mucosa by cone-beam computerized tomography. *J Clin Periodontol*. 2009 Jul;36(7):564–8.
12. Eger T, Müller HP, Heinecke A. Ultrasonic determination of gingival thickness. Subject variation and influence of tooth type and clinical features. *J Clin Periodontol*. 1996 Sep;23(9):839–45.
13. Daly CH, Wheeler JB 3rd. The use of ultra-sonic thickness measurement in the clinical evaluation of the oral soft tissues. *Int Dent J*. 1971 Dec;21(4):418–29.
14. Vandana KL, Savitha B. Thickness of gingiva in association with age, gender and dental arch location. *J Clin Periodontol* [Internet]. 2005 Jul;32(7):828–30. Available from: <https://onlinelibrary.wiley.com/doi/10.1111/j.1600-051X.2005.00757.x>

15. Kan JYK, Rungcharassaeng K, Umez K, Kois JC. Dimensions of peri-implant mucosa: an evaluation of maxillary anterior single implants in humans. *J Periodontol*. 2003 Apr;74(4):557–62.
16. Fu J-H, Yeh C-Y, Chan H-L, Tatarakis N, Leong DJM, Wang H-L. Tissue biotype and its relation to the underlying bone morphology. *J Periodontol*. 2010 Apr;81(4):569–74.
17. Rogers MJ, Gordon S, Benford HL, Coxon FP, Luckman SP, Monkkonen J, et al. Cellular and molecular mechanisms of action of bisphosphonates. *Cancer*. 2000 Jun;88(12 Suppl):2961–78.
18. Vitté C, Fleisch H, Guenther HL. Bisphosphonates induce osteoblasts to secrete an inhibitor of osteoclast-mediated resorption. *Endocrinology*. 1996 Jun;137(6):2324–33.
19. Lezcano V, Bellido T, Plotkin LI, Boland R, Morelli S. Osteoblastic protein tyrosine phosphatases inhibition and connexin 43 phosphorylation by alendronate. *Exp Cell Res [Internet]*. 2014 May;324(1):30–9. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S0014482714001323>
20. Duque G, Rivas D. Alendronate has an anabolic effect on bone through the differentiation of mesenchymal stem cells. *J bone Miner Res Off J Am Soc Bone Miner Res*. 2007 Oct;22(10):1603–11.
21. Pan B, Farrugia AN, To LB, Findlay DM, Green J, Lynch K, et al. The nitrogen-containing bisphosphonate, zoledronic acid, influences RANKL expression in human osteoblast-like cells by activating TNF-alpha converting enzyme (TACE). *J bone Miner Res Off J Am Soc Bone Miner Res*. 2004 Jan;19(1):147–54.
22. Pietschmann P, Stohlawetz P, Brosch S, Steiner G, Smolen JS, Peterlik M. The effect of alendronate on cytokine production, adhesion molecule expression, and transendothelial migration of human peripheral blood mononuclear cells. *Calcif Tissue Int*. 1998 Oct;63(4):325–30.
23. Price U, Le H-OT, Powell SE, Schmid MJ, Marx DB, Zhang Y, et al. Effects of local simvastatin-alendronate conjugate in preventing periodontitis bone loss. *J Periodontal Res*. 2013 Oct;48(5):541–8.
24. Shibutani T, Inuduka A, Horiki I, Luan Q, Iwayama Y. Bisphosphonate inhibits alveolar bone resorption in experimentally-induced peri-implantitis in dogs. *Clin Oral Implants Res*. 2001 Apr;12(2):109–14.
25. Stein JM, Lintel-Höping N, Hammächer C, Kasaj A, Tamm M, Hanisch O. The gingival biotype: measurement of soft and hard tissue dimensions - a radiographic morphometric study. *J Clin Periodontol*. 2013 Dec;40(12):1132–9.
26. La Rocca AP, Alemany AS, Levi PJ, Juan MV, Molina JN, Weisgold AS. Anterior maxillary and mandibular biotype: relationship between gingival thickness and width with respect to underlying bone thickness. *Implant Dent*. 2012 Dec;21(6):507–15.